

conditions and has ability to grow in the alkaline, acidic and saline soil [6, 7]. In Pakistan, the average production is about 4.9 tons/ha, that makes the country 11th biggest rice producer in the world [8]. Various rice varieties in Pakistan are super kernel, basmati, kinaat and super Shaheen etc.

AFTs are common mycotoxin which affects the crops especially staple food (wheat, rice, maize and grains). High temperature and humidity are favorable conditions for fungus growth [9]. International Agency for Research on Cancer (IARC) ranked some materials as class one carcinogen in which AFTs also involved. *Aspergillus flavus*, *Aspergillus parasiticus*, *Aspergillus nomius* and *Aspergillus pseudotamarii* are the fungi that are responsible for the production of AFTs [10-12]. Different food commodities that have great chances of contaminations from AFTs include maize, corn, barely, silage, milk and hay etc. Mycotoxins cause the serious complications for animal and human health. These are contaminants of vital plant products like oil seeds, nuts, spices, corn, rice, barley and wheat [13]. Mycotoxins maximum levels are fixed by specific countries for different types of food products. AFTs are significant mycotoxins for foodstuff contaminants worldwide, especially for the developing countries. The important types of the AFTs are AFG1, AFG2, AFB1, AFB2, AFM1 and AFM2. AFB1 is the vital toxic variety as compared to other AFTs and it is the most persuasive human carcinogen. AFB1 type is categorized into the groups of carcinogenic compounds through the IARC of World Health Organization (WHO) [8]. AFB1 remain stable during processing operations. These contaminations may lead to huge economic damages in various countries. Different countries have specific regulations to control the mycotoxin levels in plant and dairy products [13]. European Unions have established the maximum tolerable limits for the AFTs in rice products as 4 ppb for the total AFTs (G1 + G2 + B1 + B2) and 2 ppb for AFB1 (Commission Regulation No. 1881/2006). Pakistan standard quality control authority has made a limit of 5 ppb for the AFB1 in rice products. Various methods can be used for the analysis of AFTs including the high performance liquid chromatography (HPLC), thin layer chromatography (TLC) and ELISA [14]. ELISA is the simplest method which has advantages as being selective, rapid, monotonous reliable and diagnostic analysis of various samples [13].

The purpose of this research was to find out the existence of AFTs in Pakistani rice and cattle feed and to determine the conditions responsible and ultimately to control AFTs production. It would be helpful to reduce the chances of hepatocellular carcinoma, immune super sensitivity and encephalopathy etc.

Material and Methods

Rice & cattle feed samples were collected different regions of Punjab, Pakistan (Table 1). All types of

samples were changed into powdered form using Molineux blender. 500 g of each sample was stored at 4-6°C in the sealed plastic bags for analysis [15].

Thin Layer Chromatography Technique (TLC)

The dilutions were made for spotting using micro syringe. Spot of 25 µL of test solution was applied on TLC plate. Standard spots of 5 and 10 µL of the AFTs (AFG1, AFG2, AFB1, and AFB2) were also spotted on the plate as an internal standard. The plate of TLC was placed in TLC Tank 1 (1st mobile phase) containing the anhydrous ether until the solvent moved half way up. After this, TLC plate was developed and it was dried. The redevelopment of the plate was done in similar direction but along with solution of chloroform and acetone to a ratio of 9:1 in the TLC Tank 2. After the exclusion of TLC plate from TLC Tank 2 (2nd mobile phase), spots were dehydrated and observed under the UV Light for the absence or presence of AFTs [16].

ELISA Analysis of AFB1

The quantitative examination of AFB1 in the samples was done, based on the competitive enzyme immunoassay via Ridascreen® Aflatoxin B₁ 30/15 test kit (R-Biopharma, Darmstadt, Germany). 25 mL methanol: water (70:30) was added to the 5 g sample and shaken robustly for 3 min. The extract was filtered and diluted with distilled water (1:1). At the end, 50 mL of the diluted remainders per well were used in the test. 50 mL of each prepared sample or standard solution in duplicate, antibody solution and enzyme conjugate were added to each micro titer well, mixed and incubated for 30 min at 25°C. The liquid was eliminated from the well through tapping of wells and were washed twice using buffer. The 100 mL of the substrate or chromogen solution was added to each micro titer well, mixed and incubated for 15 min at 25°C in dark. Finally, the 100 mL of stock solution (1 N H₂SO₄) was added to each micro titer well. The absorbance was measured at 450 nm.

Detoxification

Process of detoxification for the reducing type AFB1 was done on the 50 g of sample products that are naturally contaminated. For more accurate study, 100 g of uncontaminated samples were spiked for contamination. After that, both types of the samples were mixed with each other to increase the homogeneity of AFTs and were examined by TLC. Samples were detoxified through *Allium sativum* treatment [17], *Nigella sativa* [18] oil treatment, citric acid treatment [19] and sodium bicarbonate treatment [20]. These treatments were performed at 26°C for 30 min in the fuming hood.

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