Characteristics of Kitchen Waste and the Formation of Floating Brown Particles (FBP) in the Anaerobic Digestion Process

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Abstract

Five types of kitchen waste (KW) from China – including hop pot (HP), fast food (FF), Hebei cuisine (HC), university canteen (UC), and other mixed KW (Other) – were investigated as feedstock for potential biogas and methane production. The biodegradability of KW was measured using batch anaerobic digestion (AD) tests and feedstock at an inoculum ratio (F/I) of 0.5. Gompertz and Cone models were used to determine the kinetic parameters of KW degradation, biogas, and methane production. Results showed that HP had the highest lag phase time of 5.46 days. Methane production varies with different sources of KW. HP had the highest methane yield of 363.9 mL/g-VS added as compared to a sample of FF (334.8 mL/g-VS added), other (278.5 mL/g-VS added), UC (239.2 mL/g-VS added), and HC (236.0 mL/g-VS added). The biodegradability of KW ranged from 39.5% to 50.4%. During the AD process a certain amount of floating brown particles (FBP) were formed, which may be the main inhibiting factor of methane production. Analysis of 13C NMR and FTIR revealed that the main component of FBP was calcium stearate. The formation mechanism of calcium stearate may contribute to the relatively high lipid content (18.6% to 30.9%) of the KW sample, which subsequently resulted in over-accumulation of long-chain fatty acids (LCFAs) and reaction with Ca2+. Using lipid-rich substrates as feedstock may be an efficient approach to adding Ca2+ artificially for reducing the inhibition of LCFAs.

Keywords: kitchen waste, biodegradability, floating brown particles, anaerobic digestion, calcium stearate
Introduction

With the growth of population and development of the worldwide economy, kitchen waste (KW), including uneaten food and food leftovers from restaurants, households, canteens, and companies, is increasingly produced. As the world’s most populous country, China produces approximate 90 million tons of KW each year [1]. Beijing, the second largest Chinese city by urban area, produces 1,700 tons of KW daily [2]. The uncontrolled discharge of large amounts of KW causes severe environmental problems such as land pollution, odors, and greenhouse gases emissions [3]. Traditional approaches for KW disposal are mainly landfills, incineration, and aerobic composting. In term of eco-friendly and cost-effective measures, anaerobic digestion (AD) is a promising method that has been increasingly studied. Additionally, KW has high moisture content of 74-90%, which is more adaptable for AD compared to conventional technologies. In the AD process, organics are gradually hydrolyzed to small molecular materials and soluble organic substrates (e.g., acetate), which could be used by methanogens for methane production.

Due to complex constituents in Chinese KW, the biogas production performance varies with different sources of KW [4]. Lipid-rich waste contributes a considerable amount to the total quantity of KW. Theoretically, the methane potential of lipids is 1,014 L/kg-VS added, a value higher than of carbohydrates and protein (e.g., 415 L/kg-VS added for glucose and 496 L/kg-VS added for protein) [5]. The actual methane production, however, is unsatisfactory due to the inhibition of main intermediate by-products (e.g., long-chain fatty acids or LCFAs) derived from the lipid degradation process as well as mass transfer problems for soluble substrates as they become adsorbed to the microbial biomass surface [6]. Later studies demonstrated that samples with higher lipid content produced more methane with relative extensions of initial lag phase time [7]. In the current work, the phenomenon of “floating brown particles” (FBP) appeared during AD of KW. To find the composition of FBP, an energy dispersive x-ray analyzer, 13C nuclear magnetic resonance (NMR), and a Fourier transform infrared spectrometer (FTIR) were used. The main objectives of this study were: 1) to investigate the effect of different KW sources on biogas and methane production performance and 2) to find the composition and formation mechanism of FBPs.

