Introduction

Lignocellulosic biomass has been regarded as an abundantly available resource to produce bioenergy and other biochemical products [1-2] since it is rich in carbohydrate polymers that can be decomposed into hexose and pentose sugars. Wheat straw is one kind of the largest lignocellulosic agricultural byproducts and its annual output throughout the world is estimated at about 850 million tons [3-4]. The traditional utilization (animal feed, agricultural fertilizer, rural solid-fuel, etc.) cannot consume wheat straw completely and the residue is a big challenge for the environment and public health [5]. In recent decades, anaerobic digestion (AD) of wheat straw to produce biogas has been studied both in the lab and on industrial scales, whereas effective and economical pretreatments are still required to improve the digestibility of wheat straw. Without pretreatment, the cellulosic biomass would be much more difficult to hydrolyze completely, resulting in a poor biogas yield and long fermentation period.

A wide variety of methods (e.g., bases, acids, high temperature, milling, steam explosion, catalyzed hydrolyses, and their combination) have been used to pretreat straw to increase its biodegradability [6-7]. Among them, alkaline pretreatment is regarded as one of the most efficient methods and sodium hydroxide (NaOH) is widely used for its outstanding delignification capacity and simple operation [8-9]. Carrillo et al. [10] indicated that the hydrolysis degree of wheat straw pretreated by NaOH was more than three times that compared with untreated wheat straw. Taherdanak et al.[11] also found that the lignin and hemicellulose in wheat plants were
reduced using 8% (wt) NaOH for pretreatment, while the highest yield of methane production and glucose content were obtained by pretreatment at 75°C for 60 min.

NaOH pretreatment was proved effective in treating biomass, particularly for lignocellulose [12]. It may cause lignocellulose to change in both chemical property and physical structure, for instance, swell leading to an increase in the internal surface area, a decrease in the degree of polymerization and crystallinity, separation of structural linkages between lignin and carbohydrates, and disruption of the lignin structure [13-14]. These changes will release a mass of organic matter (lipid, monose, and other carbohydrates) into liquid. On the one hand, NaOH pretreatment can enhance hydrolysis of substrates and improve yield of sugar [15]. But on the other hand, a lot of low molecular weight organic matter that is easily available for microorganisms will be wasted and may pollute the environment [16].

Organic carbon is an important carbon resource for microorganisms in AD [17-18]. As a consequence, to evaluate a pretreatment method, not only the yield of sugar or rate of hydrolysis should be taken into consideration, but the loss of organic matters should be given greater importance.

This study investigated the effects of NaOH pretreatment on wheat straw at a mesophilic temperature. The hydrolysis of pretreated wheat straw by different NaOH concentrations and raw material were compared. The changes in composition and structure of the pretreated wheat straw were also detected. Finally, the losses of total organic carbon in different pretreatment conditions were presented and compared.

**Materials and Methods**

**Wheat Straw**

The wheat straw was collected from a farm in Henan province, China. After collection, it was cut into 2 cm and washed twice with purified water to remove soil and dust. Afterward it was dried at 70°C to constant weight and stored at 3°C to keep raw material fresh according to the previous report [19]. The basic compositions of raw wheat straw are given in Table 1.

<table>
<thead>
<tr>
<th>Constituent</th>
<th>Unit</th>
<th>Mean ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>TS</td>
<td>%FM*</td>
<td>94.7±1.4</td>
</tr>
<tr>
<td>VS</td>
<td>%TS</td>
<td>95.3±2.1</td>
</tr>
<tr>
<td>Hemicellulose</td>
<td>%TS</td>
<td>23.1±2.2</td>
</tr>
<tr>
<td>Cellulose</td>
<td>%TS</td>
<td>34.1±2.7</td>
</tr>
<tr>
<td>Lignin</td>
<td>%TS</td>
<td>16.7±1.8</td>
</tr>
<tr>
<td>Ash</td>
<td>%TS</td>
<td>4.4±1.1</td>
</tr>
</tbody>
</table>

*SFM: fresh matter

**Sodium Hydroxide Pretreatment**

The pretreatments of wheat straw using NaOH were carried out in bottles with a total volume of 3 L. Firstly, 65 g of wheat straw was added into each bottle. And then 1.5 L NaOH solution with concentrations of 0%, 4%, 6%, and 8% (wt) was added to each bottle. The bottles were kept in an incubator (Hadonglian, China) at 35°C. During the pretreatment, 4 g substrate was taken from each bottle for analysis after two, five, and eight days.

