Review

Drought-Induced Changes in Leaf Morphology and Anatomy: Overview, Implications and Perspectives

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Abstract

The global climate change scenario intensified various environmental factors, especially in arid and semi-arid regions. Drought is one of the most severe environmental stresses affecting plant productivity. Plants in the Mediterranean climate zone are exposed to heat and drought in summer, and these conditions have a significant effect on plant growth and development. However, in this case, the entry of CO₂ into mesophyll cells is prevented and therefore the rate of photosynthesis decreases which ultimately causes a reduction in plant growth. In order to acclimate to stressful environmental conditions, plants exhibit several structural modifications to cope with these harmful conditions. This review highlights some aspects of anatomical adaptive changes in plants under drought stresssuch as a reduction in leaf size and angle, stomatal position, epidermal thickness and deposition of the cuticle to prevent the loss of water from the leaf surface. Furthermore, it elaborates the role of buliform cells in leaf rolling, structural adaptation in the mesophyll cells, and the presence of trichomes. Mesophyll cells and bulliform cells provide easier rolling of leaves in case of intense drought. In arid conditions, the economical use of water by plants is possible by closing the stomata and reducing transpiration.

Keywords: bulliform cell, drought, leaf rolling, mesophyll cells, stomata

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Introduction

The beginning of 21st century intensified the various environmental factors, especially in arid and semi-arid regions, due to extreme and rapid changes in climatic conditions [1], viz, rising air temperatures, atmospheric CO, levels, and high wind, which is projected to accelerate global climate change in future [2, 3]. All these events produce changes in rainfall patterns and distribution. Along with extreme rainfall events, the destruction of the ecological balance and freshwater scarcity has become serious environmental problem worldwide [4]. Under natural and agricultural conditions, plant growth and development require an optimal level of soil moisture, fluctuation in soil moisture beyond the optimal level can affect plant productivity. However, low water availability in the rhizosphere slows plant growth and limits the plant's ability to absorb nutrients [5, 6] Water constitutes a significant proportion of total fresh biomass in plants, ranging from 80% to 95%, which plays an essential role in several morpho-physiological and biochemical processes, including plant growth, development, and metabolism [7, 8].

Drought is the condition of prolonged dryness, which adversely affects the anatomical, morpho-physiological and other fundamental processes in the plant leading to limited plant growth and productivity [9, 10]. It also affects the location and density of the stomata on the leaf surface. The deposition of a waxy layer (cuticle) and positive thickness in the epidermal cells are the important features, that plant leaves adapt under drought stress conditions [11, 12]. Under drought stress, plants undergo several anatomical and morphological changes at cell to organ levels to cope with stress conditions. In the early phases of drought stress, plants normally maintain water balance by boosting water intake via the root system and decreasing water loss via stomata closure to prevent transpiration [13]. In the lateral stage, when plants are subjected to severe drought stress, a variety of structural changes are triggered in plant leaves. These changes include an accumulation of solutes in the cells to maintain turgor pressure [14], a change in leaf size, angle, and thickness [15, 16], an increase in the thickness of the cuticle and the number of stomata to minimize transpiration and water loss [17,

Plant leaves are the most important organs which are responsible life maintaining process like photosynthesis and plays a vital role in the transport of water and nutrients. Drought is one of the most severe abiotic stress and has detrimental affects on the morphological and anatomical characteristics of leaves which ultimately cause a reduction in crop growth and productivity. This review attempted to provide an overview of leaf morphological and anatomical responses under drought stress to maintain plant water status and crop productivity, mainly focusing on the role of leaf size and angle, the genesis of trichomes, the formation of bulliform cells, and their role in leaf rolling.

On the cellular level, functional traits associated with drought stress tolerance are also discussed.

Drought-Induced Changes in Plant Leaves

The changes observed in the plants during their vegetative periods of life by the harmful effects of drought stress are a gradual decline in the leaves' numbers and area, leaf wilting, cell elongation in the leaves, and enhanced senescence of leaves which ultimately cause a reduction in the total plant height. This reduction in plant height affects the morphology of the different plant essential organs. Plant leaves are the main organ for transpiration and act as an indicator for the water deficient condition. They have the capability to adapt their structural features such as increased thickness, tissue density, and reduce the area under such environments [19]. The first response of plants to drought is the change in the osmotic pressure of the cell. As the turgor pressure decreases, the cell water potential decreases, and cell expansion is limited, this is reflected in the morphological characteristics of the plant, and the first affected organ is the leaf, which is responsible for photosynthesis [20].

Specific leaf area is one of the morphological responses of the plant to drought stress. Specific leaf area components play an important role in photosynthesis, as they determine the flow and distribution of photons within leaves, limiting mesophyll to CO, diffusion to carboxylation sites in chloroplasts, especially under stress conditions [21]. Several previous studies reported that the reduction in the leaf area, change in the turgor, and canopy temperature, under drought, affect plant growth through a decline in the rate of photosynthesis [22]. Moreover, plants can roll their leaves which is the most important morphological feature of the leaves to cope with drought via loss the water in the upper epidermal cells which reduces press potential as a result of leaf roll as observed in the flag leaf of the wheat [23]. In severe harsh dry environments, some plants have thick film on their leaves and the rolling motion help to improve the loss of water under direct sunlight [24].

Since leaf morphology is important in plant growth and development, it significantly affects plant yield. The symptoms observed in the leaves in response to drought are important in the anatomical adaptation of the cultivars and are used in cultivar selection [10]. The responses of leaves to drought stress can be evaluated by looking at the leaf's anatomical features such as bulliform cells on the leaf surface, epidermal cells, and stomatal size. Drought affects the dehydration of leaf mesophyll cells by causing the closure of stomata and causes damage to photosynthetic organs [25].

Water-related morphological changes are the formation of fine roots, leaf rolling, and changes in growth patterns to increase water uptake from the soil [26]. Previous studies showed that drought stress significantly decreased the leaf numbers in maize [27],

and declined the total biomass and leaf area, and caused detrimental effects on photosynthesis [28, 29]. These complex modifications at the leaf level are an aspect to withstand drought, plants modify their growth patterns, distribute nutrients, and turn on stress-response genes. In addition to coping with water shortages, plants are able to secure their long-term resilience and capacity to recover when conditions improve due to their diverse adaptations.

Leaf Size and Angle under Drought Stress

The incline produced between the leaf blade and stem is referred to as the leaf angle [30, 31]. The cell wall' composition, expansion, and division at the lamina junction that connects the leaf blade determine how the leaf angle forms which is regulated by the hormones [32]. There are two types of physiological mechanisms responsible for the leaf angel, first one is the change in the growth of the cells on the upper and lower surface of the petiole and the second mechanism is the change in turgor potential at the part present at the base of leaves called the pulvinus. Plants alter their leaf angle in response to environmental factors such as light, water, gravity, and carbon dioxide. A leaf curling under water stress is an extreme example of how various portions of a leaf may alter its angles at different speeds [33].

The relationship between leaf angle and leaf rolling is inverse [34]. Resistance to water stress is correlated with a change in leaf angle. A shift in leaf angle can lower leaf temperature, conductivity, and transpiration by reducing photosystem inhibition and affecting the efficient utilization of water. In response to drought stress, several grass species roll their leaf blades, minimizing their exposure to stress [35].

Leaf size is the morphological feature of the plants responsible for the photosynthesis efficiency connected to variation in the leaf size [36]. It has been observed in the wheat leaves that the photosynthetic rate is high due to the narrow, smaller, erect, and larger deposition of cuticular wax on the epidermis of the leaf. These abovementioned overcome the water loss in the plants facing water deficiency [37]. The narrow-sized leaves have more resistance to drought stress as compared to the large-sized leaves [38]. The plants have flag leaves smaller in size and erect leaf angles are more adaptive to improve photosynthesis and decline in water loss through the evaporation process [39]. Therefore, reducing leaf angle could serve as one of the breeding objectives for wheat growth to increase plant density, enhance light absorption, and boost chlorophyll levels [40].

