Introduction

China is a large agricultural country; in 2016, it produced a total of 3.8 billion tons of livestock and poultry manure. To promote sustainable agricultural development, on 12 June 2017 the General Office of the State Council issued goals to accelerate the recycling of livestock and poultry waste. This will result in more livestock and poultry manure being treated for use on fruit and vegetable farmland. Many studies have shown that livestock and poultry manure contains large amounts of heavy metal pollutants [1] and, if used as a fertilizer, may contaminate farmland soil [2] and accumulate in grain, fruits, and vegetables [3], ultimately negatively impacting human health [4].
Heavy metal pollution is not only related to pollutant concentrations, but also to their speciation, migration, and bioavailability characteristics. According to the Tessier speciation scheme [5], heavy metals can be divided into exchangeable fractions, carbonate bound fractions, Fe-Mn oxide bound fractions, organically bound fractions, and residual fractions. Exchangeable heavy metal fractions are sensitive to environmental change, susceptible to migration and transformation, and can be absorbed by plants [6]. Carbonate bound heavy metal fractions are related to the pH of the soil. An increase in soil pH will result in carbonate co-precipitation. When soil pH decreases, the heavy metals are released again into the environment. This type of heavy metal can also be utilized by living organisms [7]. Fe-Mn oxide bound fractions are generally not easily utilized [8]. Organically bound fractions can be dissolved under certain conditions and may affect the environment [9]. Residual heavy metal fractions are generally stable in nature; they can remain in the soil in a passive state for long periods of time and are not easily absorbed by plants [10].

Therefore, in recent years, some researchers have attempted to change the forms of heavy metals in the environment using treatment methods that transform them from bioavailable to bio-unavailable forms. Dong et al. [11] reported that high solid anaerobic digestion increased the bioavailability of Cu, Zn, Ni, and Cr, while decreasing the bioavailability of Pb. Lv et al. [12] found that vermicomposting decreased the mobility and bioavailability of heavy metals, and increased the relative content of bio-unavailable heavy metal forms. Liu et al. [13] showed that composting can reduce 70% of exchangeable Cd, and immobilize Cd in soil; thus, composting can be used for remediation of Cd-contaminated soils.

China is one of the biggest apple producing countries in the world, where apple orchards cover an area of 23,292 million m². At a rate of 3 kg of fertilizer per m² per year, 69.84 million tons of organic fertilizer are required each year. Additionally, at a rate of 0.3-0.6 tons of organic fertilizer per ton of livestock and poultry manure, approximately 116.4-232.8 million tons of manure are used. Therefore, heavy metal contamination of orchard soil and fruit is a problem deserving of significant attention. Wang et al. [14] studied the heavy metal content of soil and fruit from 48 apple orchards located in the Liaodong Peninsula. They found that Cr and Cu concentrations in some soil samples exceeded the allowable limits for heavy metal contents in orchard soils in China, and Cr, Cd, and Zn concentrations in some fruit samples exceeded the Chinese agricultural product safety requirements for pollution-free fruit. These results were attributed to the natural physicochemical properties of the soil [15, 16]. Wang et al. [17] reported that long-term use of Cu-based pesticides causes the Cu content in apple orchard soils to increase each year, indicating that pesticide and fertilizer use affects heavy metal contents in the soil. In this study, the content and forms of heavy metals found in chicken manure, compost, apple orchard soil, and fruit were studied in order to clarify the migration and accumulation patterns of heavy metal contaminants within the livestock manure-compost-soil-fruit system.

Material and Methods

Sampling

The experiments in this study involved an apple orchard in Shandong Province, China. There are 11 broiler chicken farms located in the vicinity of the apple orchard (breeding scale of 20,000-100,000). The resulting chicken manure is consolidated and transported to a nearby organic fertilizer plant where it is mixed with rice husk powder to a C:N ratio of 25. Next, 1% calcium superphosphate is added, and a dose of fermentation bacteria is introduced. The water content is adjusted to approximately 50%-60%, and after being fully mixed, the mixture is placed into solid-state fermentation tanks. After 24 hours of forced-ventilation fermentation, the resulting material is stockpiled into 1-1.5 m high and 4-5 m wide heaps that continue to decompose. The heaps are turned over once every day and, after 21 days, the fermentation process is completed. The heaps are sun-dried until the water content reaches 30%-35% and then stored. Every October, chicken manure compost is applied to the orchard at a rate of 3 kg/ m², using a combination of trenching and sprinkling. Trenching application involved digging a 40-cm wide and 50-cm deep trench between the fruit tree rows, and filling the bottom 10 cm of the trench with straw and weeds. Next, a mix of the manure compost and soil was applied and the trench was filled with soil. The sprinkling application method involved spreading the manure compost directly under the trees and turning over the soil. Sampling was performed during September 2016. The chicken manure samples were taken from the breeding farms, the manure compost samples were taken from the organic fertilizer plant, and the soil and apple fruit samples were taken from the apple orchards. A Dutch shovel was used for the variable-depth soil samples. At each depth (0-10 cm, 10-20 cm, 20-30 cm, 30-40 cm), five random sampling sites were selected and 200 g of soil for each sampling site was mixed evenly, dried naturally, ground to pass through a 1-mm sieve, and stored at 4°C. The apple fruit sample surfaces were thoroughly washed, peeled, cored, homogenized, and stored at 4°C.

