

Original Research

***JrbHLH* Gene Family: Genome-Wide Identification and Transcriptional Expression in Persian Walnut (*Juglans regia* L.)**

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Abstract

In plants, the basic helix-loop-helix (*bHLH*) transcription factors play key roles in diverse biological processes. Genome-wide comprehensive and systematic analyses of *bHLH* proteins have been well conducted in Arabidopsis (*AtbHLH*), rice (*OsbHLH*) and tomato (*SlbHLH*) species. However, there are no reports of *bHLH* family genes have been functionally characterized in Persian walnut (*Juglans regia*). We identified a total of 189 *bHLH* genes in *J. regia*, which are clustering into 18 subfamilies. All *JrbHLH* contained the conserved domains *HLH*. A total of 69 *JrbHLH* genes were expanded as whole genome duplications (WGD). The syntenic analysis was conducted for Persian walnut (*J. regia*) with other two woody plants (*P. trichocarpa* and *O. europaea*) to infer the *bHLH* genes' evolutionary relationship between these species. We identified a total of 42 pairs of orthologous *bHLH* genes between *J. regia* and *P. trichocarpa*, while only 29 orthologous gene pairs between *J. regia* and *O. europaea*, while 25 collinear gene pairs were found in three species *J. regia*, *O. europaea* and *P. trichocarpa*. Furthermore, *JrbHLHs* had different expressed patterns between reproduction and vegetative tissues based on the transcriptome expression profiles of female flowers, male flowers, leaves, and hulls, respectively. Specifically, both transcriptome data and quantitative polymerase chain reaction (qRT-PCR) assessment showed that two *JrbHLH* genes (*JrbHLH3-5* and *JrbHLH13-13*) were highly represented in female and male flowers, while one *JrbHLH* gene (*JrbHLH13-4*) was highly expressed in leaves. Genome-wide identification, gene structure, phylogeny and expression analysis of the *JrbHLH*

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gene in *J.regia*, supplied a theoretical foundation for the functional study of the *JrbHLH* gene family and provide an understanding of the further facilitated molecular studies of Persian walnuts.

Keywords: phylogenetic analysis, transcription factor, DNA binding with one finger, *Juglans regia*, expression profile

Introduction

The basic helix-loop-helix (*bHLH*) gene, its transcriptional factors were first observed in mammals and subsequently detected in eukaryotic species [1-3]. In many species, members of the *bHLH* gene family play a role in eukaryotic organisms with transcriptional regulators [4, 5]. The *bHLH* superfamily is the second-largest transcription factor (TF) family, defined by a highly conserved *bHLH* domain with a length of approximately 50-60 amino acids and divided into two separate areas, the critical region and the *HLH* region [2, 5]. The base area consists about 10-15 amino acids for DNA binding located at N-terminus domain and C-terminal site involving of two amphipathic α -helices acting as a dimerization domain [6-8]. Concerning the highly conserved *bHLH* domain, there are also other preserved motifs in some *bHLH* superfamily [9, 10].

Genome-wide comprehensive and systematic analyses of *bHLH* proteins in plants have been carried out in many annual plant species, such as rice (*Oryza sativa*), maize (*Zea mays*), thaliana (*Arabidopsis thaliana*), and tomatoes (*Solanum lycopersicum*) [11, 12], including fruitful plants species such as apple (*Malus domestica*) and poplar (*Populus trichocarpa*) [2, 13]. However, the *bHLH* genes have not been functionally reported in Persian walnut (*Juglans regia*) previously. The function of *JrbHLH* proteins in plants is also involved in biological and biochemical processes, including the development of petals and the reaction to drought and salt stress and biosynthesis [13-15]. Besides, *bHLH* proteins are involved in the regulation of fruit growth and anthocyanin biosynthesis in flowers and fruit [16, 17].

Due to the importance of the *bHLH* genes in various physiological programs, it would be of interest to make a systematic investigation of the *bHLH* family in Persian walnut. The walnut genome sequencing provided an opportunity to reveal the organization, expression and evolutionary traits of walnut *bHLH* gene family at the genome-wide level [18]. In the present study, we identified 189 walnut *bHLH* genes and classified them into 18 main groups. The comprehensive analysis including the exon-intron organization, motif compositions, gene duplications, phylogenetic and synteny analysis were further investigated. Expression and quantitative polymerase chain response (qRT-PCR) analysis was performed to identify involvement of specific *bHLH* gene family members in flowering process in walnut. This study provided valuable clues for functional characterization of *bHLH* gene family members in walnut.

Material and Methods

Identification of bHLH Transcription Factors in *J. regia*

The whole protein sequence of walnuts downloaded from the NCBI Genbank (<https://treegenesdb.org/FTP/Genomes/Jure/v1.4/annotation/>). The members of *Arabidopsis JrbHLH* family were downloaded from the TAIR website (<https://www.arabidopsis.org/index.jsp>). We used *Arabidopsis bHLH* protein sequences as a query using a local alignment search tool (BLAST) to search against walnut protein sequences, considering those with an E-value of less than 1×10^{-10} . Therefore, we implemented the Markov hidden profile model (HMM) in HMMER v.3.2.1 for window, with default parameters for the search of *bHLH* proteins and domains in the protein family (Pfam) database [19].

Phylogenetic, Domain Analysis, Motif, Gene Structure Analysis of *JrbHLH* Transcription Factors

The neighbor-joining (NJ) tree of 189 *bHLH* transcription factors were constructed using MEGA v.7.0 software [16, 20], with the pairwise deletion of 1,000 bootstraps and a Poisson model, in the presence of potential domains using the Pfam webserver [21]. The same domains were also obtained from Pfam with an E-value cutoff of 1.0 to validate the result using the simple modular architecture research tool (SMART) program through BLASTN, the whole genome sequence of *J. regia* were examined *JrbHLH* transcription factors. Similarly, from the NCBI the conserved domain database (CDD) [22], and the whole CDS database was downloaded (<https://treegenesdb.org/FTP/Genomes/Jure/v1.4/annotation/>). The exon and intron structure were displayed using the online gene structure display server (<http://gsds.cbi.pku.edu.cn/>) [11]. The genome explorer (<https://www.ncbi.nlm.nih.gov/genome/>) was used to report the specific sequences of walnut genes and the motif identification was done by MEME software with default parameters, the maximum number of motifs was (20) and the optimum motifs were (30-50) [23].