The grass plants can overcome the adverse effects of drought stress at a moderate level, because they have narrow, small-sized, and erect-angle leaves which lead to the utilization of the radiation that perfectly comes from the sun and contribute to improving the process of photosynthesis [41]. Plants alter their leaf

size and angle in order to achieve an ideal equilibrium between absorbing sunlight for photosynthesis and preserving valuable water under drought stress. These modifications serve as a dynamic survival strategy that highlights the incredible ability of plants to harsh environmental conditions. Such flexibility increases the chance that plants will survive, enabling them to tolerate water under drought stress.

Drought and Leaf Thickness

The leaf thickness (LT), which measures the space between its top and bottom surfaces, reveals the optical route that light takes through it and determines whether it will be repelled or absorbed. LT is closely related to productivity, responses to drought stress, and biomass partitioning [42, 43].

Under water deficiency the leaves have two-way approaches a) increase the thickness, and b) decrease the thickness of the leaves. Palisade and spongy tissue growth, as well as a reduction in leaf and stomata size, are ways to increase the capacity of plants to store water and minimize water loss in the former [17, 44]. To boost the capacity of CO₂ and inorganic nutrients to penetrate the leaves as well as enhance the exchange of gases to repair and maintain respiration under stress, certain plants thin their leaves or develop unique leaves [45, 46]. In order to adapt the stomata and optimize transpiration under water stress, the leaves' internal framework is altered. However, the cause for this is unclear, necessitating additional research.

Leaf area, leaf thickness, leaf density, and stomatal structure in the plant vary depending on the plant's water content [47]. The increase in leaf thickness in plants in drought conditions is due to the increase in upper epidermis thickness. The thick epidermis layer in grass plants prevents high water loss from leaves in dry conditions. The leaf thickness of the varieties with moderate leaf rolling in arid conditions in rice decreased more than the ones with more leaf rolling [48]. Drought stress reduces leaf thickness in relation to the net carbon absorption and photosynthetic performance of plants [20]. If a plant has a high drought tolerance, it shows that biomass losses will be less [10].

In arid conditions, lamina thickness increased in the moderately drought-tolerant and highly tolerant sugarcane genotypes. The cell walls and cuticle thickness of the lamina epidermal cells of the genotype, which is moderately drought resistant, also increased. In moderately drought-tolerant sugarcane, with drought, stomata size increased in leaves, while a decrease was observed in the highly drought-tolerant genotype [18]. The complex interactions between drought and leaf thickness demonstrate the mechanisms that plants use to survive in harsh environments. By enabling plants to tolerate water stress, these adaptations not only benefit in immediate survival but also contribute to long-term resilience in the ecosystem.

Leaf Rolling under Drought Stress

Leaves are the most important organ in the plant body which are responsible for photosynthesis. Reduction in the leaf expansion, stomatal conductance and the assimilate produced through the photosynthesis in the leaves via the adverse effect of drought stress cause to reduce the yield of the crops. Morphological adaptation in the leaves such as leaf rolling, loss of turgor, and osmotic adjustment to mitigate the abovementioned issues by the drought [49].

Leaf rolling is the most observed phenomenon caused by the change in the water potential in the bulliform cells present in the epidermis of the leaves. This phenomenon plays a vital role to slow down the process of transpiration and enhance the yield-contributing indices in plants that face water deficiency [50]. The top layer of the epidermis of the leaf loses water, which decreases the pressure potential, which makes the leaf roll. This process is beneficial by lowering leaf temperature, higher light absorption, and enhancing the rate of transpiration. Leaf area and leaf rolling were significantly increased in *Zea mays* L. (maize) leaves grown under drought stress regimes [51].

The primary indicator of plants is leaf rolling in response to drought stress involving the amount of thickness in the lamina and swelling of the epidermal cells [52]. By adjusting the leaf rolling, it is possible to control the efficiency of photosynthetic activity per unit leaf area. The amount of dry matter accumulation and transpiration may both be increased and decreased by minimizing the effect of sunlight on the leaves and achieving optimal leaf roll [10]. The rolling that occurs in the leaves also reduces the damage caused by light by reducing the areas exposed to the sun in the plant [53]. The water loss in rice is 36% in the case of partial curling of the leaf and 52% in the case of full rolling [54]. Therefore, a positive correlation was observed between the degree of leaf rolling and water loss.

The patterns of leaf rolling, which include rolling inward or outward, facilitate effective photosynthetic processes in leaves [55]. As compared to entirely rolled leaf behavior, the optimal expression of leaf rolling is helpful for increasing water usage efficiency [56]. A similar pattern of leaf rolling was observed in the wheat plant which contribute to reducing the loss of water from the leaf surface via lowering interaction between the leaf surface and direct sunlight, while simultaneously enabling light to penetrate farther into the canopy [57, 58]. Under drought stress, rice (Oryza sativa L.) leaves frequently show two leaf rolling (LR) patterns known as adaxial and abaxial rolling which is the inward and outward motion of the leaves [59, 60]. It is always believed that optimal/partial LR is a better option for mitigating the effects of dryness than fully rolling or fully flattened leaf appearances [53]. The optimal LR is the most effective strategy which takes part to improve the photosynthetic rate under drought stress and overcome water loss. This

phenomenon can be observed in the afternoon when the direct sunlight interaction with the surface of the leaves attains a high level [55, 61, 62].

In the rice plant exposed to drought, rolling occurs in the leaves due to bulliform cell shrinkage [10]. Similarly, a decrease in bulliform cell size is observed in rice varieties in arid conditions [20]. The decrease in cell size caused a decrease in transpiration rate in the plant and prevented high water losses. As a result of all these, rounding of the leaves of the plant has occurred. Vascular bundle size and amount were found to be associated with leaf rolling in rice under drought conditions [52].

Environment-related factors such as a shortage of water, a warm climate, and exposure to sunlight are the main causes of leaf rolling in plants. Changes in photosynthetic rates, ion levels, fluctuations in the systems that produce antioxidants, and cell forms are additional factors that contribute to leaf rolling. In plants that become more drought-resistant, a lack of water, an increase in temperature, and the rays of the sun accelerate leaf rolling, this process plays an important role in reducing the water loss from stomata [63]. Leaf rolling is a drought-induced response that helps plants maintain vital physiological functions and conserve moisture, allowing them to rely on it until favorable conditions return, thus increasing their chances of survival during drought periods.

Bulliform Cells and Their Positive Role in Leaf Rolling

Bulliform cells have been linked to the leaf-rolling response observed commonly in the leaves of grasses under drought stress. These cells develop in longitudinal strips on the adaxial leaf surface. During the water deficiency in the leaf, bulliform cells showed significant shrinkage in comparison to other epidermal cell types, offering a potential mechanism for facilitating leaf rolling [64]. Plants need an adequate amount of water for proper development during their vegetative stage of growth, its consumption varies in every stage of their life cycle, that's why LR acts as a vital adaptation under drought stress [13]. The change in the turgor pressure in the bulliform cell present on the upper layer of epidermal cells in the leaves cause leaf rolling to be a quick response against water-deficient conditions and prevents the leaves from large exposure to sunlight [65, 52, 64].