Methods

Feedstock and Inoculum

Five kinds of kitchen waste (KW) were sampled from different places (Table 1). Each kind of KW was obtained from mixing two or three different samples collected from the same type of place. Once collected, KW was shredded into particles with an average size of 3.0 mm and kept at -18°C for later use. Digested sludge from a stable operated CSTR reactor of Asuwei Landfill (Changping District, Beijing, China) was selected as inoculum. Prior to inoculation, inoculum was acclimated (pre-incubation) for two weeks in a constant temperature incubator shaker (ZWYR-D2402, China) by adding mixed KW at 42±1°C [8-9]. Unpublished data from biogas plant surveys suggest that average temperatures of mesophilic (35-45°C) digesters of full-scale biogas plants was 40°C, therefore the performance of trials at 42°C seems legitimate to us. After acclimatization, the total solids (TS), volatile solids (VS), and pH of inoculum was determined to be 2.02%, 0.88%, and 7.17, respectively. As previous research suggested that thermo-pretreatment could reduce the content of oil and accelerate the hydrolysis of KW [10], KW used in this study was treated by a high-temperature sterilization pot (Panasonic MLS-3751L) at 120°C for two hours. Processed samples were then kept at 4°C before use.

Batch Experiments

Biochemical methane potential (BMP) tests were conducted in batch mode using glass bottles with a total volume of 250 mL and a working volume of 200 mL. The initial VS concentration of KW was 6.67 g-VS/L and the corresponding feedstock-to-inoculum (F/I) ratio was adjusted to 0.5 (on VS basis) [11]. After the addition of inoculum and substrates, tap water was added to digesters to make a final working volume of 200 mL. To ensure an oxygen-free environment, nitrogen was continuously passed though the loaded glass bottles for two minutes. Digesters were then placed in a constant temperature incubator shaker (ZWYR-D2402, China) for three weeks. The digestion temperature was 42±1°C and shaking rate was 100 rpm. Control samples were set up by adding the same amount of inoculum and water instead of KW.

Analytical Methods

TS and VS were determined according to standard methods [12]. Major elements of organic of the KW samples (such as C, H, N, and S) were measured by an elemental analyzer (EA3000, Italy). The contents of Fe, Cu, Mn, Zn, Ca, and Mg of the KW samples were analyzed by means of x-ray fluorescence (Panalytical, Petro-Axiosm AX, Italy). The content of Ca, and Mg of the KW samples were analyzed by means of x-ray fluorescence (Panalytical, Petro-Axiosm AX, Italy). The content of Ca, and Mg of the KW samples were analyzed by means of x-ray fluorescence (Panalytical, Petro-Axiosm AX, Italy).
Characteristics of Kitchen Waste...

Hemlock, cellulose, hemicellulose, and lignin were tested by an ANKOM fiber analyzer (A200, America). Crude fat was measured according to the Soxhlet extraction method with a crude fat determination instrument (SZF-06G, China). Crude protein was determined by a Kjeldahl nitrogen instrument (KDY-9820, China).

Biogas production was measured once per day using an ANKOM RFS gas production system. Biogas composition was analyzed once per day using a gas chromatograph (FULI 9790II, China) equipped with a thermal conductivity detector and helium as the carrier gas. The temperatures of the injector, oven, and detector were 150, 130, and 160°C, respectively.

After digestion, floating brown particles (FBP) were formed in all digesters. To investigate the composition of FBP, an energy-dispersive x-ray analyzer (EX-350, HORIBA EMAX Energy, Japan) was used under the test conditions of 15 kV. The images were taken using a scanning electron microscope (SEM) at 10,000 and 30,000 magnification. The functional groups of FBP were analyzed using an FTIR spectrometer (Bruker vertex 70v, Germany). The solid samples were mixed with KBr, ground to a fine powder, and pressed into pellets for infrared (IR) transmission studies. Spectral information was collected over a wave number range from 4000 to 400 cm⁻¹. Furthermore, the functional groups of FBP were analyzed by ¹³C nuclear magnetic resonance (NMR).