**Enzymatic Digestion Of Wheat Straw**

Enzymatic digestion was performed according to previous reports [20-21]. Raw material and pretreated wheat straw were milled into 40 meshes and then sealed in a bag with bore diameter smaller than 80 meshes so that the products of hydrolysis could pass through the bag and the residual wheat straw would be left in it. Then 0.2 g wheat straw and 4 ml enzyme cocktail (15FPU/g wheat straw) dissolved in 16 ml acetate buffer (pH 4.7) were put into a 50 ml erlenmeyer flask. Then all the erlenmeyer flasks were sealed and incubated at 45°C and 120 rpm for 8, 24, and 48 h. Each enzymatic hydrolysis assay was performed in triplicate. Enzymatic digestion rate was calculated as in Eq. (1) [20]:

\[
\text{Enzymatic digestion} \% = \frac{m_1-(m_2\times c)}{m} (1)
\]

...where \(m\) is the dry weight of substrate, \(m_1\) is the total weight of substrate and the bag, \(m_2\) is the total weight of remaining substrate and bag, and \(c\) is the correction coefficient of the bag (weight ratio of the bag before hydrolysis to the bag after hydrolysis).

**Substrate Analysis**

Total solids (TS) and volatile solids (VS) were determined according to standard methods [22]. Neutral detergent fiber (NDF), acid detergent fiber (ADF), and acid detergent lignin (ADL) were measured by a fiber analyzer (200l, ANKOM, USA) according to procedures supplied by ANKOM Technology [23]. After analyzing ADL, samples were ignited at 550°C for 4 h in a muffle oven to get the ash. Hemicellulose, cellulose, and lignin were calculated according to Eqs. (2), (3), and (4):

\[
\text{Hemicellulose} \% = \text{NDF} \% - \text{ADF} \% \quad (2)
\]
\[
\text{Cellulose} \% = \text{ADF} \% - \text{ADL} \% \quad (3)
\]
\[
\text{Lignin} \% = \text{ADL} \% - \text{Ash} \% \quad (4)
\]

**Microscope Observation**

A scanning electron microscope (SEM, S-4800, Hitachi, Japan) was used to visualize the superficial differences between untreated and pretreated wheat straw.
at a magnification of 500×. The wheat straw was metalized with a thin layer of gold in a sputter coater polaron (E-1045, Hitachi, Japan) to ensure the conductivity of its observation.

FTIR Observation

Fourier transform infrared spectroscopy (FTIR) spectra was obtained by a Nicolet iS10 FTIR spectrophotometer (Thermo Fisher Scientific, USA). Each spectrum was obtained with an average of two scans and a resolution of 4 cm⁻¹, from 500 to 4,000 cm⁻¹ [11].

TOC Analysis

Total organic carbon (TOC) was measured by a DR6000 spectrophotometer (Hach, USA). The liquid remaining in each bottle was collected and centrifuged at 10,000 rpm for 10 min (HC-2066, ZONKIA, China), and the supernatant was analyzed. According to a TOC test method provided by the Hach Company [24], the standard agents were added into the diluted sample successively, and then the sample was digested at 102ºC for 2 h. After cooling to room temperature, the sample was measured at 430 nm. The loss ratio of TOC was calculated according to Eq. (5):

\[
\text{Loss ratio of TOC (\%) = } \frac{V \times c}{m} \quad (5)
\]

…where \( c \) is the concentration of TOC (g/L), \( V \) is the volume of liquid phase in L, and \( m \) (g) is the dry weight of the substrate.

Results and Discussion

The Influence of NaOH-pretreatment on Hydrolysis of Wheat Straw

In order to understand the effects of NaOH pretreatment on wheat straw, the compositions, including hemicellulose, cellulose, and lignin (as well as ash) of wheat straw before and after pretreatment were tested. TS and VS of wheat straw pretreated by different concentrations of NaOH for 2, 5, and 8 days were compared. On the whole, as listed in Table 2, TS and VS were reduced with the increase of pretreatment time and NaOH concentration.

