

Short Communication

# Chemical Composition of Volatile Oil from Two Emergent Plants and Their Algae Inhibition Activity

Hong-Qiang Wang<sup>1,2</sup>, Feng Liang<sup>1</sup>, Ning Qiao<sup>1</sup>, Jin-Xia Dong<sup>1</sup>,  
Lie-Yu Zhang<sup>3\*</sup>, Yi-Fei Guo<sup>1</sup>

<sup>1</sup>School of Municipal and Environmental Engineering, Henan University of Urban Construction,  
Pingdingshan, Henan province 467036, PR China

<sup>2</sup>State Key Laboratory of Freshwater Ecology and Biotechnology, Institute of Hydrobiology,  
Chinese Academy of Sciences, Wuhan, Hubei Province 430072, PR China

<sup>3</sup>Chinese Research Academy of Environmental Sciences, Beijing 100012, PR China

Received: 3 November 2013

Accepted: 10 March 2014

## Abstract

The volatile oils of two emergent plants (*Typha latifolia* and *Arundo donax*) were extracted with alcohol and then fractionated with some chemical reagent. Chemical composition of volatile oils were analyzed and identified by gas chromatograph-mass spectrometry (GC-MS). The volatiles appeared to form a complex mixture of 27 identified constituents, containing mainly fatty acids, ester, aldehyde, sterol, ketone, phenol, etc. Allelopathic activities of volatile oils on *Microcystis aeruginosa* were also determined. The results indicated that volatile oils from two emergent plants inhibited the growth of *M. aeruginosa* with inhibition rates of 43.3% and 47.9%, respectively, when the concentration of extracts was 50.0 mg/L.

**Keywords:** allelopathic activity, volatile oil, *Typha latifolia*, *Arundo donax*, *Microcystis aeruginosa*

## Introduction

Eutrophication has been a major water quality problem all over the world [1]. During the past decades many efforts have been made to combat eutrophication of shallow lakes by various measures. Recent studies have demonstrated that macrophytes with allelopathic potential may play an important role in the restoration of eutrophic lakes [2]. In other words, the growth of harmful algal blooms can be controlled by using allelochemical excreted from some macrophytes [3, 4]. Allelopathy has been described for many macrophytes, such as *Phragmites communis* [5], *Vallisneria spiralis* [6], *Myriophyllum spicatum* [7], *Stratiotes aloides* [8], and *Potamogeton maackianus* [9, 10]. Macrophytes can

produce a wide variety of secondary metabolites such as polyphenols, terpenes, ester, fatty acid, and ketone to effectively inhibit the growth of blue-green algae. Volatile organic compounds such as major allelochemicals in some aquatic plants have been reported. For example, dihydroactinidinolide from *Vallisneria spiralis* showed inhibition activity against *Microcystis aeruginosa* [6]. Shao et al. [11] also showed that  $\beta$ -ionone had strong inhibitory effects on *M. aeruginosa*. Ozaki et al. [12] reported that volatile organic compounds have lytic effects on *Microcystis*. In addition, Harada et al. [13] also found that  $\beta$ -ionone can cause cellular lysis and a decrease of Chl *a* content on *Microcystis*. It is known that cellular lysis and a decrease of Chl *a* are common phenomena of algal cell death [11].

Release of volatile organic compounds can help plants acclimatize to abiotic stress, contributing to thermotoler-

---

\*e-mail: zhanglieyu@163.com

ance and the removal of reactive oxygen species, and there is a growing appreciation that chemical emissions may have wider importance for plant biology [14].

These results indicate that algal growth could be controlled by using some macrophytes with allelopathic potential. The objective of this study was to investigate the components of the volatile oils from *Typha latifolia* and *Arundo donax* as well as their allelopathic effect on the growth of *M. aeruginosa*.