Syntenic Analysis

To identify the potential homologous gene pairs ($E < 1 \times 10^{-5}$, top 5 matches) across multiple genomes using BLASTP. The identification of syntenic chains was determined using homologous gene pairs through MCScanX and detected the duplicate pairs of two

woody plants (*P. trichocarpa* and *O. europaea*) [24]. These detected duplicate gene pairs further analyzed through MCScanX including whole-genome duplication (WGD)/segmental and tandem duplication of walnut genes pair with *P. trichocarpa* and *O. europaea* species gene pairs.

GO Annotation

The Blast2GO v2.5 with a cutoff E-value of 1×10^{-6} was used to conduct GO annotations [25]. First, we use the *JrbHLLH* protein sequence to perform a blastp with an E-value of $1e-05$. The GO mapping analysis was used to obtain the GO annotation from the *JrbHLLH* members the GO enrichment analysis was carried out via the online GO enrichment program on the Omicshare website (<https://www.omicshare.com/tools/Home/Sof/gogsea>).

Interaction Network of *JrbHLLH* Proteins

The function of *JrbHLLH* protein based on *bHLLH* protein predicted in *Arabidopsis* due to lack of relevant information on walnut protein. The 189 walnut *bHLLH* proteins aligned with *Arabidopsis bHLLH* proteins using the Blastp program with an E-value of $1e-05$. The *JrbHLLH* protein interaction network was developed with *Arabidopsis* using homologous *bHLLH* proteins. The network was developed with default parameters on the String database (<https://string-db.org/>) using the input proteins of *J. regia* and six predicted input proteins [26].

Plant Materials, Treatments and Collections

A total of 26 samples from walnut were collected in this study, including 15 female flowers in different stages, the first opening of female flowers took place on April 10, April 15 and 22, and the full opening of female flowers took place (specifically, on April 15 and 22, stigma was not fully developed) and May 1 was the last date, 3 male flowers, 3 leaves, and 3 hulls. The female flowers (consist of 3 replicates) were collected on March 23, April 1, 8, 16 and April 23 respectively, the male flowers were collected on April 10, 11 and May 2 respectively, the leaves and the hull transcriptome data was from 'Chandler' walnut [18]. After harvesting, the pericarp was dissected immediately and stored at -80°C in liquid nitrogen. The total RNA has been isolated from leaves and other tissues using the Pre-Plant Kit (Tiangen, Beijing, China) [27]. After enough RNA extraction, the quality and quantity were checked with the help of NanoReady (Model: F-1100 made in China). Finally, build 23 cDNA libraries using FastKing RT Kit (Tiangen) and sequenced on the Illumina HiSeq 2500 platform. The differential gene expression analysis (DESeq) was performed using package DESeq R v.1.1.1. The predicted $P\text{-value} > 0.05$ have been assigned as expressed differently. In order to compare the

differences between the *bHLLH* gene expression level in reproductive tissues and vegetative tissues, we selected male and female flowers, young leaves, and hull with biological replicates to create a clear expression pattern.

Quantitative Real-Time PCR (qRT-PCR)

To validate the pattern of expression results, we used the tissues of female and male flowers, and leaves. The primer sequences details as *JrbHLLH13-5* (Forward: GTGGACGAGATAGCTCACGG, Reverse: TTCGTAGCCAGCGTCTTTGT), *JrbHLLH13-13* (Forward: CGAAGAGACGGGAAGACTGG, Reverse: ATGGTTCAAGCTCGCTCTCC), and *JrbHLLH13-14* (Forward: ACCAGGCCCTGAGTTCTGTA, Reverse: AGACCCTTGGCCTTTTGCTT). The synthetic cDNA was used as qRT-PCR template when diluted to 1/10 with sterile water. The iQTM SYBR[®] Green Supermix was used to perform qRT-PCR (Cat. 170-8880AP; Bio-Rad) on a Light Cycler 480 Real-Time PCR system (Roche Diagnostics, Laval, QC, Canada). The 18S rRNA has been used for internal gene control [28]. The primer specificities and corresponding melting curves were verified and repeated the experiment three times (triplicates) [10].

Results and Discussion

The Number, Phylogenetic Relationships and Location of *JrbHLLHs* in Persian Walnut

The conserved domain of *bHLLH* genes from *Arabidopsis* were as a query to identify the gene family members in walnut, the results showed that a total of 189 genes belong to the walnut *bHLLH* gene family. The *JrbHLLHs* were used to construct a phylogenetic tree with NJ method, it is clear that the *JrbHLLHs* were classified into 18 groups, and the number in these groups were unevenly distributed (Fig. 1). Based on their phylogenetic relationship walnut *bHLLH* genes were divided into 18 subfamilies (Fig. 1). Subfamily No. 13 reported the highest number of *JrbHLLH* genes (25), while family No. 11 reported the minimum number of *JrbHLLH* genes (4). In this study, we identified a total of 189 *bHLLH* genes in Persian walnut, compared to other plant members such as *Arabidopsis thaliana*, which contains 133 *bHLLH* genes [4, 16] and significant plants such as peanuts, potatoes, wheat, beans, rice, carrots, and tomatoes have *JrbHLLH* genes 132, 124, 188, 225, 155, 167, 146 and 159 [29, 30] (Fig. 1).