Bulliform, collenchymal, mesophyll, and vascular bundle cells are among the cells involved in LR. However, one of the main causes of LR in rice is the shrinking due to decreased turgor pressure of bulliform cells on both sides of the leaf [66, 67]. Bulliform cells expand in arid conditions in sugar beet [68]. The response to drought in sugarcane is the thickness of the cuticle layer, the enlargement of the vesicles in the bulliform cells, and the increase in the number of veins in the leaf. In addition, the lower and upper cuticle

thicknesses increased under drought stress. The number of green leaves, chloroplast content, chloroplast length, width, and width/length ratio are very important in selecting drought-resistant varieties in sugarcane [15]. Twelve days after drought in rice, a decrease in bulliform cell size, which is easily affected by turgor pressure, is observed. Thus, a high rate of evapotranspiration is prevented [25]. In short, bulliform cells are crucial in the physiological adaptation of leaf rolling, which improves water-saving behaviors and boosts the plant's tolerance to drought stress, making it an advantageous modification for plants.

Is Leaf Rolling Necessary in Plants?

However, since the leaf blade becomes flattened if rolling does not occur, greater amounts of sunlight will strike the leaf surface, enhancing transpiration through the stomata in order to prevent plant heating up [69]. As a consequence of this, plants will rapidly wilt. If stress takes a long time, it could also cause problems with photosynthetic activity and change in the metabolic function of the plant causing plant death. However, leaf rolling occurs in the leaves which contributes to improving the metabolic function in the leaves and induces resistance against water deficiency is a positive step toward the survival of the plants [6]. It has been reported in a number of studies that moderate LR enhances the photosynthetic efficiency, while high-level or complete rolling decrease this efficiency and causes adverse effects on the plant [70, 71].

Photosynthetic activity ceased by the extreme rolling of the leaves which is considered to not play a positive role in the other beneficial process in the plant. Therefore, the rolling approach of the leaves is the most critical process for the perfect functioning of the plants [72]. Bulliform cells are an example of a vital strategy to water conservation and photosynthesis enhancement. The crucial role of leaf rolling shows how plants can adapt to drought stress. Understanding the complex workings of bulliform cells not only increases our understanding of how resilient plants are but also stimulates new ideas for improving water use efficiency in agriculture and reducing the effects of water shortages in the face of global warming.

Drought Stress and Trichomes

Trichomes are the hair-like fine outgrowth on the surface of the leaves which have variability in their density, size, locality, and functioning according to plant species. There are two main types of trichomes first one is the glandular trichomes which secret the chemicals to prevent the plants from external factors such as herbivorous and pathogens attacks and the second type is the non-glandular trichomes which function to act as barriers against harsh conditions by the reflecting the rays from the sun, reduce the process of transpiration

and also absorbed water [73-75]. The two types have the capacity to secrete or store vast amounts of specialized metabolites, which increase a plant's resistance to harsh climatic circumstances [76].

In terms of morphology, leaves with more trichomes in terms of leaf area are more resistant to abiotic degradation because transpiration prevents excessive water loss and regulation of temperature [77]. According to a study, trichome numbers significantly increased in the dry season compared to the wet season, which affected the physiological processes of plants. A number of physiological functions including stomatal conductance, transpiration rate, and water consumption efficiency, can be significantly affected by the presence of leaf trichomes [78].

Because bulliform cells, which absorb and store water, are abundant in the basal section of the leaf trichomes in grasses provide water to leaves. Trichomes influence how water interacts with leaf surfaces by directing water droplets away from the stem and soil and towards them, which helps plants absorb water. By decreasing transpiration, modifying the energy balance, and lowering light absorption, it also improves water storage [79, 78, 80]. The connection between trichome density and physiological features under water-deficit stress in many plant species is still not evident even though this information is well-documented in many herbaceous crops. It will be easier to explain the growing susceptibility of plant species to drought if we have a better knowledge of this link.

Drought-Induced Changes in Stomata

Drought stress not only affects the morphological features of the plants but also has adverse effects on physiological attributes such as relative water content, relative water humidity, and stomatal conductance of the plants and also photosynthetic pigments which directly attach with the anatomical features of the leaves. Leaf anatomical features to be observed due to water deficiency can be used as important visual indicators to determine the sensitivity and tolerance of the plant to arid conditions [20].

Plants that have experienced a long period of drought stress have smaller stomata which are fewer in number to reduce water loss and maximize plant water consumption which leads to early and quick response against drought. Usually, controlling leaf vein density is used to attain this equilibrium [81]. The plants under stress conditions close their stomata to overcome the loss of water by the phenomena of transpiration, as a result, decrease in the amount of carbon dioxide (CO₂) in the internal part of leaves. During the light reaction of photosynthesis, the disturbance in the electron transport chain and the activation of the glycolate oxidase pathway produce a high amount of reactive oxygen species (ROS) which cause oxidative stress in the plants [20].

The opening and number of stomata decreased with the increase in the drought. These characteristics of the stomata such as size, density, and conductivity are very important to reduce the loss of water and the flow of CO2 in the cells of plant leaves that take part in photosynthesis [15, 68]. Changes in the structure of stomata are closely related to abscisic acid (ABA), a hormone produced during times of stress. During dehydration in the root zone, ABA is rapidly synthesized and transported. This causes interactions with jasmonic acid and nitric oxide to stimulate stomatal closure. This occurs through changes in signal transduction, including changes in guard cell turgor and ion pumping through ion channels [20].

The plants of the family Poaceae have special anatomical features and are known as C_4 plants which regulate their stomata to prevent the loss of water and enhance carbon assimilation leading to positive help in the physiological process under water stress conditions [82]. So, the C_4 plants have higher water use efficiency and are considered to become drought tolerant than the plants belonging to the C_3 species. The number of stomata in grass plants demonstrated a substantial negative connection with the drought period [26].

Stomatal size plays an important role in the rice cultivars under water deficiency, the cultivars with large stomatal size have high transpiration which contributes to wilting more frequently. The reduction in the turgor pressure changes the size of the stomata. Therefore, the cultivars that have larger-sized stomata in drought stress are more sensitive [25]. In plants, stomatal density increases significantly under moderate stress [83] (Kofidis et al., 2004) and decreases under severe stress conditions [84]. The increase in stomatal density in dry conditions also helps to control sweating better [85].

Under conditions of water stress, the density of the stomata, which reduce moisture in the plant through the leaves, is crucial for plants. Different cotton (Gossypium hirsutum L.) genotypes have different stomatal densities in dry circumstances. Under drought stress, stomatal density in cotton genotypes decreased in the range of 10-23 mm² compared to control conditions. The maximum reduction in the number of stomata resulted in greater water retention in the plant due to less transpiration [86]. Plants have extensive mechanisms for optimizing gas exchange and preservation of water, which is highlighted by the extraordinary plasticity of stomata in response to drought. This illustrates their capacity to grow even in difficult circumstances.

Cuticular Changes under Drought

The outermost layer of the leaves is synthesized from the epidermal cells and is called the cuticle, which constitutes cutin and cuticular waxes. It protects the inner tissues from other environmental factors such as biotic and abiotic stress such as harmful radiation, fungus and pathogens effects, and water loss under

dry conditions [87, 88]. Cuticular waxes, on the other hand, play a major role in controlling non-stomatal water loss, making them an important adaptation in the development of terrestrial plants [89, 90]. It plays a vital role in the physiology of plants by delaying cellular water loss from the leaves and acts as a key adaptation [91].

Glaucousness, a bluish-white coloring caused by densely dispersed epicuticular wax crystalloids, is a common outcome of epicuticular wax accumulation on plant surfaces. These phenomena decrease the leaf temperature by the reflectance enhancement of radiations which is helpful for the survival of leaves in water-deficient environments [88, 92]. Xerophytic plants have more thickness in their cuticles to the enhancement in the production of cuticular waxes from the epidermal cells of the leaves [93]. It has been observed in the Arabidopsis plant that the increase in thickness of the cuticle per unit area of the leaf with the deposition of cuticular wax on the leaf surface [94]. This change in the cuticle surface decreases the permeability of the cuticle which is a positive step toward the stress via decline in the water loss.