**Kinetic Models**

First-order reaction constant ($k$) is a vital parameter for predicting the feasibility of using feedstocks for large-scale biogas production. Besides, the lag phase time ($\lambda$) is also important for designing an AD plant. In this study, modified Gompertz (Eq. (1)) and Cone (Eq. (2)) models were used to determine the $k$ and $\lambda$ values [13].

$$B = B_0 \exp \left( -\exp \left( \frac{\mu_m \tau}{B_0} (\lambda - t) + 1 \right) \right)$$

(1)

$$B = \frac{g}{1 + (kt)^{-n}}$$

(2)

**Table 2. Elemental composition of KW examined in this study and in literature.**

<table>
<thead>
<tr>
<th>Parameters</th>
<th>HP</th>
<th>HC</th>
<th>FF</th>
<th>UC</th>
<th>Other</th>
</tr>
</thead>
<tbody>
<tr>
<td>TS, %*</td>
<td>14.5±1.14</td>
<td>19.35±1.29</td>
<td>20.02±2.22</td>
<td>16.85±2.86</td>
<td>23.37±1.60</td>
</tr>
<tr>
<td>VS, %*</td>
<td>13.09±1.17</td>
<td>18.03±1.71</td>
<td>18.72±2.23</td>
<td>15.31±3.09</td>
<td>21.45±1.37</td>
</tr>
<tr>
<td>VS/TS, %*</td>
<td>89.84±1.28</td>
<td>93.13±2.65</td>
<td>93.50±0.75</td>
<td>90.59±2.95</td>
<td>91.80±1.12</td>
</tr>
<tr>
<td>C, %TS</td>
<td>55.80±3.83</td>
<td>52.31±0.08</td>
<td>53.03±5.92</td>
<td>49.95±5.55</td>
<td>56.98±1.86</td>
</tr>
<tr>
<td>N, %TS</td>
<td>2.71±0.10</td>
<td>2.74±0.40</td>
<td>1.99±0.02</td>
<td>2.87±0.11</td>
<td>3.48±0.10</td>
</tr>
<tr>
<td>C/N, %</td>
<td>20.59±1.74</td>
<td>19.33±2.82</td>
<td>26.69±2.71</td>
<td>17.46±2.62</td>
<td>16.39±1.00</td>
</tr>
<tr>
<td>H, %TS</td>
<td>8.35±0.43</td>
<td>7.98±0.02</td>
<td>8.25±0.97</td>
<td>7.60±0.99</td>
<td>8.10±0.84</td>
</tr>
<tr>
<td>O, %TS</td>
<td>23.90±4.17</td>
<td>28.80±1.23</td>
<td>30.23±5.81</td>
<td>30.87±4.85</td>
<td>24.61±2.97</td>
</tr>
<tr>
<td>S, %TS</td>
<td>0.31±0.39</td>
<td>0.10±0.04</td>
<td>0.08±0.02</td>
<td>0.30±0.14</td>
<td>0.18±0.07</td>
</tr>
<tr>
<td>Cl, %TS</td>
<td>7.39±0.72</td>
<td>7.10±0.13</td>
<td>8.03±2.97</td>
<td>8.11±3.40</td>
<td>6.60±0.89</td>
</tr>
<tr>
<td>P, %TS</td>
<td>0.68±0.13</td>
<td>0.66±0.16</td>
<td>0.84±0.69</td>
<td>0.39±0.05</td>
<td>0.75±0.17</td>
</tr>
<tr>
<td>K, %TS</td>
<td>2.82±0.14</td>
<td>2.65±0.53</td>
<td>2.22±0.72</td>
<td>1.82±0.28</td>
<td>2.14±0.33</td>
</tr>
<tr>
<td>Ca, %TS</td>
<td>3.94±0.94</td>
<td>3.75±1.69</td>
<td>2.74±1.05</td>
<td>3.18±0.17</td>
<td>3.83±0.50</td>
</tr>
<tr>
<td>Na, %TS</td>
<td>2.98±0.09</td>
<td>2.97±0.03</td>
<td>3.49±2.53</td>
<td>3.31±2.09</td>
<td>2.23±0.69</td>
</tr>
<tr>
<td>Mg, %TS</td>
<td>0.18±0.02</td>
<td>0.16±0.01</td>
<td>0.18±0.13</td>
<td>0.12±0.04</td>
<td>0.12±0.03</td>
</tr>
<tr>
<td>Al, %TS</td>
<td>0.03±0.01</td>
<td>0.02±0.01</td>
<td>0.04±0.01</td>
<td>0.04±0.02</td>
<td>0.02±0.01</td>
</tr>
<tr>
<td>Zn, %TS</td>
<td>0.02±0.00</td>
<td>0.02±0.00</td>
<td>0.02±0.00</td>
<td>0.01±0.00</td>
<td>0.02±0.00</td>
</tr>
<tr>
<td>Cu, %TS</td>
<td>0.02±0.01</td>
<td>0.01±0.00</td>
<td>0.01±0.01</td>
<td>0.01±0.00</td>
<td>0.01±0.00</td>
</tr>
<tr>
<td>Mn, %TS</td>
<td>0.01±0.01</td>
<td>&lt;0.01</td>
<td>0.01±0.01</td>
<td>&lt;0.01</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Fe, %TS</td>
<td>0.07±0.02</td>
<td>0.08±0.00</td>
<td>0.09±0.03</td>
<td>0.06±0.03</td>
<td>0.06±0.01</td>
</tr>
<tr>
<td>Ni, %TS</td>
<td>&lt;0.01</td>
<td>&lt;0.01</td>
<td>&lt;0.01</td>
<td>&lt;0.01</td>
<td>&lt;0.01</td>
</tr>
</tbody>
</table>