The raw wheat straw containing 23.1% hemicellulose, 34.1% cellulose, 16.7% lignin, and 4.4% ash (Table 1) was similar to other reports [25-26]. As shown in Fig. 1, after pretreatment the cellulose and hemicellulose had a small reduction comparing to the untreated straw. With the increase of NaOH concentration and pretreatment time, the contents of cellulose and hemicellulose decreased slowly. The maximum losses of cellulose and hemicellulose were received when using 8% NaOH to treat for eight days, which were 41.2% and 51.1%, respectively, on the basis of contents in raw wheat straw. While for lignin, the loss was greatly caused once NaOH was added, and the maximum was 77.2% when 6% NaOH was used to treat for five days. These changes indicated that NaOH pretreatment affected the physical structure and chemical properties of wheat straw.

Lignin is thought to be one of the strongest resistance materials for enzymes and microorganisms during anaerobic digestion. The results above proved that, compared with raw material, NaOH pretreatment of wheat straw at a mesophilic temperature could reduce lignin significantly, but when the NaOH concentration and pretreatment time increased, there was no obvious enhancement of this effect. So the pretreatment conditions should be optimized in order to keep cellulose and hemicellulose and decrease lignin, as well as to reduce the treatment cost and environmental impact.

<table>
<thead>
<tr>
<th>Pretreatment time</th>
<th>Concentration of NaOH (wt)</th>
<th>TS (%FM)</th>
<th>VS (%TS)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2 days</td>
<td>0%</td>
<td>93.4 ± 4.8</td>
<td>92.3 ± 5.6</td>
</tr>
<tr>
<td></td>
<td>4%</td>
<td>92.7 ± 7.3</td>
<td>95.1 ± 2.2</td>
</tr>
<tr>
<td></td>
<td>6%</td>
<td>89.2 ± 12.9</td>
<td>94.4 ± 2.7</td>
</tr>
<tr>
<td></td>
<td>8%</td>
<td>90.5 ± 5.3</td>
<td>89.9 ± 4.7</td>
</tr>
<tr>
<td>5 days</td>
<td>0%</td>
<td>87.2 ± 4.1</td>
<td>93.5 ± 1.1</td>
</tr>
<tr>
<td></td>
<td>4%</td>
<td>90.6 ± 6.7</td>
<td>92.7 ± 4.8</td>
</tr>
<tr>
<td></td>
<td>6%</td>
<td>84.3 ± 0.9</td>
<td>93.6 ± 3.1</td>
</tr>
<tr>
<td>8 days</td>
<td>0%</td>
<td>87.6 ± 3.4</td>
<td>91.4 ± 1.8</td>
</tr>
<tr>
<td></td>
<td>4%</td>
<td>81.1 ± 1.2</td>
<td>93.1 ± 4.8</td>
</tr>
<tr>
<td></td>
<td>6%</td>
<td>82.4 ± 3.3</td>
<td>90.8 ± 1.7</td>
</tr>
<tr>
<td></td>
<td>8%</td>
<td>78.6 ± 3.5</td>
<td>88.3 ± 6.7</td>
</tr>
</tbody>
</table>

Table 2. Comparison of TS and VS of wheat straw pretreated by different NaOH concentration and time.
straw can be distinguished by inspecting the peak at 1,731-1,733 cm\(^{-1}\) region, which corresponds to ester linkages of hemicellulose [27]. It is obvious that the transmittance of pretreated wheat straw at 1,732 cm\(^{-1}\) is higher than that of raw wheat straw, which indicates that some hemicellulose was removed after pretreatment [14]. There are some notable differences in bands at 1,610 cm\(^{-1}\) and 1,510 cm\(^{-1}\), which are characteristic bands of the aromatic ring of lignin, suggesting that the structure of lignin changed. This result is consistent with the composition analysis and SEM observation. After pretreatment, the transmittance

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**Fig. 1.** Time course of chemical composition of wheat straw treated at different concentration of NaOH. Changes of cellulose a), hemicellulose b), lignin c) and ash d) in wheat straw at different pretreatment conditions. Composition is on the basis of dry matter of raw wheat straw. The legend in a) applies to b), c) and d).

**Fig. 2.** Comparison of the SEM images of raw wheat straw a) and the pretreated wheat straw by 4%, 6%, 8% NaOH for 8 days (b, c, d), respectively.