Gene Structure and Protein Domains of the *bHLLH* Gene Family of Persian Walnut

Structures including exons and introns were mapped to compare the structural parts of the 189 *JrbHLLH* genes (Fig. 2). The precise number and location of the domains of each Persian walnut protein were determined

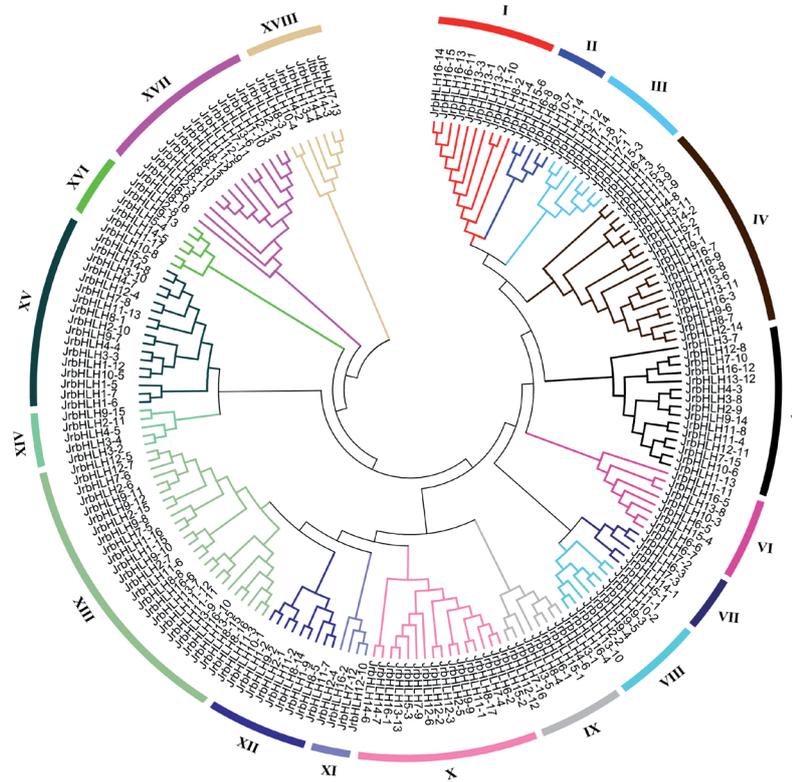


Fig. 1. Phylogenetic analysis of bHLH proteins in Persian walnut (189). A neighbor-joining (NJ) tree was constructed using 189 JrbHLH sequences. The tree further clustered into 18 subfamilies.

using the data available in the Pfam database (Fig. 2b). Conserved domain and motif valuation indicated that Motif 1 was present in all prevalent walnut *JrbHLH* genes. The subfamily of 16 and 17 consists of various motifs, such as 7, 8, 10, 15, 16, 17, and 19 also, as shown in the 18 family includes only motif 5, as well as a conserved motif (Fig. 2b). The analysis of the *JrbHLH* gene structure revealed large gene variations. It has been noted that the number of exons of Persian walnut *JrbHLH* transcript varies from 1 to 17 (Fig. 2c).

MEME analysis has shown that the walnut *JrbHLH* gene family consists of twenty-four residues of amino acids has been conserved and has unique domains for 189 walnut *JrbHLH* gene families. Among all these twenty-four conserved domains, one conserved domain (*HLH*) is made up of walnut *JrbHLH* genes, while the other 23 are unique domains. All of these conserved amino acid residues are compatible with previous *JrbHLH* domain research in other crops, such as peaches twenty-nine conserved amino acid residues. Based on conserved domains, walnut *JrbHLH* genes consist of 23 unique domains and one conserved domain (*HLH*) found in all *JrbHLH* genes [4, 22].

Synteny Analysis of *JrbHLH* Genes

The syntenic analysis was conducted for Persian walnut (*J. regia*) with other two woody plants (*P. trichocarpa* and *O. europaea*) to infer *bHLH* genes

evolutionary relationship between these species (Fig. 3). We identified 42 pairs of orthologous *bHLH* genes between *J. regia* and *P. trichocarpa*, while only 29 orthologous gene pairs between *J. regia* and *O. europaea* (Fig. 3). A total of 24 collinear gene pairs identified between *J. regia* and *P. trichocarpa* were not found between *J. regia* and *O. europaea*, 3 collinear gene pairs identified between *J. regia* and *O. europaea* were not found between *J. regia* and *P. trichocarpa*, importantly, 25 collinear gene pairs were found in three species *J. regia*, *O. europaea* and *P. trichocarpa* (Fig. 3). Gene organization plays a vital role in the evolution of multigene families [28]. Analysis of the gene structure indicated that exons present in the walnut *JrbHLH* gene ranged from 1-17 considered to exhibit the lower expression levels in plants [31, 32]. In the case of the *JrbHLH* gene in Persian walnuts, almost all gene structures have an intron 0 phase; therefore, the *JrbHLH* gene is evolutionarily preserved. The position of intron at 1, 2, 3, 4 and 5 were present throughout the *JrbHLH* family gene showed that the *JrbHLH* family is evolutionarily protected [14]. Since the evolutionary studies of *JrbHLH* transcription factors are limited in walnut need to be explored in further study (Fig. 3). There are collinear genes between the walnut and *P. trichocarpa* and *O. europaea* these results suggested that the *bHLH* genes may have evolved from the Persian ancestor in different plants [29, 33, 34].