The results of the previous studies witnessed that an increase in the deposition of the cuticular waxes on the surface of the leaves is the key adaptation to increase the resistance under water-deficient environments [95, 96]. Enhancement in the wax accumulation on the upper epidermal surface in the arabidopsis (Arabidopsis thaliana L.) [97], alfalfa (Medicago sativa L.) [98], and Camelina (Camelina sativa L.) [99] through transcriptional factor overexpression under drought stress. The connection between the cuticle and drought stress was also observed in the barley plants which are bread to high tolerance and yield under this stress [100]. Cuticle and epidermal structures are effective features in the adaptation of plants to drought [25]. The continuation of functions of some species in arid soils may be due to various physiological changes such as an increase in cuticle thickness and a decrease in stomatal size and density [101, 26]. Drought-induced stress in plants leads to significant alterations in their tissues, such as increased wax accumulation, which is crucial for reducing water loss and enhancing their climatic resistance.

Cuticle Thickness and Stomatal Density Responses in Monocots vs Dicots

The formation of a hydrophobic cuticle on the outer surface of the leaves which plays a crucial role against external environmental changes, was one of the most crucial adaptive features for survival [102]. Deposition in the cuticle on the epidermal cells is one of the modifications that plants adopt to enhance tolerance in water-deficient conditions [95]. A positive increase in the thickness was observed in both monocot and dicot plant leaves such as *Hordeum vulgare* L. [103], *Gossypium*

hirsutum L. [104], Nicotiana Tabacum L. [105], Glycine max L. [106], Triticum aestivum L. [88], Arabidopsis thaliana L. [107] and Sorghum bicolor L. [108] under water stress. However, the thickness of the leaf cuticle is varying among the plant species or even in the same plants under different environmental conditions. In some cases, the monocot has thick cuticles as a key adaptation in response to drought stress, while in the same circumstance, the dicot develops thicker cuticle on the leaf surface. So, on a broad spectrum stating that the monocot has more thick cuticles than the dicot according to the external environment is not accurate and vice versa.

Stomata defines the ability of plants for gas exchange, including atmospheric carbon dioxide uptake for photosynthesis and water vapor release during transpiration. Stomata density may vary among plant species under different environmental conditions like drought stress [109]. A number of monocot species such as *Hordeum vulgare* L. [110, 111], *Oryza sativa* L. [112] and *Triticum aestivum* L. [113] may show a drop in stomatal density under drought stress. In dry situations, this can reduce transpiration to lower water loss and improve waterar conservation.

The dicot plants may also respond differently to drought stress in terms of their stomatal density, depending upon the species and its mechanisms of adaptation. Some dicot reduces their stomatal density like monocots to decline in water loss such as *Amaranthus viridis* L. [114] and *Vitis vinifera* L. [115]. Some dicot species that can withstand drought may continue to have larger stomatal densities to continue photosynthesis as well as development when water is limited [116, 117]. The comparative study of monocot and dicot in responses to cuticle thickness and stomatal density showed in Table.1.

Mesophyll Cells under Drought Stress

Mesophyll cells, the predominant cell type in leaves and the fundamental site of photosynthesis are organized in cylinders called palisade mesophyll cells (PMCs) on the upper side and spherical, spongy mesophyll cells (SMCs) on the lower side of the leaf. Modifications in leaf phenotypes result from PMCs and SMCs losing their dorsoventral differentiation [121].

In arid conditions, leaf thickness is associated with photosynthesis rate and plant growth. In drought-resistant plants, leaf thickness increased under drought-stress conditions, resulting in an increase in mesophyll density [18]. Changes in mesophyll thickness, vascular sheath, and sclerenchyma layer were observed in plants with drought stress [52, 20]. Compared to drought-susceptible plants, drought-adapted plants' leaves usually feature more densely packed, elongated cells, but thinner spongy mesophyll cells.

The influence of drought stress on the growth of plant leaves has been extensively investigated [122]. The growth of leaf epidermal cells and physical leaf shape are closely related phenomena [123]. The mesophyll, a kind of cell that experiences significant fluctuations in turgor status, can be found near the guard cells. This means it is the optimal tissue for converting rapid fluctuations in water stress into the quick ABA biosynthesis necessary to regulate stomatal responses [124, 125]. Additionally, many seed plant species depend on the numerous chloroplasts in mesophyll cells as a virtually endless source of carotenoid precursors to fuel the continuous production of ABA, which is necessary for keeping stomatal closure throughout protracted periods of soil water deficiency [126].

Mesophyll Palisade Cells

With drought stress, dense and smaller mesophyll palisade cells are observed in the plant [101, 25], and the space between mesophyll cells decreases

Table 1. Response	s of cuticle thickness	and stomatal density	in monocot and dicot pl	lants.

Group of plants	Cuticle thickness	Stomatal density	Plant species	Responses	References
Monocot	Increased	Decreased	Barley (Hordeum vulgare L.)	Reduced evaporation of water on the surface of leaves, high water use efficiency	[103, 110]
			Rice (Oryza sativa L.)	Enhanced tolerance against drought stress and improved yield	[118, 112]
			Wheat (Triticum aestivum L.)	Improved water use efficiency and yield	[88, 113]
Dicot	Increased	Increased	Arabidopsis (Arabidopsis thaliana L.)	-	[107, 119]
	increased	Increased	Cotton (Gossypium hirsutum L.)	Improved physiological mechanisms to reduce drought stress effects	[120, 117]
	-	Decreased	Green amaranth (Amaranthus viridis)	Positive enhancement in the physiological indices	[114]

and increases the number of epidermal cells and mesophyll chlorenchyma cells [83]. It also significantly reduced leaf and epidermis thickness, number of palisade cells, length of palisade mesophyll, number of sponge cells, and length of sponge mesophyll in peanuts [127].

Leaf dry matter per area limits the photosynthetic efficiency by affecting the conductivity of mesophyll to CO₂ [128-130]. The conductivity of the mesophyll to CO₂ is largely determined by the anatomical features of the mesophyll in adapting to long-term stresses such as drought [131, 132]. Drought cell wall thickness increased the thickness between the palisade and spongy mesophyll. In drought conditions, olive also increases the number of mesophyll cells and sclereids in order to maintain photosynthesis as well as to reduce transpiration, the spaces between mesophyll cells are reduced and the amount of non-glandular surface in the leaf increases significantly [85].

According to a previous investigation, the plants that face the issue of drought stress have thicker and large numbers of palisade cells as compared to the well-watered plants [133]. Starch stores were depleted as a result of structural damage imposed by drought stress in palisade cells. The efficiency in exchanges of gases within the leaves and ultimately the creation of starch reserves is both impacted by damage to the vein network of the leaf [134, 135]. Lack of water can cause the leaves to shrink, which can seriously harm the vein networks, mesophyll tissues, and plastids, where starch is produced, structurally [136]. Previous research has shown that starch stores are crucial for preserving plant energy as well as growth and during the course of the photoperiod, plant starch stores can be increased and transformed into soluble sugars as osmolytes to sustain plant development during stress caused by drought [137]. Many plants have been shown to deplete starch stores in response to drought stress, but further research is needed to determine whether structural destruction of palisade cells occurs when the leaves shrink. Plants possess a remarkable capacity to overcome the adverse effects of harsh environmental conditions by modifying the structure of their cells and maintaining vital processes while decreasing the water loss. Mesophyll cells complex interaction reflects the complexity of its resistance and provides facts about plant survival mechanisms that might motivate environmentally friendly farming methods.

Epidermal Cells under Drought Stress

When plants are exposed to drought conditions, transpiration decreases and leaf epidermis thickness increases, cells, and intercellular spaces decrease, vascular tissue, root/shoot ratio increases [85, 138]. In dry conditions, the upper and lower epidermis and cuticle thickness decrease in the leaf. Drought reduces many anatomical features in the leaf, and differences are observed in drought-tolerant and sensitive species [139].