## Materials and Methods

### Chemicals

Anhydrous sodium sulfate (AR), alcohol (AR), diethyl ether (AR), and dimethyl sulfoxide (DMSO, AR) were purchased from Chemical Reagent Company, Shanghai, China. Methanol (HPLC) was purchased from Tedia Company, INC, USA.

### Macrophytes and Extraction of Chemicals

*T. latifolia* and *A. donax* were collected from Pingxi Lake, Pingdingshan, China. Plant materials were washed free of debris with regular water and later by deionized water, then dried and powdered. An appropriate amount of the powdered sample was soaked with alcohol for 72 h at room temperature, then filtered with GF/C glass fibre filters (47 mm, 1.2  $\mu$ m, purchased from Whatman Maidstone, UK) with reducing pressure using a vacuum pump, subsequently collected the filtrates. The alcohol filtrates were evaporated to be close to dryness by rotary evaporator at 39°C, then appropriate amounts of ultrapure water were added to the alcohol filtrates, finally fractionated with diethyl ether three times. The diethyl ether filtrates were first dried with anhydrous sodium sulfate and then evaporated to dryness by rotary evaporator at 39°C. The diethyl ether extracts were stored at 4°C until being used for GC-MS analysis and biological assay.

### Bioassay

Axenic *M. aeruginosa* were obtained from the Culture Collection of Algae at the Institute of Hydrobiology, Chinese Academy of Sciences. The algae were cultured in sterilized BG11 medium (pH 7.4) at 25°C and light intensity of 2500 lux, 12:12 h light:dark cycle. The algae were cultured for 4 days to reach the exponential phase with the density of  $10^5$ - $10^6$  cells/mL, which were used for the assay of growth inhibition. The growth medium of all cultures was BG11 [15].

The concentration-response relationships between the allelochemicals and the tested organisms were studied in 50 mL flasks containing 20 mL test solution, to which  $10^6$  cells·mL<sup>-1</sup> of *M. aeruginosa* were inoculated. The tested organisms were exposed, in triplicate, to one concentration level and a control, respectively. The final concentrations of compounds in the test solution were 50 mgL<sup>-1</sup> for volatile oils

Table 1. Composition of the volatile oil from *T. latifolia* and *A. donax*.

Rent time (min)	Compounds	<i>T. latifolia</i>	<i>A. donax</i>
7.31	2-metoxo-4-vinyl-phenol		
7.54	4-hydroxybenzaldehyde		
7.90	vanillin		
8.25	dimethyl phthalate		
8.79	lauric acid	–	
9.11	3,4,5-trimethoxyphenol	–	
9.5	syringaldehyde	–	
9.95	myristic acid		–
10.24	phytane	–	
10.37	P-hydroxyl ethyl cinnamate		–
10.41	3,7-dimethyloct-6-en-1-yl acetate		–
10.45	6,10,14-trimethyl-2-pentadecanone		–
11.01	hexadecanoic acid		
11.10	dibutyl phthalate		
11.18	palmitic acid ethyl ester		
11.78	phytol		–
11.86	linoleic acid		
11.96	stearic acid		–
11.99	ethyl linoleate		–
12.03	linolenic acid ethyl ester		
12.11	ethyl stearate		
12.89	4,8,12,16-tetramethylheptadecan-4-olide		–
13.68	unknown		–
21.84	campesterol	–	
22.51	stigmasterol	–	
23.85	$\gamma$ -sitosterol	–	
24.07	stigmastanol	–	
27.58	stigmast-4-ene-3-one		

“–” undetectable

from *T. latifolia* and *A. donax*. The 50% inhibition concentration of every volatile oil based on cell density of the tested organisms (EC<sub>50</sub>) were determined after exposure for 72 h. The stock solutions of volatile oils were prepared with DMSO, which in test solution was lower than 0.2% (v/v). The test results indicated that the concentrations of DMSO added had no effect on the growth of the tested organisms.

Table 2. Inhibitory effects of the volatile oil from *T. latifolia* and *A. donax* on growth of *M. aeruginosa*.