**Fig. 3.** Comparison of the FT-IR spectra of raw wheat straw a) and the pretreated wheat straw by 4%, 6%, 8% NaOH for 8 days (b, c, d), respectively.
of bands, like 3,412 cm\(^{-1}\) (H-bonded O-H stretching) and 990 cm\(^{-1}\) (C-H deformation) have some increase because the content of these groups in substrate increased, reflecting the depolymerization of lignocellulose polymers and disruption of the crystalline structure [28-29].

Loss of Total Organic Carbon During Pretreatment

The changes of TOC at different NaOH concentrations and different pretreatment times are presented in Fig. 4. It is clear that both purified water and NaOH pretreatment caused TOC loss. Seven-hundred milligrams of TOC (based on 65 g substrate) were lost after two days pretreated by purified water and no more than 3 g TOC

Fig. 4. Lost TOC in NaOH pretreatment. Y-axis means the lost TOC comparing with the mass of substrate, g/65g.

Fig. 5. Enzymatic digestion of wheat straw and loss ratio of TOC. Samples for enzymatic hydrolysis were taken after 2 a), 5 b) and 8 days c), Y-axis means percentage that the outcome of enzymatic hydrolysis to total mass; the legend in a) applies to b) and c). The lost TOC ratios were also described d).
were lost after eight days. This lost TOC might be from some low-weight matter, like organic acid. Compared with pure water pretreatment, the content of TOC in NaOH pretreatment liquid substantially increased. The lost TOC was 4 g after two days pretreatment by 4% NaOH, and doubled after eight days. During the first five days, 6% and 8% NaOH pretreatment caused a close and stable loss, which was about 6 to 7 g, but the TOC loss had a sharp increase after eight days. It was also found that after eight days, the TOC loss was 8 g from the wheat straw pretreated by 4% NaOH while increasing to 13 g when pretreated by 8% NaOH. The results indicate that the NaOH concentration and pretreatment time had great effects on wheat straw.

### Comparison of Enzymatic Digestion and Loss Ratio of TOC

Enzymatic digestion experiments were carried out for 48 h at 45°C, which is regarded as optimal [30]. The results are shown in Figs 5a-5c.

The enzymatic digestion of substrates pretreated by purified water and raw material had very low hydrolysis, which remained at around 4% after 48 h. The enzyme attacked the target with difficulty because most hemicellulose and cellulose were covered by lignin. However, after pretreatment enzymatic digestion appeared to increase distinctly. The maximal enzymatic digestion of the substrates taken after two and five days was 19.7% with 6% NaOH and 19.5% with 4% NaOH, respectively (Figs 5a-5b). After pretreatment for eight days, 8% NaOH resulted in a maximum of 22% enzymatic digestion for 48 h and the percentage would continue to rise (Fig. 5c). This led us to assume that 8% NaOH pretreatment at 35°C could generate the largest enzymatic yield in enough enzymatic time.

However, even though NaOH pretreatment resulted in prime enzymatic digestion, unexpected loss of TOC (Eq. 5) was aggravated in long-term pretreatment with such alkali concentrations (Figure 5d). After two days of pretreatment, the most serious loss ratio of TOC was 10%, while this value reached 20% after eight days with 8% NaOH pretreatment. The lost organic matters were some low-weight molecules that could easily be converted into biofuel by fermentation. Therefore, it is not economic and causes a huge waste of carbon resource if organic matter is lost in pretreatment. Unfortunately, the loss of organic matter in the pretreatment process is not given as much importance. It is worth thinking about how to control the balance between the loss of TOC and enhancement of hydrolysis.

### Conclusion

NaOH pretreatment can change the composition and structure of wheat straw, even at a mesopholic temperature. The lignin has been removed effectively after NaOH pretreatment. Compared with purified water pretreatment and non-pretreatment, NaOH pretreatment exhibits good performance on enhancing enzymatic hydrolysis. But at the same time, a large ratio of TOC loss is caused. So the pretreatment conditions should be carefully chosen to ensure the treatment effect while reducing organic loss. The evaluation criteria of a pretreatment method should be included with comprehensive factors such as TOC loss, hydrolytic yield, process complexity, treatment cost, environmental friendliness, etc.

### Acknowledgements

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