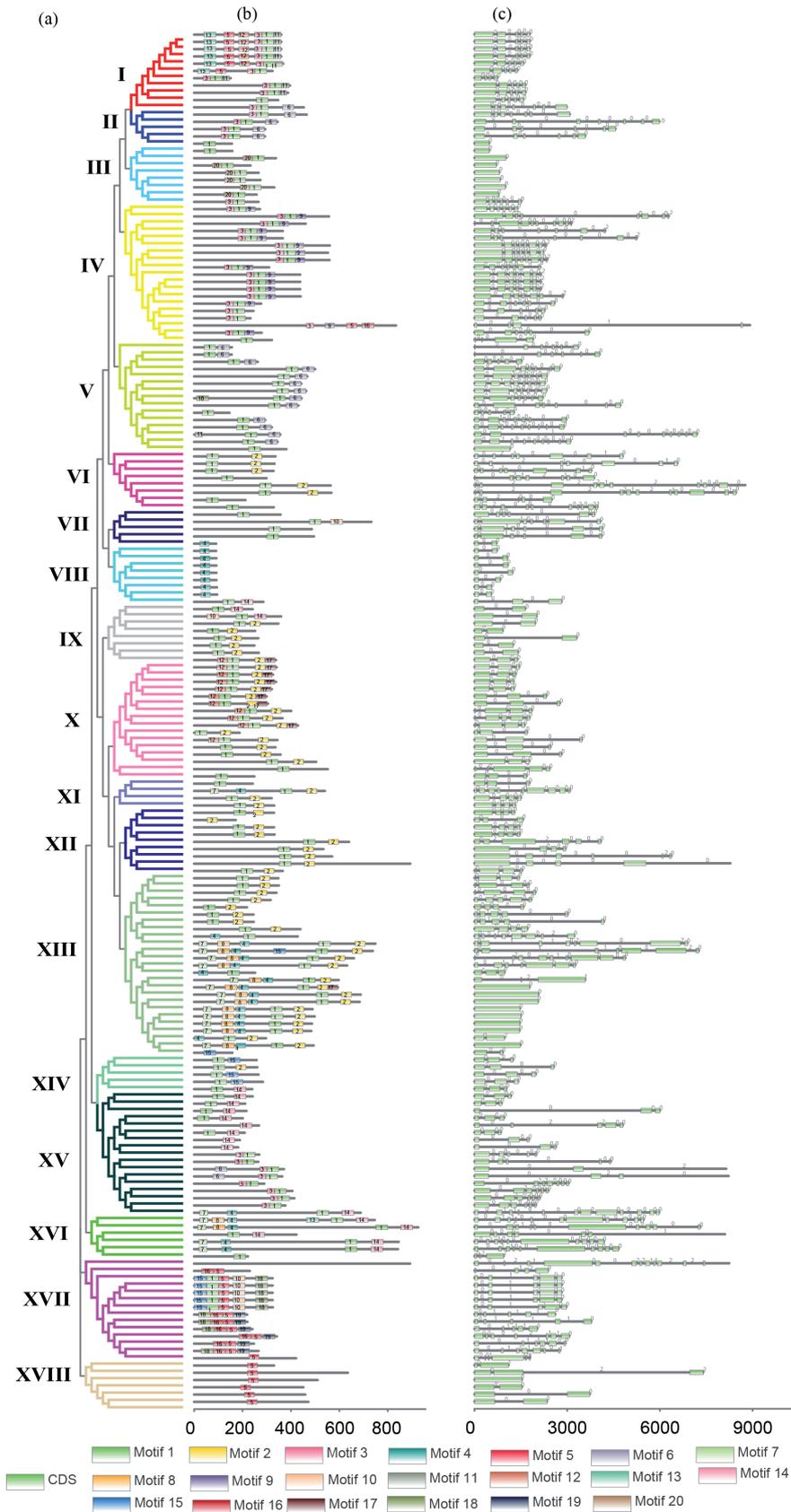


Fig. 2. The Phylogenetic relationship, motif compositions, and gene structure of JrbHLH transcription factors. a) The phylogenetic relationships of JrbHLHs based on NJ method. The various colors characterized the ten subfamilies. b) motif compositions of JrbHLHs. Gray lines indicated non-conserved sequences, and colored boxes represent conserved motifs. The motifs are displayed proportionally in each protein. c) gene structure of JrbHLHs. CDS represents coding sequence, green boxes indicate CDS, and gray lines represent Introns; 0, 1, and 2 represent different types of phase. Phase 0: located between two consecutive codons; Phase 1: splitting codon between the first and second nucleotides; Phase 2: between the second and third nucleotides of a codon.