Detection and Analysis

The total extraction method involved weighing 1-5 g samples (to a precision of 0.001 g) and placing
them in a 100-mL beaker. Next, 20 mL of aqua regia (HCl: HNO₃ = 3:1) was added, and the beaker was covered with a watch glass. Next, the contents were micro-boiled on a heating plate at a temperature of 150ºC-200ºC for 30 minutes. When the contents were almost dry, the beaker was removed from the heat. The watch glass and the inner wall of the beaker were then rinsed with a small amount of water. After cooling, 2 mL of 50% hydrochloric acid solution was added and heated until dissolved, then cooled and filtered. The filtrate was collected into a 50-mL graduated flask. After the filtrate had dried, it was rinsed more than three times using a small amount of ultra-pure water. The filtrates were then combined, and ultra-pure water was added to 50 mL and mixed thoroughly before measurement.

The Tessier continuous extraction method was used for heavy metal speciation analysis [5]. The following fractions were obtained: exchangeable, carbonate bound, Fe-Mn oxide bound, organically bound, and residual. An ICPE-9000 spectrometer (Shimadzu, Japan) and inductively coupled plasma optical emission spectrometry analysis (ICP-OES) were used. The heavy metal standard was supplied by the National Standard Material Center of the Beijing Wanjia Shouhua Biotechnology Co., Ltd.

Statistical Analysis

The experimental data were expressed using the mean values of three parallel sets of data. Standard deviations were also calculated. Origin 9.0 was used for mapping, and SPSS (19.0) was used for the statistical analysis. ANOVA (analysis of variance) and Tukey’s range test were employed for the statistical analysis, with a significance level of \( p < 0.05 \).

Results and Discussion

Heavy Metal Content in Chicken Manure

Table 1 shows the heavy metal content in chicken manure raw materials from large-scale commercial broiler farms. The order of mean heavy metal contents was found to be Zn>As>Cu>Cr>Pb>Hg>Cd. The median values were essentially consistent with the mean values. Low standard deviation values reflect small variations in the heavy metal contents of various manure samples. This might be due to similarities in chicken coop design, breeding scale and technology, the proportion and quantity of feed, and other conditions at the various farms. The skewness coefficient, which is a measure of the asymmetry of the distribution relative to a normal distribution, confirms a normal distribution of heavy metals in the chicken manure. The kurtosis values were extremely low owing to the fact that most samples clustered around the central mean value.

These statistical properties show that the mean and median values correspond to the concentrations of the heavy metals. As, Cd, and Hg contents in the chicken manure exceeded the limits for heavy metal contents of organic fertilizer products, as specified by the Organic Fertilizer Standard NY 525-2012. The main source of As in the chicken manure was found to be As-containing antibiotics used during the breeding process. Most of these antibiotics are not absorbed by the poultry, and are excreted through urine and feces [18]. The Cd and Hg contents in the chicken manure also exceed standard values because fishmeal, which is commonly added to chicken feed, is generally high in Cd and Hg [19]. No abnormal values were observed in box plots (Fig. 1), meaning that the heavy metal content of the chicken manure was not affected by external factors.

Table 1. Heavy metal contents (mg/kg) of chicken manure.