The smooth walls and small size of epidermal cells create resistance to dry conditions, while small epidermal cells show 20 times more resistance to dry conditions than large ones [85]. After drought stress, the thickness of the epidermis, which is the protector of photosynthetic organs, increases in rice cells [25]. The width/length of the upper epidermal cells (μm/μm) varied between 0.67±0.13 under control conditions and 0.75±0.11 under drought stress, while the width/length (µm/µm) of lower epidermal cells was 0.67 under control conditions. ± 0.11 while it was 0.7 ± 0.07 under stress conditions [140]. Plants in extreme drought conditions can enhance their survival by thickening epidermal cells, which store water and respond to stress through stomatal control and trichome growth, thereby enhancing their water conservation and drought resistance.

Conclusions

In arid and semi-arid regions, drought stress poses a significant global challenge, severely limiting both the quantity and quality of crop production. The morphological and structural strategies during drought stress hold significant importance concerning the enhancement of water use efficiency, drought tolerance, and crop plant productivity. Mesophyll and bulliform cells play a crucial role in the tolerance against drought stress. Mesophyll cells photosynthesize the leaf. During drought stress, mesophyll cells store osmolytes to maintain turgor pressure and boost antioxidant enzyme activity to decrease dehydration damage. Bulliform cells are thin-walled monocot epidermal cells that cover the leaf. Bulliform cells shriveled more than other epidermal cells and reduced water loss during leaf dryness. The combination of bulliform and mesophyll cells increases a plant's tolerance to drought, providing a potential mechanism in leaf rolling and storing osmolytes that helps plants to conserve water and protect their photosynthetic cells. In addition, drought-tolerant plants have evolved a range of other structural changes such as a change in leaf angle, size, and area, an increase in the thickness of the cuticle, and the number of stomata to minimize transpiration and water loss has been observed in response to drought stress. Moreover, this review illustrates how bulliform cells take part in leaf rolling which is the key trait to overcome drought stress.

In crux, this review will help researchers understand the morphological and anatomical traits in plants, but further studies are needed to fully comprehend its genetics and molecular mechanisms and its potential application in plant breeding. Further studies are also needed to understand the molecular and genetic mechanisms behind the leaf's anatomical changes of stressed plants, as they play a vital role in developing drought early warning systems, identifying crop risks, and minimizing crop damage. Genetic control of these features can enhance leaf modification for improved

agricultural productivity and water use efficiency, in conjunction with phonemics strategies.

Authors' Contributions

IY conceptualized the manuscript. IY, MAJ and KU prepared the initial manuscript. S.H. and MF reviewed the manuscript. All authors read and approved the final manuscript.

Conflict of Interests

The authors declare that they have no conflict of interest.

References

- TRENBERTH K.E., DAI A., VAN DER SCHRIER G., JONES P.D., BARICHIVICH J., BRIFFA, K.R., SHEFFIELD, J. Global warming and changes in drought. Nature Climate Change, 4 (1), 17, 2014.
- YIN J., GENTINE P., ZHOU S., SULLIVAN S.C., WANG R., ZHANG Y., GUO S. Large increase in global storm runoff extremes driven by climate and anthropogenic changes. Nature communications, 9 (1), 4389, 2018.
- YANG H., HUNTINGFORD C., WILTSHIRE A., SITCH S., MERCADO L. Compensatory climate effects link trends in global runoff to rising atmospheric CO₂ concentration. Environmental Research Letters, 14 (12),124075, 2019.
- SIVAKUMAR M.V., MOTHA R.P., DAS H.P. Natural disasters and extreme events in agriculture: impacts and mitigation. eds., Berlin, Heidelberg: Springer Berlin Heidelberg, 2005.
- DA SILVA E.C., DE ALBUQUERQUE M.B., DE AZEVEDO NETO A.D., DA SILVA JUNIOR C.D. Drought and its consequences to plants – From individual to ecosystem. Responses of organisms to water stress, 18, 2013.
- SELEIMAN M.F., AL-SUHAIBANI N., ALI N., AKMAL M., ALOTAIBI M., REFAY Y., DINDAROGLU T., ABDUL-WAJID, H.H., BATTAGLIA, M.L. Drought stress impacts on plants and different approaches to alleviate its adverse effects. Plants, 10 (2), 259, 2021.
- 7. OLIVEIRA C., ANTUNES C.H., A multi-objective multi-sectoral economy–energy–environment model: Application to Portugal. Energy, **36** (5), 2856, **2011**.
- 8. HUSSAIN H.A., HUSSAIN S., KHALIQ A., ASHRAF U., ANJUM S.A., MEN S., WANG L. Chilling and drought stresses in crop plants: implications, cross talk, and potential management opportunities. Frontiers in plant science, 9, 393, 2018.
- ANJUM S.A., XIE X., WANG L.C., SALEEM M.F., MAN C., LEI W. Morphological, physiological and biochemical responses of plants to drought stress. African journal of agricultural research, 6 (9), 2026, 2011.
- GUNNULA W., KANAWAPEE N., SOMTA P., PHANSAK P. Evaluating anatomical characteristics associated with leaf rolling in northeastern Thai rice cultivars during drought by decision tree. Acta Agrobotanica, 75 (1), 2022.

- ABOBATTA W.F. Drought adaptive mechanisms of plants
 A review. Advances in Agriculture and Environmental Science, 2 (1), 62, 2019.
- BANO C., AMIST N., SINGH N. B. Morphological and anatomical modifications of plants for environmental stresses. Molecular plant abiotic stress: biology and biotechnology, 29, 2019.
- 13. HUSSAIN H.A., MEN S., HUSSAIN S., CHEN Y., ALI S., ZHANG S., ZHANG K., LI Y., XU Q., LIAO C., WANG, L. Interactive effects of drought and heat stresses on morphophysiological attributes, yield, nutrient uptake and oxidative status in maize hybrids. Scientific reports, 9 (1), 3890, 2019.
- SANDERS G.J., ARNDT S.K. Osmotic adjustment under drought conditions. In Plant responses to drought stress: From morphological to molecular features. Berlin, Heidelberg: Springer Berlin Heidelberg. 199, 2012.
- ZHANG F.J., ZHANG K.K., DU C.Z., LI J., XING Y.X., YANG L.T., LI Y.R. Effect of drought stress on anatomical structure and chloroplast ultrastructure in leaves of sugarcane. Sugar Tech, 17, 41, 2015.
- 16. JÚNIOR S.D.O.M., DE ANDRADE J.R., DOS SANTOS C.M., SILVA J.A.C., D SANTOS K.P., SILVA J.V., ENDRES L. Leaf thickness and gas exchange are indicators of drought stress tolerance of sugarcane. Emirates Journal of Food and Agriculture, 29, 2019.
- NAWAZ T., HAMEED M., ASHRAF M., AHMAD M.S.A., BATOOL R., FATIMA S. Anatomical and physiological adaptations in aquatic ecotypes of Cyperus alopecuroides Rottb. under saline and waterlogged conditions. Aquatic botany, 116, 60, 2014.
- 18. TARATIMA W., RITMAHA T., JONGRUNGKLANG N., MANEERATTANARUNGROJ P. KUNPRATUM N. Effect of stress on the leaf anatomy of sugarcane cultivars with different drought tolerance (Saccharum officinarum, Poaceae). Revista de Biología Tropical, 68 (4), 1159, 2020.
- PATMI Y.S., PITOYO A. Effect of drought stress on morphological, anatomical, and physiological characteristics of Cempo Ireng cultivar mutant rice (*Oryza sativa* L.) strain 51 irradiated by gamma-ray. In Journal of Physics: Conference Series 1436, 012015, 2020.
- 20. SALSINHA Y.C.F., MARYANI INDRADEWA D., PURWESTR Y.A., RACHMAWATI D. Leaf physiological and anatomical characters contribute to drought tolerance of Nusa Tenggara Timur local rice cultivars. Journal of Crop Science and Biotechnology, 24, 337, 2021.
- 21. VELIKOVA V., ARENA C., IZZO L.G., TSONEV T., KOLEVA D., TATTINI M., ROEVA O., DE MAIO A., LORETO F. Functional and structural leaf plasticity determine photosynthetic performances during drought stress and recovery in two Platanus orientalis populations from contrasting habitats. International journal of molecular sciences, 21 (11), 3912, 2020.
- TAIZ L., ZEIGER E., MØLLER I.M., MURPHY, A. Plant physiology and development (No. Ed. 6). Sinauer Associates Incorporated. Pp, 761, 2015.
- 23. WILLICK I.R., LAHLALI R., VIJAYAN P., MUIR D., KARUNAKARAN C., TANINO, K.K. Wheat flag leaf epicuticular wax morphology and composition in response to moderate drought stress are revealed by SEM, FTIR-ATR and synchrotron X-ray spectroscopy. Physiologia plantarum, 162 (3), 316, 2018.
- 24. RUEDA M., GODOY O., HAWKINS B.A. Spatial and evolutionary parallelism between shade and drought tolerance explains the distributions of conifers in the conterminous United States. Global Ecology and Biogeography, 26 (1), 31, 2017.