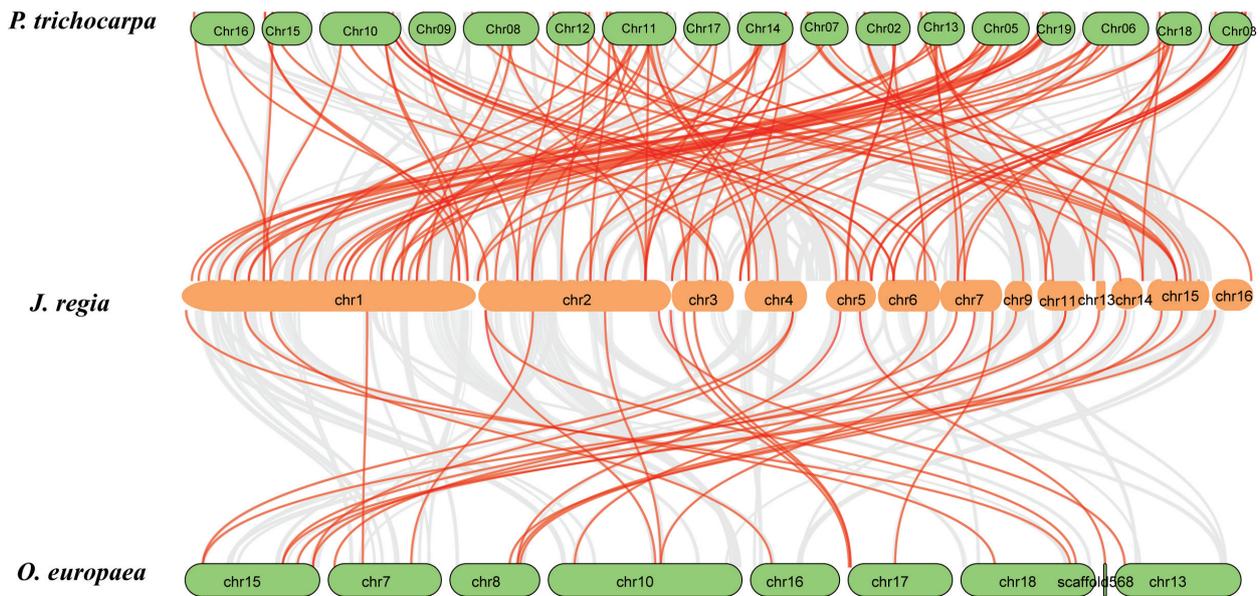


Fig. 3. Duplicate type, synteny analysis of *bHLH* genes between *J. regia* and two groups of representative plant species. Synteny analysis of *bHLH* genes between *J. regia* and three plant species. Gray lines in the background indicate the collinear blocks within Persian walnut and other plant genomes, while the red lines highlight the syntenic *BHLH* gene pairs. The species names with the prefixes '*P. trichocarpa*', '*O. europaea*' indicate *Populus trichocarpa*, *Olea europaea*, and *Quercus. robur*, respectively.

The Expansion and Evolution of the *JrbHLH* Gene Family of Persian Walnut

The analysis of GO enrichment based on 189 *JrbHLH* proteins annotated in the GO database. In the category of biological processes, bio-regulation, metabolic processes, cellular processes, and stimulus-response are significantly enriched terms. In the cellular component category, cell, organelle and cell parts are significant. GO terms for the transcription factor activities of nucleic acid binding were highly represented in the molecular function category. Besides, the most GO term enriched by members of the *JrbHLH* is GO: 003674 (molecular function). Tandem duplication, segmental duplication, and WGD were most likely chosen by gene families as forms of expansion [10]. However, a total of 69 *JrbHLHs* were duplicated as WGD, and no gene pair experienced tandem duplication events. These results indicated that the evolutionary expansion patterns of *bHLH* transcription factor family members were duplicated by WGD events (Fig. 1 and 2). These finding contrasts with several previous reports in which a similar phenomenon was analyzed [27, 35]. For example, the expansions of the *Gossypium*, *bHLH* genes were tandem duplication events [29, 34].

Transcriptome Expression Profile Analysis, qRT-PCR, and Interaction Work of *JrbHLH* Genes in Persian Walnut

Transcript expression analysis of different tissues of Persian walnut (female flower, male flower, young leaf, and hull) showed that the *JrbHLHs* expression levels were different in female flowers, and male flowers,

leaves, and hulls (Fig. 4). Expression profiling revealed that a total of 28 *JrbHLH* genes have higher expression in multiple tissues of Persian walnut [the values of FPKM (fragments per kilobase per million) were more than 60] (Fig. 4). A total of 4 and 8 members of *JrbHLH* genes show higher expression in female flowers and male flowers, respectively. For vegetative tissues transcriptome expression levels, we found that a total of 4 and 12 *JrbHLH* genes were expressed highly in leaves, while 12 *JrbHLH* genes were expressed highly in hulls. Among these high expressed genes, the *JrbHLHI3-10*, *JrbHLHI15*, and *JrbHLHI4-8* showed highly expression levers in both tissues (Fig. 4). Moreover, five *JrbHLH* genes were extremely high expressed (the values of FPKM were more than 400) (Fig. 4). Furthermore, by compared the differences between the *JrbHLH* gene expression levels in reproductive tissues and vegetative tissues, and we found that the two genes (*JrbHLHI3-13* and *JrbHLHI4-8*) both expressed high in reproductive tissues (female flowers and male flowers), while three genes (*JrbHLHI3-4*, *JrbHLHI4-3*, and *JrbHLHI4-4*) both expressed high in vegetative tissues (leaves and hulls) (Fig. 4). In woody plants, previous research showed that the *JrbHLH* gene plays an important role in flowering, such as peach and other fruit varieties, and some *JrbHLH* superfamily genes may be involved in fruit development [16]. This finding is consistent with previous findings of the *JrbHLH* protein, which also plays a regulatory role in tomato fruit ripening [29]. Especially in the reproductive development members of rice and *A. thaliana*, the functional subfamilies, *OsBHLHI42*, *AtbHLH091*, *OsBHLHI41*, *AtbHLH010* and *AtbHLH089* of these genes have similar expression patterns. While from the present we