	<i>T. latifolia</i>	<i>A. donax</i>
Inhibition ratio	43.3%	47.9%

### Identification of Organic Acid

The dried volatile oils were analyzed by gas chromatograph-mass spectrometry (GC-MS) (Agilent computerized system consisting of a 6890 gas chromatograph coupled to a Agilent 5973N quadrupole mass spectrometer) using an HP-5MS capillary column (30 m × 0.25 mm × 0.25 mm id). The oven temperature was programmed at 60°C (initial temperature maintained 1 min) to reach 260°C at a rate of 4°C/min, kept constant 20 min. Helium was used as a carrier gas with a flow rate of 1 mL/min. Mass fragments of the components were compared to the mass fragmentation data contained in the NIST 02.

## Result and Discussion

### Composition of the Volatile Oil from *T. latifolia* and *A. donax*

The components of the volatile oils from *T. latifolia* and *A. donax* were listed in Table 1. 27 compounds were analyzed and identified by GC-MS, in which fatty acids, ester, aldehyde, sterol were primary compositions. Compared components of the volatile oils from *T. latifolia* and *A. donax*, the amount and components of volatile oils were different in two macrophytes. There were differences in the amount and components of volatile oil, which might be due to the differences of plant species.

### Allelopathic Effects of Volatile Oils on Algae

The allelopathic effects of volatile oils from *T. latifolia* and *A. donax* were explored by bioassay of *M. aeruginosa*. The results showed that volatile oils from these plants inhibited the growth of *M. aeruginosa* with inhibition rates of 43.3% and 47.9%, respectively, when the concentration of extracts was 50.0 mg/L.

The role of allelopathy in aquatic systems has received increasing attention as a potential means of controlling algal blooms [2]. Yang et al. [16] reported that vanillin at a concentration of 40 mg/L had obvious inhibition on the growth of *Alexandrium tamarense*. Five allelopathic compounds (lauric acid, myristic acid, hexadecanoic acid, linoleic acid, stearic acid) can inhibit the growth of *M. aeruginosa*, *Chlorella pyrenoidosa* Chick, and *Scenedesmus obliquus* Kütz [17] and *Selenastrum capricornutum* [18] (Table 3). These compounds were also found in *T. latifolia* and *A. donax*.

*T. angustifolia* had a strong allelopathic effect on *Bolboschoenus fluviatilis* (river bulrush), reducing the longest leaf length and root, shoot, and total biomass of *B. fluviatilis* [20]. Aliotta et al. [19] reported that *T. latifolia* contains six compounds ( $\beta$ -sitosterol, (20s)24-methylphenol, stigmast-4-ene-3,6-dione,  $\alpha$ -linolenic, linoleic, and an unidentified C18:2) with inhibitory activity to the growth of some microalgae. Compared to the results reported by Aliotta et al. [19], linoleic acid and  $\gamma$ -sitosterol were detected. Other compounds did not find it in the study. There was a difference in components of compounds, which might be due to the differences of research methods.

Hong and Hu [21] reported the effects of aquatic extract of *Arundo donax* Linn. (*A. donax*) on the growth of four freshwater algae (*M. aeruginosa*, *Selenastrum capricornutum*, *S. obliquus*, and *C. pyrenoidosa*) to control algal blooms. Hong et al. [22] isolated and identified gramine (N,N-dimethyl-3-amino-methylindole) from *Arundo donax*, gramine with 0.47 mg·L<sup>-1</sup> of medium effective concentration to inhibit the growth of *M. aeruginosa*. After gramine exposure, esterase and total dehydrogenase activities were increased firstly but decreased later. In contrast with the controls, the cells exposed to gramine showed apparent ultrastructure alterations with thylakoids in breakage, phyco-bilins in decrease; lipid and cyanophycin granules were abundant at first but dissolved afterwards, DNA in fragmentation [23]. The results indicated that the allelopathic effects of extract of *A. donax* L. on algae were strain-specific and *A. donax* L. may control the toxic cyanobacterium *M. aeruginosa*. Based on the observation of algal morphology and the measurement of algal density and cell size, the results showed that allelochemicals extracted with all three solvents inhibited *M. aeruginosa*. These results indicate that the production of metabolites by *T. latifolia* and *A. donax* might control harmful algal growth.