 ZAGOTO A.D.P., VIOLITA V. X Leaf anatomical modification in drought of rice varieties (*Oryza sativa* L.). Eksakta: Berkala Ilmiah Bidang MIPA, 20 (2), 42, 2015.

- BACHLE S., ZARICOR M., GRIFFITH D., QUI F., STILL C.J., UNGERER M.C., NIPPERT J.B. Physiological responses to drought stress and recovery reflect differences in leaf function and anatomy among grass lineages. bioRxiv, 2022, 2022.
- 27. SHEMI R., WANG R., GHEITH E.S.M., HUSSAIN H.A., HUSSAIN S., IRFAN M., CHOLIDAH L., ZHANG K., ZHANG S., WANG L. Effects of salicylic acid, zinc and glycine betaine on morpho-physiological growth and yield of maize under drought stress. Scientific Reports, 11 (1), 3195, 2021.
- 28. HUSSAIN H.A., MEN S., HUSSAIN S., ZHANG Q., ASHRAF U., ANJUM S.A., ALI, I., WANG, L. Maize tolerance against drought and chilling stresses varied with root morphology and antioxidative defense system. Plants, 9 (6), 720, 2020.
- 29. KUMAR S., ISLAM A.R.M.T., ISLAM H.T., HASANUZZAMAN M., ONGOMA V., KHAN R., MALLICK J. Water resources pollution associated with risks of heavy metals from Vatukoula Goldmine region, Fiji. Journal of environmental management, 293, 112868, 2021.
- 30. ZHAO S.Q., HU J., GUO L.B., QIAN Q., XUE H.W. Rice leaf inclination2, a VIN3-like protein, regulates leaf angle through modulating cell division of the collar. Cell research, 20 (8), 935, 2010.
- 31. MANTILLA-PEREZ M.B., SALAS FERNANDEZ M.G. Differential manipulation of leaf angle throughout the canopy: current status and prospects. Journal of Experimental Botany, 68 (21-22), 5699, 2017.
- 32. KONG F., ZHANG T., LIU J., HENG S., SHI Q., ZHANG H., WANG Z., GE L., LI P., LU X., LI G. Regulation of leaf angle by auricle development in maize. Molecular Plant, 10 (3), 516, 2017.
- 33. YANG X., LI R., JABLONSKI A., STOVALL A., KIM J., YI K., MA Y., BEVERLY D., PHILLIPS R., NOVICK K., XU X. Leaf angle as a leaf and canopy trait: Rejuvenating its role in ecology with new technology. Ecology Letters, 26 (6), 1005, 2023.
- 34. HAKEEM S., ALI Z., SADDIQUE M.A.B., MERRIUM S., ARSLAN M., HABIB-UR-RAHMAN M. Leaf wettability and leaf angle affect air-moisture deposition in wheat for self-irrigation. BMC Plant Biology, 23 (1), 1, 2023.
- 35. BRIGLIA N., WILLIAMS K., WU D., LI Y., TAO S., CORKE F., MONTANARO G., PETROZZA A., AMATO D., CELLINI F., DOONAN J.H. Image-based assessment of drought response in grapevines. Frontiers in plant science, 11, 595, 2020.
- 36. SEVANTO S. X Phloem transport and drought. Journal of experimental botany, **65** (7),1751, **2011**.
- 37. BALOTA M., PAYNE W. A., EVETT S. R., PETERS T. R. Morphological and physiological traits associated with canopy temperature depression in three closely related wheat lines. Crop Science, 48 (5), 1897, 2008.
- 38. KUMAR A., BHARTI B., KUMAR J., TRIPATHI A., GAHTYARI N.C., JAISWAL J.P., VISHWAKARMA S.R. Genetic variability for morphological traits in released varieties of barley (*Hordeum vulgare* L.) under partially reclaimed saline-sodic soil. Indian Journal of Plant Genetic Resources, 29 (01), 45, 2016.
- 39. LIU J., XU Z., FAN X., ZHOU Q., CAO J., WANG F., JI G., YANG L., FENG B., WANG T. A genome-wide association study of wheat spike related traits in China. Frontiers in plant science, 9, 1584, 2018.

- 40. FISCHER R.A., EDMEADES G.O. Breeding and cereal yield progress. Crop science, 50, 85, 2010.
- 41. ZHAO C., BAO Y., WANG X., YU H., DING A., GUAN C., CUI J., WU Y., SUN H., LI X., ZHAI D. QTL for flag leaf size and their influence on yield-related traits in wheat. Euphytica, 214, 1, 2018.
- 42. PAULI D., WHITE J.W., ANDRADE-SANCHEZ P., CONLEY M.M., HEUN J., THORP K.R., FRENCH A.N., HUNSAKER D.J., CARMO-SILVA E., WANG G., GORE, M.A. Investigation of the influence of leaf thickness on canopy reflectance and physiological traits in upland and Pima cotton populations. Frontiers in plant science, 8, 1405, 2017
- 43. CONEVA V., CHITWOOD D.H. Genetic and developmental basis for increased leaf thickness in the Arabidopsis Cvi ecotype. Frontiers in Plant Science, 9, 322, 2018.
- 44. ZÚÑIGA-FEEST A., BUSTOS-SALAZAR A., ALVES F., MARTINEZ V., SMITH-RAMÍREZ, C. Physiological and morphological responses to permanent and intermittent waterlogging in seedlings of four evergreen trees of temperate swamp forests. Tree Physiology, 37 (6), 779, 2017.
- 45. BRODERSEN K.E., HAMMER K.J., SCHRAMEYER V., FLOYTRUP A., RASHEED M.A., RALPH P.J., KÜHL M., PEDERSEN O. Sediment resuspension and deposition on seagrass leaves impedes internal plant aeration and promotes phytotoxic H2S intrusion. Frontiers in plant science, 8, 657, 2017.
- 46. LAWSON J.R., FRYIRS K.A., LEISHMAN M.R. Interactive effects of waterlogging and atmospheric CO₂ concentration on gas exchange, growth and functional traits of Australian riparian tree seedlings. Ecohydrology, 10 (3), 1803, 2017.
- 47. BINKS O., MEIR P., ROWLAND L., DA COSTA A.C.L., VASCONCELOS S.S., DE OLIVEIRA A.A.R., FERREIRA L., MENCUCCINI M. Limited acclimation in leaf anatomy to experimental drought in tropical rainforest trees. Tree Physiology, 36 (12), 1550, 2016.
- CONESA M.A., MUIR C.D., MOLINS A., GALMÉS
 J. Stomatal anatomy coordinates leaf size with Rubisco kinetics in the Balearic Limonium. AoB Plants, 12(1), 050, 2020
- 49. MOSSA M.M., MAMATI E., REDA T. Evaluation of physiological and agronomic responses as screening techniques for yield and water stress tolerance in wheat Cultivars In Tigray Ethiopia. Academia Journal of Agricultural Research, 4 (1), 6, 2016.
- 50. YOUZHONG L., ZUJIAN Z., XINGYOU G., JIANCHANG Y., QINGSEN Z. Physiological and ecological effects of crimpy leaf character in rice (*Oryza sativa* L.) II. Photosynthetic character, dry mass production and yield forming. Zuo wu xue bao, 30 (9),883, 2004.
- CAI F., ZHANG Y., MI N., MING H., ZHANG S., ZHANG H., ZHAO X. Maize (*Zea mays* L.) physiological responses to drought and rewatering, and the associations with water stress degree. Agricultural Water Management, 241, 106379, 2020
- 52. CAL A.J., SANCIANGCO M., REBOLLEDO M.C., LUQUET D., TORRES R.O., MCNALLY K.L., HENRY A. Leaf morphology, rather than plant water status, underlies genetic variation of rice leaf rolling under drought. Plant, cell & environment, 42 (5), 1532, 2019.
- XU P., ALI A., HAN B., WU X. Current advances in molecular basis and mechanisms regulating leaf morphology in rice. Frontiers in plant science, 9, 1528, 2018.
- 54. SINGH S., SINGH T.N., CHAUHAN J.S. Living with limited water, part II: Dynamics of leaf rolling, leaf water