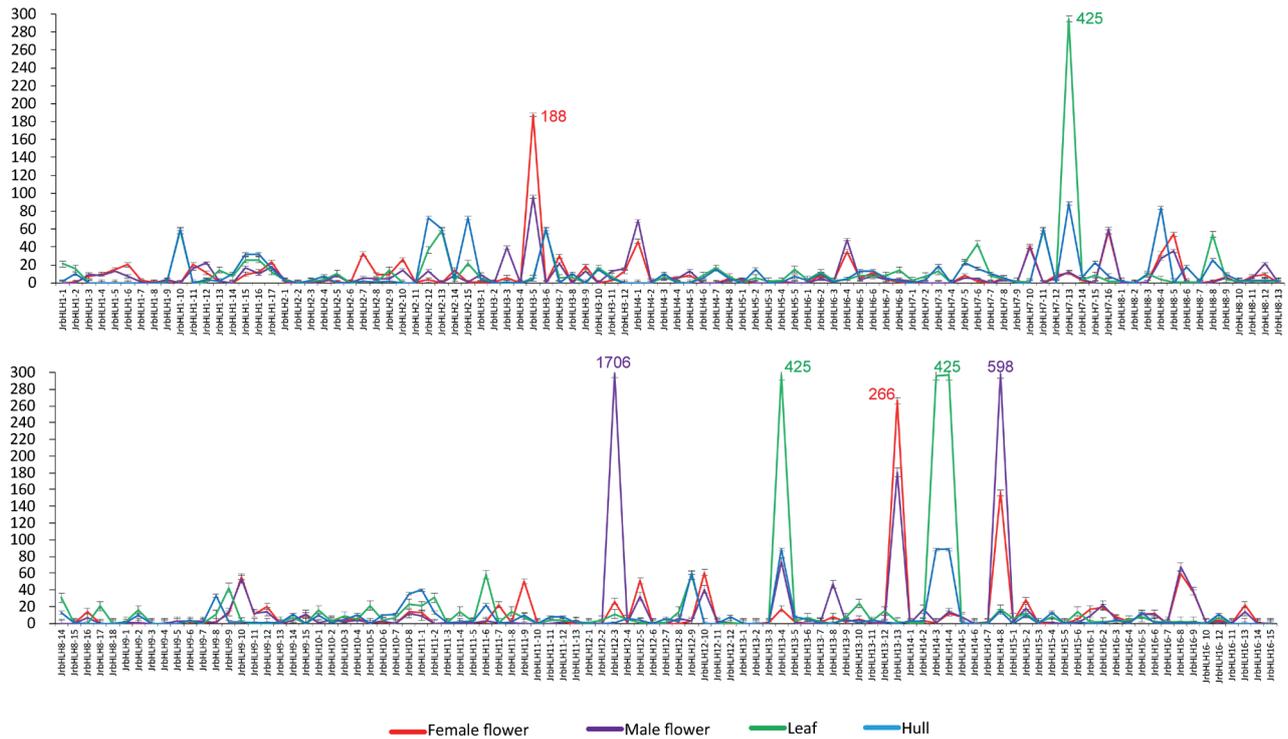


Fig. 4. The transcriptome expression profiles of a total of 189 Persian walnut bHLH genes in four tissues. Female flower, male flower, leaves, and hull as 3 biological replicates.

reported *JrbHLH13-4*, *JrbHLH1-15* and *JrbHLH14-8* in reproductive and vegetative tissues (Fig. 4). The same applies in the potato as the *StbHLHs* (*StbHLH26*, 85 and 99) have relatively high expression in flowers, in this study, 41 are highly expressed in flower tissues. These

genes may be involved in flower development [19, 36, 37].

The different expression patterns involving gradual increases and decreases observed in this study illustrate that some bHLH superfamily genes

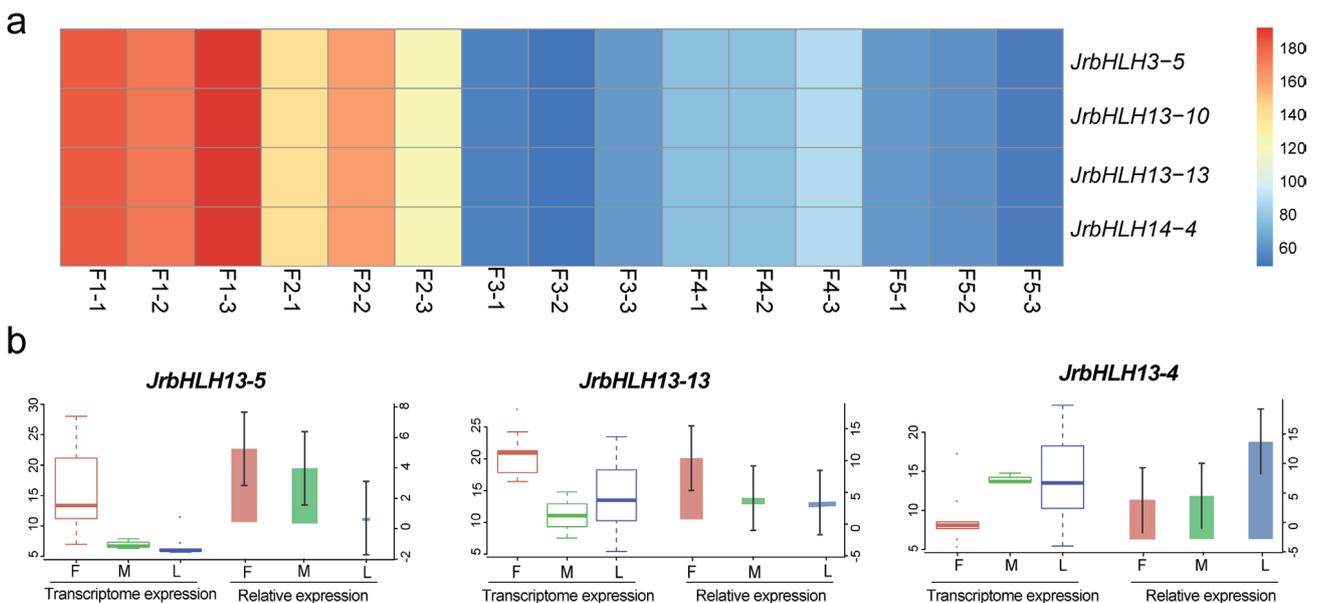


Fig. 5. Expression profiles of the Persian walnut bHLH genes. a) The heatmap exhibit the ratio of the expression levels of 12 bHLH genes in five developmental stages, 1, 2, 3, 4, and 5 represent different stages of Persian walnut flower. b) Expression analysis of 3 *JrbHLHs* in three representative samples by *qRT-PCR*. Data were normalized to β -actin gene and vertical bars indicate standard deviation. The details of samples see Table S2.

may be associated with fruit development. This finding was consistent with previous findings that *bHLH* proteins play a regulatory role in fruit ripening in walnut and other fruit species [37]. Additionally, the findings of this study are in agreement with transcriptomic results [37]. During the developmental stage (1st-5th) of the female flower, the transcriptome expression levels of *JrbHLH* genes were increased i.e. *JrbHLH3-5*, *JrbHLH13-4*, *JrbHLH13-13*, and *JrbHLH14-4* [37] (Fig. 5a). We further verified the difference gene expression level patterns in Persian walnut flowers and leaves used by qRT-PCR experiment (Fig. 5b). The results showed that *JrbHLH3-5* and *JrbHLH13-13* were highly expressed in female compared to male flowers, while *JrbHLH13-25* expressed highly in leaves of *J. regia* (Fig. 5b). These genes differentially expressed in female flowers, male flowers and leaves (Fig. 5b), which could be subsequently prioritized in plant functional studies for further analysis. Based on transcriptome and relative (qRT-PCR) expression, the walnut *JrbHLH* genes *JrbHLH3-5* and *JrbHLH13-13* show high expression in the female flower followed by the male flower, while the *JrbHLH13-4* shows greater expression in the vegetative part of the leaves followed by the male flower and also decreased expression in the female walnut family members of the *JrbHLH* gene. Therefore, further characterization of the 28 *JrbHLHs* is highly important and will provide a new insight to understand the molecular mechanism which may play some roles in the expression of the Persian walnut in multiple tissues.