* Treatment by high-temperature sterilization pot at 120°C for two hours plus removal of lipids. * “O” was calculated based on Eq.: 99.5%VS = C + H + O + N. * Based on wet basis, NG: not given.
... where $B$ is the cumulative methane yield (mL/g-VS$_{added}$), $B_0$ is the ultimate methane yield (mL/g-VS$_{added}$), $\lambda$ refers to the lag phase time (d), $\mu_{max}$ stand for maximum methane production rate (mL/g-VS$_{added}$/d), $e$ is equal to 2.718281828, $t$ is the digestion time (d), $k$ is the first-order rate constant (d$^{-1}$), and $n$ is the shape factor (dimensionless).

## Results and Discussion

### Characteristics of Substrates

Characteristics of KW from different sources are shown in Table 2. The VS/TS ratio ranged from 89.8% to 93.5% in all KW samples, which is desirable for biomethane production as a higher VS/TS ratio indicates higher organic content [14]. On a total weight basis, KW from the canteen showed the lowest TS and VS contents. The C/N ratios of HP, HC, FF, UC, and Other were 20.6, 19.3, 26.7, 17.5, and 16.4, respectively, which is suitable for methane production as previous research suggested that a proper C/N ratio for AD ranged from 15.0 to 30.0 [15]. Compared to previous research (Table 2), KW contains relatively high moisture (>70%), high VS/TS ratio (>85%), and suitable C/N ratio (14.8-26.7).

Besides, abundant nutrient elements were detected in all KW samples (Table 2). Among them, elements of P (0.39-0.84%), K (1.82-2.82%), Na (2.23-3.49%), and Ca (2.74-3.94%) (on TS basis) are essential for microbial growth. Trace metal elements of Al, Zn, Cu, Mn, and Fe were also detected. Metal concentration is a vital parameter in methane formation as bioavailable trace metals could stimulate the conversion of pollutants into methane and improve the tolerance of microorganisms to toxic substances [18-19]. But metal concentrations should not exceed the allowable standards [20-21].