- homeostasis and water economy by hybrid rice. Journal of Crop Improvement, 25(4), 435, 2011.
- YUAN S., LI Y., PENG, SLeaf lateral asymmetry in morphological and physiological traits of rice plant. PloS one, 10 (6), 0129832, 2015.
- JUAREZ M.T., TWIGG R.W., TIMMERMANS M.C. Specification of adaxial cell fate during maize leaf development. Nature, 131, 4544, 2004.
- REBETZKE G.J., MORRISON A.D., RICHARD R.A., BONNETT D.G., MOORE C. Genotypic variation for leaf rolling in wheat. Wheat breeding society of Australia (eds), Mildura, 172, 2001.
- 58. SIRAULT X.R.R., CONDON A.G. WOOD J.T., FARQUHAR G.D., REBETZKE G.J. "Rolled-upness": phenotyping leaf rolling in cereals using computer vision and functional data analysis approaches. Plant Methods, 11 (1), 1, 2015.
- ZHANG C., YANG H., WU W., LI W. Effect of drought stress on physiological changes and leaf surface morphology in the blackberry. Brazilian Journal of Botany, 40, 625, 2017.
- TEE E.E. Journey and Destination: KORRIGAN1 Subcellular Localization Dynamically Changes during Plant Growth and Stress Tolerance. 32, 291, 2020.
- 61. ISLAM M.M., KAYESH E., ZAMAN E., URMI T.A., HAQUE M.M. Evaluation of rice (*Oryza sativa* L.) genotypes for drought tolerance at germination and early seedling stage. The Agriculturists, **16** (1), 44, **2018**.
- 62. BARET F., MADEC S., IRFAN K., LOPEZ J., COMAR A., HEMMERLÉ M., DUTARTRE D., PRAUD S., TIXIER M.H. Leaf-rolling in maize crops: from leaf scoring to canopy-level measurements for phenotyping. Journal of experimental botany, 69 (10), 2705, 2018.
- 63. BEN-AMAR A., BOUIZGAREN A., MAHBOUB S., NSARELLAH N. E., EL BOUHMADI K. Role of leaf rolling on agronomic performances of durum wheat subjected to water stress. African Journal of Agricultural Research, 16 (6), 791, 2020.
- 64. MATSCHI S., VASQUEZ M.F., BOURGAULT R., STEINBACH P., CHAMNESS J., KACZMAR N., GORE M.A., MOLINA I., SMITH L.G. Structure-function analysis of the maize bulliform cell cuticle and its potential role in dehydration and leaf rolling. Plant Direct, 4 (10), 00282, 2020.
- 65. ZOU L.P., SUN X.H., ZHANG Z.G., LIU P., WU J.X., TIAN C.J., QIU J.L., LU T.G. Leaf rolling controlled by the homeodomain leucine zipper class IV gene Roc5 in rice. Plant physiology, 156(3), 1602, 2011
- 66. FANG L., ZHAO F., CONG Y., SANG X., DU Q., WANG D., LI Y., LING Y., YANG Z., HE G. Rolling-leaf14 is a 2OG-Fe (II) oxygenase family protein that modulates rice leaf rolling by affecting secondary cell wall formation in leaves. Plant biotechnology journal, 10 (5), 524, 2012.
- 67. LI W.Q., ZHANG M.J., GAN P.F., QIAO L., YANG S.Q., MIAO H., WANG G.F., ZHANG M.M., LIU W.T., LI H.F., SHI C.H. CLD 1/SRL 1 modulates leaf rolling by affecting cell wall formation, epidermis integrity and water homeostasis in rice. The Plant Journal, 92 (5), 904, 2017.
- 68. TARATIMA W., RITMAHA T., JONGRUNGKLANG N., RASO S., MANEERATTANARUNGROJ, P. Leaf anatomical responses to drought stress condition in hybrid sugarcane leaf (*Saccharum officinarum* 'KK3'). Malaysian Applied Biology, 48 (3), 181, 2019.
- 69. LAFITTE R., BLUM A., ATLIN G. Using secondary traits to help identify drought-tolerant genotypes. Breeding rice for drought-prone environments, 37, 2003.

- NAR H., SAGLAM A., TERZI R.A.B.İ.Y.E., VARKONYI Z., KADIOGLU A. Leaf rolling and photosystem II efficiency in Ctenanthe setosa exposed to drought stress. Photosynthetica, 47, 429, 2009.
- SINGH B., REDDY K.R., REDONA E.D. WALKER T. Screening of rice cultivars for morpho-physiological responses to early-season soil moisture stress. Rice Science, 24 (6), 322, 2017.
- 72. ZHOU Z., FAN J., ZHANG J., YANG Y., ZHANG Y., ZAN X., LI X., WAN J., GAO X., CHEN R., HUANG Z. OsMLP423 Is a Positive Regulator of Tolerance to Drought and Salt Stresses in Rice. Plants, 11 (13), 1653, 2022.
- WERKER E. Trichome Diversity and Development-II. Trichome Variability and Classification. Advances in Botanical Research, 31, 4, 2000.
- DADA O., OHU, O. Growth and stomatal response of Amaranthus spp. to different watering regimes, organic and inorganic soil amendments. Nigerian Journal of Science, 47, 15, 2013.
- 75. MERSHON J.P., BECKER M., BICKFORD C.P. Linkage between trichome morphology and leaf optical properties in New Zealand alpine Pachycladon (*Brassicaceae*). New Zealand Journal of Botany, **53** (3), 175, **2015**.
- 76. HUCHELMANN A., BOUTRY M., HACHEZ C. Plant glandular trichomes: natural cell factories of high biotechnological interest. Plant physiology, 175 (1), 6, 2017.
- 77. CHEN J.J., SUN Y., KOPP K., OKI L., JONES S.B., HIPPS L. Effects of water availability on leaf trichome density and plant growth and development of Shepherdia× utahensis. Frontiers in Plant Science, 13, 855858, 2022.
- 78. KONRAD W., BURKHARDT J., EBNER M., ROTH-NEBELSICK A. Leaf pubescence as a possibility to increase water use efficiency by promoting condensation. Ecohydrology, 8 (3), 480, 2015.
- 79. FERNÁNDEZ V., SANCHO-KNAPIK D., GUZMÁN P., PEGUERO-PINA J.J., GIL L., KARABOURNIOTIS G., KHAYET M., FASSEAS C., HEREDIA-GUERRERO J.A., HEREDIA A., GIL-PELEGRÍN E. Wettability, polarity, and water absorption of holm oak leaves: effect of leaf side and age. Plant Physiology, 166 (1), 168, 2014.
- 80. BICKFORD C.P. Ecophysiology of leaf trichomes. Functional Plant Biology, **43** (9), 807, **2016**.
- 81. LI S., HAMANI A.K.M., ZHANG Y., LIANG Y., GAO Y., DUAN, A. Coordination of leaf hydraulic, anatomical, and economical traits in tomato seedlings acclimation to long-term drought. BMC Plant Biology, 21 (1), 1, 2021.
- 82. ZHOU H., HELLIKER B.R., HUBER M., DICKS A., AKÇAY E. C4 photosynthesis and climate through the lens of optimality. Proceedings of the National Academy of Sciences, 115 (47), 12057, 2018.
- 83. KOFIDIS G., BOSABALIDIS A.M., CHARTZOULAKIS K. Leaf anatomical alterations induced by drought stress in two avocado cultivars. Journal of Biological Research-Thessaloniki, 1, 115, 2004.
- 84. XU Z., ZHOU G. Responses of leaf stomatal density to water status and its relationship with photosynthesis in a grass. Journal of experimental botany, 59 (12), 3317, 2008.
- 85. BOSABALIDIS A.M., KOFIDIS G. Comparative effects of drought stress on leaf anatomy of two olive cultivars. Plant science, 163 (2), 375, 2002.
- 86. VEESAR N.F., JATOI W.A., GANDAHI N., AISHA G., SOLANGI, A.H., MEMON S. Evaluation of Cotton Genotypes for Drought Tolerance and Their Correlation Study at Seedling Stage. Biomedical Journal of Scientific & Technical Research, 29 (1), 22090, 2020.