Conclusions

In summary, we have recognized 189 *JrbHLH* genes in Persian walnut (*Juglans regia*) in this research. Phylogenetic analysis showed the division of the *JrbHLH* gene into 18 subfamilies. Also, expression profiling revealed that the *JrbHLH* gene showed distinct patterns of expression in distinctly prevalent walnut tissues. A total of 10 genes (*JrbHLH14-8*, *JrbHLH12-3*, *JrbHLH13-13*, *JrbHLH7-10*, *JrbHLH4-1*, *JrbHLH16-8*, *JrbHLH16-9*, *JrbHLH6-4*, *JrbHLH3-5*, and *JrbHLH11-9*) were highly reflected in multiple phases of female flowers and male flowers, which may play a significant role in Persian walnut flowers. During the flowering of female flowers, it was shown that some genes had increased expression rates as females grew. Overall, 28 members of the *JrbHLH* gene family are highly expressed in reproductive and vegetative tissues, while *JrbHLH13-4*, *JrbHLH1-15*, and *JrbHLH14-8* have been highly expressed in all tissues. Specifically, both transcript information and (qRT-PCR) analysis showed that two *JrbHLH* genes (*JrbHLH3-5*, *JrbHLH13-13*) were extremely expressed in female and male flowers, while one *JrbHLH* gene (*JrbHLH13-4*) was strongly expressed in leaves. Our outcome was consistent with that in Persian walnuts, but without any

combination with other tissues, the expression of the *JrbHLH* gene in male flowers was fully associated with female flowers.

Acknowledgments

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Conflict of Interest

The authors declare no conflict of interest.

References

- MURRE C., MCCAWE P.S., BALTIMORE D. A new DNA binding and dimerization motif in immunoglobulin enhancer binding, daughterless, MyoD, and myc proteins. *Cell*, **56**, 777, 1989.
- MAO K., DONG Q., LI C., LIU C., MA F. Genome wide identification and characterization of apple *bHLH* transcription factors and expression analysis in response to drought and salt stress. *Frontiers in plant science*, **8**, 480, 2017.
- SHEN W., CUI X., LI H., TENG R.-M., WANG Y.-X., LIU H., ZHUANG J. Genome-wide identification and analyses of bHLH family genes in *Brassica napus*. *Canadian journal of plant science*, **99**, 589, 2019.
- HEIM M.A., JAKOBY M., WERBER M., MARTIN C., WEISSHAAR B., BAILEY P.C. The basic helix-loop-helix transcription factor family in plants: a genome-wide study of protein structure and functional diversity. *Molecular biology and evolution*, **20**, 735, 2003.
- ZHANG T., LV W., ZHANG H., MA L., LI P., GE L., LI G. Genome-wide analysis of the basic Helix-Loop-Helix (*bHLH*) transcription factor family in maize. *BMC plant biology*, **18**, 235, 2018.
- MEIER-ANDREJSZKI L., BJELIĆ S., NAUD J.-F., LAVIGNE P., JELESAROV I. Thermodynamics of b-HLH-LZ Protein Binding to DNA: The Energetic Importance of Protein- DNA Contacts in Site-Specific E-Box Recognition by the Complete Gene Product of the Max p21 Transcription Factor. *Biochemistry*, **46**, 12427, 2007.
- WANG P., SU L., GAO H., JIANG X., WU X., LI Y., ZHANG Q., WANG Y., REN F. Genome-wide characterization of *bHLH* genes in grape and analysis of their potential relevance to abiotic stress tolerance and secondary metabolite biosynthesis. *Frontiers in plant science*, **9** 64, 2018.