Table 3. EC<sub>50</sub> (mg/L) of five allelochemicals on toxic *M. aeruginosa*, *C. pyrenoidosa* Chick, *S. obliquus* Kütz (the fifth day), and *S. capricornutum* (the third day).

Compounds	<i>M. aeruginosa</i>	<i>C. pyrenoidosa</i> Chick	<i>S. obliquus</i> Kütz	<i>S. capricornutum</i>
Lauric acid	4.563	6.265	5.813	10.36
Myristic acid	15.498	16.003	15.977	18.65
Hexadecanoic acid	18.234	19.876	20.146	35.46
Linoleic acid	0.047	0.089	0.059	17.27
Stearic acid	19.849	20.849	20.754	40.27
Reference	[17]	[17]	[17]	[18]

## Acknowledgements

This work was kindly supported by the National Science-technology Support Plan Projects of China (2012BAJ21B06), National Natural Science Foundation of China (51108447; 51208485), China Postdoctoral Science Foundation Project (20100471208), China Postdoctoral Science special Foundation (201104499), the open foundation of State Key Laboratory of Freshwater Ecology and Biotechnology (2011FB03) and Science Foundation of Henan University of Urban Construction (2013JBS008).

## References

- DAI Y.R., JIA C.R., LIANG W., HU S.H., WU Z.B. Effects of the submerged macrophyte *Ceratophyllum demersum* L. on restoration of a eutrophic waterbody and its optimal coverage. *Ecol. Eng.* **40**, 113, **2012**.
- PAKDEL F. M., SIM L., BEARDALL J., DAVIS J. Allelopathic inhibition of microalgae by the freshwater stonewort, *Chara australis*, and a submerged angiosperm, *Potamogeton crispus*. *Aquat. Bot.* **110**, 24, **2013**.
- GROSS E.M., MEYER H., SCHILLING G. Release and ecological impact of algicidal hydrolysable polyphenols in *Myriophyllum spicatum*. *Phytochemistry* **41**, (1), 133, **1996**.
- NAKAI S., INOUE Y., HOSOMI M., MURAKAMI A. *Myriophyllum spicatum*-releasing allelopathic polyphenols inhibiting growth blue-green algae *Microcystis aeruginosa*. *Water Res.* **34**, 3026, **2000**.
- LI F.M., HU H.Y. Isolation and characterization of a novel antialgal allelochemical from *Phragmites communis*. *Appl. Environ. Microbiol.* **71**, (11), 6545, **2005**.
- XIAN Q.M., CHEN H.D., LIU H.L., ZOU H.X., YIN D.Q. Isolation and Identification of Antialgal Compounds from the Leaves of *Vallisneria spiralis* L. by Activity-Guided Fractionation. *Environ. Sci. Pollut. Res.* **13**, (4), 233, **2006**.
- LEU E., KRIEGER-LISZKAY A., GOUSSIAS C., GROSS E.M. Polyphenolic allelochemicals from the aquatic angiosperm *Myriophyllum spicatum* inhibit photosystem II. *Plant Physiol.* **130**, (4), 2011, **2002**.
- MULDERIJ G., MOOIJ W.M., SMOLDERS A.J.P., VAN DONK E. Allelopathic inhibition of phytoplankton by exudates from *Stratiotes aloides*. *Aquat. Bot.* **82**, (4), 284, **2005**.
- WANG H.Q., CHENG S.P., ZHANG S.H., HE F., LIANG W., ZHANG L.P., HU C.Y., GE F.J., WU Z.B. Chemical Composition in Aqueous Extracts of *Potamogeton malaianus* and *Potamogeton maackianus* and their Allelopathic Effects on *Microcystis aeruginosa*. *Pol. J. Environ. Stud.* **19**, (1), 213, **2010**.