### Biomethane Potential

Biogas production from five different sources of KW is shown in Fig. 1. After 21 days digestion, HP produced the highest methane yield at 363.9 ±38.9 mL/g-VS$_{added}$ among all the samples, followed by FF (334.8 mL/g-VS$_{added}$), Other (278.5 mL/g-VS$_{added}$), UC (239.2 mL/g-VS$_{added}$), and HC (236.0 mL/g-VS$_{added}$). Large standard deviations were found in BMP tests due to the sampling method, indicating that sample sources significantly influenced the biomethane potential of KW. It is worth noting that HP had the longest lag phase compared to other samples (Figs 1-2). This may be related to the relatively high lipid content of HP (Fig. 3), as previous research demonstrated that the sample with higher lipid content produced more methane with relative extension of the lag phase [22]. The lipid content in KW ranged from 18.6% to 30.6%. The daily methane generation of HP increased dramatically from day 4 and reached peak production of 43.9 ±10.7 mL/g-VS$_{added}$ on day 11 (Fig. 2). Daily methane production of FF and HC increased gradually from day 1 and reached the peak of 32.4 ±27.4 mL/g-VS$_{added}$ and 38.4 ±5.2 mL/g-VS$_{added}$ on day 10, respectively (Fig. 2), which is relatively earlier than HP. According to methane production results, HP had the highest methane production potential.

As can be seen from Fig. 3, theoretical methane yields of different sources of KW ranged from 605.8 mL/g-VS$_{added}$ to 721.4 mL/g-VS$_{added}$ by using elemental and organic composition analysis methods [5]. Thus, the biodegradability of KW was found to be 39.5-50.4%. The methane yields and biodegradability obtained in this study was relatively low as compared to the values reported by Zhang et al. [4], where methane yield was 435 mL/g-VS$_{added}$ and VS destruction was 81%. It is worth noting that a certain amount of floating brown particles (FBP) formed during the AD process. In this research, further analysis was performed to investigate the physicochemical property of floating brown particles (data is shown in 3.4).

### Kinetic Models

Modified Gompertz and Cone models were used in this study for describing and predicting AD processes [13]. The calculated parameters are shown in Table 3. For the Cone model, the determination coefficient ($R^2$) ranged
from 0.994 to 0.999. For the modified Gompertz model, \( R^2 \) ranged from 0.992 to 0.998. Higher \( R^2 \) indicated that the assumption of Cone and modified Gompertz models can suit AD tests well. Based on the Cone results, the hydrolysis rate constants \( (k) \) for HP, HC, FF, UC, and Other were 0.098, 0.124, 0.099, 0.132, and 0.112 d\(^{-1}\), respectively, which were comparable with values of 0.050-0.240 d\(^{-1}\) and 0.127-0.193 d\(^{-1}\) as reported by Li et al. [5] and Qiao et al. [23]. According to the results of modified Gompertz, KW from HP showed the highest lag phase time \( (\lambda) \) of 5.46 d, which indicates the longest startup occurring in the HP digester. It is worth noting that HP contains the highest content of lipids (30.9% of TS, Fig. 3), while the degradation rate of lipids is slow for microorganisms. Maximum methane production rate \( (\mu_m) \) and ultimate methane yield \( (B_0) \) were found in HP, which may also be due to the contribution of lipids.

Analysis of Floating Brown Particles

**SEM and Energy Spectrum Analysis**

The scanning electron micrograph (SEM) showed that the floating brown particles have multi-layer structure (Fig. 4), which may form superimposed “particles.” As shown in Fig. 5, During AD, FBP were found at the centimeter level. Energy spectrum analysis showed that the floating brown particles mainly contained three elements: C, O, and Ca. The mass fractions of C, O, and Ca in the brown particles are 67.49%, 26.78%, and 4.28%, respectively, and the atomic numbers are 75.40%, 22.46%, and 1.43%, respectively. The analysis revealed organic matter composition of the floating brown particles.

\[ ^{13}C \text{ NMR and FTIR Analysis} \]