8. CARRETERO-PAULET L., GALSTYAN A., ROIG-VILLANOVA I., MARTÍNEZ-GARCÍA J.F., BILBAO-CASTRO J.R., ROBERTSON D.L. Genome-wide classification and evolutionary analysis of the *bHLH* family of transcription factors in Arabidopsis, poplar, rice, moss, and algae. *Plant physiology*, **153**, 1398, **2010**.
9. PIRES N., DOLAN L. Early evolution of *bHLH* proteins in plants. *Plant signaling & behavior*, **5**, 911, **2010**.
10. FELLER A., MACHEMER K., BRAUN E.L., GROTEWOLD E. Evolutionary and comparative analysis of MYB and bHLH plant transcription factors. *The plant journal*, **66**, 94, **2011**.
11. HU Z., LV X., XIA X., ZHOU J., SHI K., YU J., ZHOU Y. Genome-wide identification and expression analysis of calcium-dependent protein kinase in tomato. *Frontiers in plant science*, **7**, 469, **2016**.
12. WEI K., CHEN H. Comparative functional genomics analysis of *bHLH* gene family in rice, maize and wheat. *BMC plant biology*, **18** (1), 309, **2018**.
13. ZHAO K., LI S., YAO W., ZHOU B., LI R., JIANG T. Characterization of the basic helix-loop-helix gene family and its tissue-differential expression in response to salt stress in poplar. *Peer J*, **6**, e4502, **2018**.
14. ZHANG X., LUO H., XU Z., ZHU Y., JI A., SONG J., CHEN S. Genome-wide characterisation and analysis of bHLH transcription factors related to tanshinone biosynthesis in *Salvia miltiorrhiza*. *Scientific reports*, **5**, 1, **2015**.
15. KAVAS M., BALOĞLU M.C., ATABAY E.S., ZIPLAR U.T., DAŞGAN H.Y., ÜNVER T. Genome-wide characterization and expression analysis of common bean *bHLH* transcription factors in response to excess salt concentration. *Molecular genetics and genomics*, **291**, 129, **2016**.
16. ZHANG C., FENG R., MA R., SHEN Z., CAI Z., SONG Z., PENG B., YU M. Genome-wide analysis of basic helix-loop-helix superfamily members in peach. *PloS one*, **13** (4), **2018**.
17. DUEK P.D., FANKHAUSER C. bHLH class transcription factors take centre stage in phytochrome signalling. *Trends in plant science*, **10**, 51, **2005**.
18. MARTÍNEZ-GARCÍA P.J., CREPEAU M.W., PUIU D., GONZALEZ-IBEAS D., WHALEN J., STEVENS K.A., PAUL R., BUTTERFIELD T.S., BRITTON M.T., REAGAN R.L. The walnut (*Juglans regia*) genome sequence reveals diversity in genes coding for the biosynthesis of non-structural polyphenols. *The plant journal*, **87**, 507, **2016**.
19. WANG R., ZHAO P., KONG N., LU R., PEI Y., HUANG C., MA H., CHEN Q. Genome-wide identification and characterization of the potato *bHLH* transcription factor family. *Genes*, **9**, 54, **2018**.
20. KUMAR S., NEI M., DUDLEY J., TAMURA K. MEGA: a biologist-centric software for evolutionary analysis of DNA and protein sequences. *Briefings in bioinformatics*, **9**, 299, **2008**.
21. EL-GEBALI S., MISTRY J., BATEMAN A., EDDY S.R., LUCIANI A., POTTER S.C., QURESHI M., RICHARDSON L.J., SALAZAR G.A., SMART A. The Pfam protein families database in 2019. *Nucleic acids research*, **47**, D427, **2019**.
22. MARCHLER-BAUER A., BO Y., HAN L., HE J., LANCZYCKI C.J., LU S., CHITSAZ F., DERBYSHIRE M.K., GEER R.C., GONZALES N.R. CDD/SPARCLE: functional classification of proteins via subfamily domain architectures. *Nucleic acids research*, **45**, D200, **2017**.
23. CE B.T.J.J.G., NOBLE W. The MEME suite. *Nucleic acids research*, **43**, W39, **2015**.
24. FOO E., YONEYAMA K., HUGILL C.J., QUITTENDEN L.J., REID J.B. Strigolactones and the regulation of pea symbioses in response to nitrate and phosphate deficiency. *Molecular plant*, **6**, 76, **2013**.
25. CONESA A., GÖTZ S., GARCÍA-GÓMEZ J.M., TEROL J., TALÓN M., ROBLES M. Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics*, **21**, 3674, **2005**.
26. SZKLARCZYK D., MORRIS J.H., COOK H., KUHN M., WYDER S., SIMONOVIC M., SANTOS A., DONCHEVA N.T., ROTH A., BORK P. The STRING database in 2017: quality-controlled protein-protein association networks, made broadly accessible. *Nucleic acids research*, **44**, 937, **2016**.
27. YAN F., LI H., ZHAO P. Genome-wide identification and transcriptional expression of the *PAL* gene family in Common walnut (*Juglans regia* L.). *Genes*, **10**, 46, **2019**.
28. XU G., GUO C., SHAN H., KONG H. Divergence of duplicate genes in exon-intron structure. *Proceedings of the national academy of sciences*, **109**, 1187, **2012**.
29. SUN H., FAN H.-J., LING H.-Q. Genome-wide identification and characterization of the bHLH gene family in tomato. *BMC genomics*, **16**, 9, **2015**.
30. MIAO L., GAO Y., ZHAO K., KONG L., YU S., LI R., LIU K., YU X. Comparative analysis of basic helix-loop-helix gene family among Brassica oleracea, Brassica rapa, and Brassica napus. *BMC genomics*, **21**, 1, **2020**.
31. REN X.-Y., VORST O., FIERS M.W., STIEKEMA W.J., NAP J.-P. In plants, highly expressed genes are the least compact. *Trends in genetics*, **22**, 528, **2006**.
32. JEFFARES D.C., PENKETT C.J., BÄHLER J. Rapidly regulated genes are intron poor. *Trends in genetics*, **24**, 375, **2008**.
33. VERVOORT M., LEDENT V. The evolution of the neural basic Helix-Loop-Helix proteins. *The scientific world journal*, **1**, 396, **2001**.
34. SONG X.-M., HUANG Z.-N., DUAN W.-K., REN J., LIU T.-K., LI Y., HOU X.-L. Genome-wide analysis of the *bHLH* transcription factor family in Chinese cabbage (*Brassica rapa* ssp. *pekinensis*). *Molecular genetics and genomics*, **289**, 77, **2014**.
35. WANG J., HU Z., ZHAO T., YANG Y., CHEN T., YANG M., YU W., ZHANG B. Genome-wide analysis of *bHLH* transcription factor and involvement in the infection by yellow leaf curl virus in tomato (*Solanum lycopersicum*). *BMC genomics*, **16**, 39, **2015**.
36. MEISSNER R.C., JIN H., COMINELLI E., DENEKAMP M., FUERTES A., GRECO R., KRANZ H.D., PENFIELD S., PETRONI K., URZAINQUI A. Function search in a large transcription factor gene family in Arabidopsis: assessing the potential of reverse genetics to identify insertional mutations in R2R3 MYB genes. *The plant cell*, **11**, 1827, **1999**.
37. TRAINOTTI L., BONGHI C., ZILLOTTO F., ZANIN D., RASORI A., CASADORO G., RAMINA A., TONUTTI P. The use of microarray μ PEACH1.0 to investigate transcriptome changes during transition from pre-climacteric to climacteric phase in peach fruit. *Plant science*, **170**, 606, **2006**.