- ZHANG S.H., CHENG S.P., SUN P.S., WANG H.Q., WU Z.B. Isolation and identification of antialgal compounds from *Potamogeton maackianus* by activity-guided fractionation. *Allelopathy J.* **28**, (1), 95, **2011**.
- SHAO J.H., XU Y., WANG Z.J., JIANG Y.G., YU G.L., PENG X., LI R.H. Elucidating the toxicity targets of  $\beta$ -ionone on photosynthetic system of *Microcystis aeruginosa* NIES-843 (Cyanobacteria). *Aquat. Toxicol.* **104**, (1-2), 48, **2011**.
- OZAKI K., OHTA A., IWATA C., HORIKAWA A., TSUJI K., ITO E., IKAI Y., HARADA K. Lysis of cyanobacteria with volatile organic compounds. *Chemosphere*, **71**, (8), 1531, **2008**.
- HARADA K.I., OZAKI K., TSUZUKI S., KATO H., HASEGAWA M., KURODA E.K., ARII S., TSUJI K. Blue color formation of cyanobacteria with  $\beta$ -cyclocitral. *J. Chem. Ecol.* **35**, (11), 1295, **2009**.
- GLINWOOD R., NINKOVIC V., PETERSSON J. Chemical interaction between undamaged plants-Effects on herbivores and natural enemies. *Phytochemistry*, **72**, (13), 1683, **2011**.
- RIPPKA R., DERUELLES J., WATERBURY J. B., HERDMAN M., STANIER R. Y. Generic Assignments, Strain Histories and Properties of Pure Cultures of Cyanobacteria. *J. Gen. Microbiol.* **111**, (1), 1, **1979**.
- YANG W.D., ZHANG X.L., LIU J.S. The influence of allelochemicals on the growth of *Alexandrium tamarense*. *China Environmental Science*, **25**, (4), 417, **2005** [In Chinese].
- ZHANG T.T., ZHENG C.Y., HE M., WU A.P., NIE L.W. Inhibition on algae of fatty acids and the structure-effect relationship. *China Environmental Science*, **29**, (3), 274, **2009** [In Chinese].
- HU C.Y., GE F.J., ZHANG S.H., LIU B.Y., WANG J., GAO Y.N., WU Z.B. Isolation of antialgal compounds from *Potamogeton malaianus* and algal inhibitory effects of common fatty acids. *J. Lake Sci.* **22**, (4), 569, **2010** [In Chinese].
- ALIOTTA G., DELLA GRECA M., MONACO P., PINTO G., POLLIO A., PREVITERA L. *In vitro* algal growth inhibition by phytotoxins of *Typha latifolia* L. *J. Chem. Ecol.* **16**, (9), 2637, **1990**.
- JARCHOW M. E., COOK B. J. Allelopathy as a mechanism for the invasion of *Typha angustifolia*. *Plant Ecol.* **204**, 113, **2009**.
- HONG Y., HU H.Y. Effects of the aquatic extracts of *Arundo donax* L. on the growth of freshwater algae. *Allelopathy J.* **20**, (2), 315, **2007**.
- HONG Y., HU H. Y., SAKODA A., SAGEHASHI M. Isolation and characterization of antialgal allelochemicals from *Arundo donax* L. *Allelopathy J.* **25**, (2), 357, **2010**.
- HONG Y., HU H. Y., SAKODA A., SAGEHASHI M. Effects of Allelochemical Gramine on Metabolic Activity and Ultrastructure of Cyanobacterium *Microcystis aeruginosa*. *World Academy of Science, Engineering and Technology*, **47**, 826, **2010**.