\(^{13}C\) NMR profiles of the samples indicated that the floating brown particle sample has different contents of C-containing groups (Fig. 6). According to resonance signals in \(^{13}C\) NMR spectra and attributed chemicals, groups appearing in the region of 0-50 ppm are either groups of methylene (–CH\(_2\)) or methyl (–CH\(_3\)); the peak at 179.30×10\(^{-6}\) is carboxylic acid radical anion group (–COO\(^{-}\)). Based on analysis results of \(^{13}C\) NMR and FTIR, the sample is mainly composed of three groups: methyle (–CH\(_2\)), methyl (–CH\(_3\)), and carboxylate anion groups (–COO\(^{-}\)). Consequently, the component of the sample can be inferred as carboxylic acid salts of saturated chain structure [CH\(_x\) (–CH\(_2\))\(_n\)COO\(^{-}\)]. An FTIR spectrogram showed that the number of methylene was

**Table 3. Parameters of Cone and modified Gompertz models for different types of KW.**

<table>
<thead>
<tr>
<th>Substrates</th>
<th>Cone model</th>
<th>Modified Gompertz model</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>( B_0 ) (mL-CH(<em>4)/g-VS(</em>{added}))</td>
<td>( \mu_m ) (mL-g-VS(_{added})/d)</td>
</tr>
<tr>
<td>HP</td>
<td>387.2±4.8</td>
<td>0.098±0.001</td>
</tr>
<tr>
<td>HC</td>
<td>245.9±3.6</td>
<td>0.124±0.002</td>
</tr>
<tr>
<td>FF</td>
<td>360.9±2.0</td>
<td>0.099±0.001</td>
</tr>
<tr>
<td>UC</td>
<td>253.1±5.2</td>
<td>0.132±0.003</td>
</tr>
<tr>
<td>Other</td>
<td>305.5±4.5</td>
<td>0.112±0.002</td>
</tr>
</tbody>
</table>
As “Ca” was the main element (2.74-3.94% on TS basis) in the floating brown particle, the component of the sample was likely to be calcium stearate. FTIR spectroscopy comparison further confirmed the similarity composition of floating brown particles and calcium stearate (Fig. 7). Although the absorption of the brown particles and the calcium stearate is slightly different, the peak position is basically the same.

Lipids were hydrolyzed by extracellular lipase generating glycerol and LCFAs (such as hexadecanoic acid and stearic acid). During the hydrolyzing process, parts of the saturated fatty acid reacted with Ca2+ and formed calcium stearate [25]. The accumulation of calcium stearate resulted in agglomeration and formed floating granules. This reaction leads to a decrease in the accumulation of LCFAs, which subsequently reduced the inhibition effect on AD reaction and methane production. For AD of lipid-rich substrates, to reduce the inhibition of LCFAs, Ca2+ can be added into the digester. However, the decrease of intermediates (LCFAs) on the other hand also results in insufficient bioavailable nutrients, which restrict methane production.

**Conclusions**

The present study investigated the potential of biogas and methane generation from different sources of KW collected from China. Results indicated that methane production vary with different sources of KW. Compared to FF, HC UC, and other sources of KW,
HP showed the highest methane production potential of 363.9±38.9 mL/g-VS added, which may be due to its relatively high lipid content. During anaerobic digestion, a certain amount of floating brown particles (FBP) were observed. Combined with energy spectrum, SEM, 13C NMR, and FTIR analysis, it can be inferred that the brown particles are likely to be calcium stearate. The formation mechanism of calcium stearate may contribute to the reaction between accumulated LCFA and Ca2+. The formation of the particles on the one hand reduced the toxicity of LCFA accumulation, whereas on the other side it may play a role in the inhibition of biogas output. Based on the results, when using lipid-rich substrates as feedstock, it may be an efficient approach to add Ca2+ artificially to reduce the inhibition of LCFA. Meanwhile, the biodegradability of floating brown particles needs to be further investigated.

Acknowledgements

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References

14. NELSON R. Methane generation from anaerobic digesters: considering different substrates Environmental Biotechnology-Iowa State University, USA, 2010.
18. YIRONG C., HEAVEN S., BANKS C.J. Effect of a trace element addition strategy on volatile fatty acid accumulation in thermophilic anaerobic digestion of food waste. Waste Biomass Valor. 6 (1), 1, 2015.