<table>
<thead>
<tr>
<th></th>
<th>As</th>
<th>Cd</th>
<th>Hg</th>
<th>Cr</th>
<th>Pb</th>
<th>Cu</th>
<th>Zn</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean</td>
<td>74.46</td>
<td>4.30</td>
<td>14.93</td>
<td>17.85</td>
<td>15.23</td>
<td>45.33</td>
<td>152.50</td>
</tr>
<tr>
<td>Standard deviation</td>
<td>13.22</td>
<td>1.37</td>
<td>2.82</td>
<td>5.01</td>
<td>7.53</td>
<td>5.56</td>
<td>22.47</td>
</tr>
<tr>
<td>Q1</td>
<td>62.19</td>
<td>3.84</td>
<td>12.93</td>
<td>13.64</td>
<td>10.85</td>
<td>41.91</td>
<td>145.61</td>
</tr>
<tr>
<td>Median</td>
<td>71.46</td>
<td>4.25</td>
<td>13.70</td>
<td>17.24</td>
<td>11.57</td>
<td>45.21</td>
<td>155.07</td>
</tr>
<tr>
<td>Q3</td>
<td>85.83</td>
<td>5.28</td>
<td>16.62</td>
<td>22.53</td>
<td>18.67</td>
<td>48.35</td>
<td>163.08</td>
</tr>
<tr>
<td>Minimum</td>
<td>56.34</td>
<td>1.52</td>
<td>11.71</td>
<td>12.11</td>
<td>7.46</td>
<td>36.83</td>
<td>107.20</td>
</tr>
<tr>
<td>Maximum</td>
<td>93.57</td>
<td>6.03</td>
<td>20.44</td>
<td>26.08</td>
<td>27.90</td>
<td>55.83</td>
<td>183.75</td>
</tr>
<tr>
<td>Skewness</td>
<td>0.12</td>
<td>−0.92</td>
<td>0.81</td>
<td>0.39</td>
<td>1.08</td>
<td>0.24</td>
<td>−0.75</td>
</tr>
<tr>
<td>Kurtosis</td>
<td>−1.59</td>
<td>0.35</td>
<td>−0.25</td>
<td>−1.48</td>
<td>−0.66</td>
<td>−0.05</td>
<td>0.54</td>
</tr>
<tr>
<td>NY 525-2012a</td>
<td>≤15</td>
<td>≤3</td>
<td>≤2</td>
<td>≤150</td>
<td>≤50</td>
<td>—</td>
<td>—</td>
</tr>
</tbody>
</table>

a Organic fertilizer; Chinese agricultural industry standard (NY 525-2012).
Effects of Composting on Heavy Metal Content and Speciation

Fig. 2 compares the heavy metal contents in chicken manure before and after composting. The heavy metal contents increase after composting, within the significance level ($p<0.05$), mainly due to the reduction in volume and weight caused by decomposition of the organic matter [12, 20, 21]. In addition to the change in the total content of heavy metals, significant variations in heavy metal speciation also occurred (Fig. 3). The most significant and important changes occurred in the exchangeable fractions of As, Cd, and Hg, which decreased from 44.58%, 16.65%, and 14.62% before composting to 35.25%, 1.82%, and 3.18% after composting, respectively, and in the residual fractions of As, Cd, and Hg, which increased from 0.68%, 3.68%, and 4.31% before composting to 14.62%, 23.02%, and 20.76% after composting, respectively.

The reduction in highly reactive forms and the increase in stable forms of heavy metals indicate that the composting process has a significant effect on the passivation of heavy metals contained in chicken manure. The increase in the carbonate bound fractions of As, Cd, and Hg after composting related to the change in pH during composting. Fe-Mn oxide bound fractions are thermodynamically unstable under hypoxic conditions. Although the high-temperature composting process in this study mainly uses aerobic fermentation, it is impossible to avoid anaerobic fermentation. The Fe-Mn oxide bound fractions of As, Cd, and Hg decreased from 36.52%, 68.22%, and 49.45% to 16.23%, 25.16%, and 28.14% after composting, respectively. Free heavy metal ions can react with and be bound by many types of organic matter. As a result, the organically bound fraction contents of As, Cd, and Hg increased from 2.11%, 7.43%, and 19.98% to 16.05%, 24.90%, and 24.94%, respectively, after composting. Studies on the increase of organically bound fractions of Cd due to composting have already been published [13]. Changes in As, Cd, and Hg speciation after composting indicates that the composting process can stabilize and immobilize heavy metals, and change their migration and bioavailability. Therefore, composting can reduce the relative amount of heavy metal absorption and accumulation in crops, reducing the risk of environmental pollution, as also shown by He et al. [22].

Heavy Metal Contents and Speciation in Apple Orchard Soils

The distribution of As, Cd, and Hg in the apple orchard soil cross-section was non-homogenous (Fig. 4), consistent with findings for Cu and Zn [23]. As and Cd concentrations were lower in the soil surface
layer (0-10 cm) than in deeper layers (10-40 cm), possibly due to the fact that As and Cd dissolve and migrate downward after rain or irrigation. Moreover, Hg in the soil surface layer reached up to 16.81 mg/kg, far exceeding the content of deeper layers. This might be because the surface layer is the most vulnerable to contamination by external Hg, and organic matter-rich soils display particularly strong adsorption and accumulation of Hg [24].

As, Cd, and Hg speciation varies significantly at different soil depths (Fig. 5). The exchangeable and carbonate bound fractions of As in the surface layer (0-10 cm) were very high (55.52% and 25.46%, respectively), with a combined value of 80.97%. The exchangeable and carbonate bound fractions of Cd and Hg were also very high (12.02% and 76.02%, and 15.75% and 60.50%, respectively), totaling 88.04% and 76.25%, respectively. The ratio of exchangeable and carbonate bound fractions to the total heavy metal content, which is known as the mobility factor or the activity coefficient, is used to assess the migration potential and bioavailability of heavy metals. Higher exchangeable and carbonate bound fraction contents indicate stronger migration potential and higher absorption and accumulation of heavy metals in crops [25, 26]. The contents and speciation of heavy metal elements As, Cd, and Hg in the deeper soil layers (10-40 cm) were approximately equal, and their activity coefficients were 43.81%-52.96%, 39.64%-43.30%, and 37.31-39.24%, respectively, lower than those in the surface layer. This indicates that heavy metals in deeper layers have lower mobility and bioavailability.

Migration and Accumulation of Heavy Metals in the Soil-Apple Fruit Systems

Table 2 shows the heavy metal contents of apple orchard soil and fruit. Cd and Hg concentrations in the apple orchard soil far exceeded the limits specified by the Environmental Quality Standard for Soils, GB 15618-1995. The contents of the other heavy metals (As, Cr, Pb, Cu, and Zn) were below the national standard. This indicates that the apple orchard soil was contaminated with Cd and Hg. As, Cd, and Hg concentrations in the apples exceeded the food pollutant limits specified by national food safety standards, especially for Hg, which was 25 times above the allowable limit. Consequently, finding and removing the contamination source merits serious attention. From highest to lowest, the order of migration rates

Fig. 4. As, Cd, and Hg concentrations at various depths of apple orchard soil. Different letters indicate significant changes (p<0.05). Columns annotated by the same letter are not significantly different (p>0.05).

Fig. 5. Speciation of a) As, b) Cd, and c) Hg at different soil depths.
of heavy metals from the soil to the apple fruit was found to be Cu>As>Zn>Hg>Cr>Cd> and Pb. The high migration rate of As in apple fruit is the main reason for As exceeding the standard. In addition to the use of chicken manure compost, whose heavy metal contents exceeded the standard, the physicochemical properties of the orchard soil, the use of pesticides and chemical fertilizers, and atmospheric deposition all contribute to soil and fruit heavy metal contents that exceed national standards [27-29].

Conclusions

This investigation of heavy metal content and speciation within the chicken manure-compost-soil-apple system showed that the contamination levels of As, Cd, and Hg remained high throughout the system, whereas levels of other heavy metals remained fairly low. This indicates that heavy metals contained in chicken manure can migrate and accumulate in soil and crops, thus posing a significant threat to ecosystem safety and human health.

Acknowledgements

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Conflict of Interest

The authors declare no conflict of interest.

Table 2. Heavy metal contents (mg/kg) of apple orchard soil and fruit.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>As</th>
<th>Cd</th>
<th>Hg</th>
<th>Cr</th>
<th>Pb</th>
<th>Cu</th>
<th>Zn</th>
</tr>
</thead>
<tbody>
<tr>
<td>Apple orchard soil*</td>
<td>10±1</td>
<td>6.15±0.64</td>
<td>5.12±0.39</td>
<td>23±3</td>
<td>0</td>
<td>12±3</td>
<td>18±4</td>
</tr>
<tr>
<td>Standard for soil*</td>
<td>≤40</td>
<td>≤0.30</td>
<td>≤0.30</td>
<td>≤150</td>
<td>≤250</td>
<td>≤150</td>
<td>≤200</td>
</tr>
<tr>
<td>Apple</td>
<td>1.1±0.1</td>
<td>0.07±0.01</td>
<td>0.25±0.05</td>
<td>0.3±0.0</td>
<td>ND</td>
<td>4±0</td>
<td>2±0</td>
</tr>
<tr>
<td>Food contamination limit*</td>
<td>≤0.5</td>
<td>≤0.05</td>
<td>≤0.01</td>
<td>≤0.5</td>
<td>≤1</td>
<td>≤10</td>
<td>≤5</td>
</tr>
<tr>
<td>Migration rate*</td>
<td>0.106</td>
<td>0.011</td>
<td>0.049</td>
<td>0.012</td>
<td>0</td>
<td>0.309</td>
<td>0.090</td>
</tr>
</tbody>
</table>

*Soil depth: ~10-40 cm.

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