87. SAMUELS L., KUNST L., JETTER, R. Sealing plant surfaces: cuticular wax formation by epidermal cells. Annu. Rev. Plant Biol., **59**, 683, **2008**.

- 88. BI H., KOVALCHUK N., LANGRIDGE P., TRICKER P.J., LOPATO S., BORISJUK N. The impact of drought on wheat leaf cuticle properties. BMC Plant Biology, 17,1, 2017.
- 89. JENKS M.A. Critical issues with the plant cuticle's function in drought tolerance. Biochemical and molecular responses of plants to the environment, **97**, 127, **2002**.
- 90. SEUFERT P., STAIGER S., ARAND K., BUENO A., BURGHARDT M., RIEDERER, M. X. Building a barrier: The influence of different wax fractions on the water transpiration barrier of leaf cuticles. Frontiers in plant science, 12, 766602, 2011.
- KOSMA D.K., JENKS M.A., Eco-physiological and molecular-genetic determinants of plant cuticle function in drought and salt stress tolerance. Advances in molecular breeding toward drought and salt tolerant crops, 91, 2007.
- 92. MCALLISTER T., CAMPOLI C., ESKAN M., LIU L., MCKIM S.M. A gene encoding a SHINE1/WAX INDUCER1 transcription factor controls cuticular wax in barley. Agronomy, 12 (5), 1088, 2022.
- 93. GORB E.V., KOZERETSKA I.A., GORB S.N. Hierachical epicuticular wax coverage on leaves of Deschampsia antarctica as a possible adaptation to severe environmental conditions. Beilstein Journal of Nanotechnology, 13 (1), 807, 2022.
- HASANUZZAMAN M., ZHOU M., SHABALA S. How does stomatal density and residual transpiration contribute to osmotic stress tolerance? Plants, 12 (3), 494, 2023
- KOSMA D.K., BOURDENX B., BERNARD A., PARSONS E.P., LU S., JOUBES J., JENKS M.A. The impact of water deficiency on leaf cuticle lipids of Arabidopsis. Plant physiology, 151 (4), 1918, 2009.
- SEO P.J., LEE, S.B. SUH M.C., PARK M.J., GO Y.S. PARK, C.M. The MYB96 transcription factor regulates cuticular wax biosynthesis under drought conditions in Arabidopsis. The Plant Cell, 23(3), 1138, 2011.
- AHARONI A., DIXIT S., JETTER R., THOENES E., VAN ARKEL G., PEREIRA A. The SHINE clade of AP2 domain transcription factors activates wax biosynthesis, alters cuticle properties, and confers drought tolerance when overexpressed in Arabidopsis. The Plant Cell, 16 (9), 2463, 2004.
- 98. ZHANG J.Y., BROECKLING C.D., BLANCAFLOR E.B., SLEDGE M.K., SUMNER L.W., WANG Z.Y. Overexpression of WXP1, a putative Medicago truncatula AP2 domain-containing transcription factor gene, increases cuticular wax accumulation and enhances drought tolerance in transgenic alfalfa (*Medicago sativa*). The Plant Journal, 42 (5), 689, 2005.
- LEE S.B., KIM H., KIM R.J., SUH M.C. Overexpression of Arabidopsis MYB96 confers drought resistance in *Camelina sativa* via cuticular wax accumulation. Plant Cell Reports, 33, 535, 2014.
- 100. GONZÁLEZ A., AYERBE L. Effect of terminal water stress on leaf epicuticular wax load, residual transpiration and grain yield in barley. Euphytica, **172** (3), 341, **2010**.
- NAWAZISH S., HAMEED M., NAURIN S. Leaf anatomical adaptations of Cenchrus ciliaris L., from the Salt Range, Pakistan against drought stress. Pakistan Journal of Botany, 38 (5), 1723, 2006.

- 102. NISHIYAMA T., SAKAYAMA H., DE VRIES J., BUSCHMANN H., SAINT-MARCOUX D., ULLRICH K.K., HAAS F.B., VANDERSTRAETEN L., BECKER D., LANG D., VOSOLSOBĚ S. The Chara genome: secondary complexity and implications for plant terrestrialization. Cell, 174 (2), 448, 2018.
- 103. SHELLAKKUTTI N., THANGAMANI P.D., SURESH K., BAALES J., ZEISLER-DIEHL V., KLAUS A., HOCHHOLDINGER F., SCHREIBER L., KRESZIES, T. Cuticular transpiration is not affected by enhanced wax and cutin amounts in response to osmotic stress in barley. Physiologia Plantarum, 174 (4), 13735, 2022.
- 104. YANG F., HAN Y., ZHU Q.H., ZHANG X., XUE F., LI Y., LUO H., QIN J., SUN J., LIU F. Impact of water deficiency on leaf cuticle lipids and gene expression networks in cotton (*Gossypium hirsutum* L.). BMC Plant Biology, 22 (1), 404, 2022.
- 105. CAMERON K.D., TEECE M.A., SMART L.B. Increased accumulation of cuticular wax and expression of lipid transfer protein in response to periodic drying events in leaves of tree tobacco. Plant physiology, 140 (1), 176, 2006.
- 106. SABHARWAL T., LU Z., SLOCUM R.D., KANG S., WANG H., JIANG H.W., VEERAPPA R., ROMANOVICZ D., NAM J.C., BIRK S., CLARK G. Constitutive expression of a pea apyrase, psNTP9, increases seed yield in field-grown soybean. Scientific Reports, 12(1), 10870, 2022.
- 107. KIM H., YU S.I., JUNG S.H., LEE B.H., SUH M.C. The F-box protein SAGL1 and ECERIFERUM3 regulate cuticular wax biosynthesis in response to changes in humidity in Arabidopsis. The Plant Cell, 31 (9), 2223, 2019.
- 108. BUCHANAN C.D., LIM S., SALZMAN R.A., KAGIAMPAKIS I., MORISHIGE D.T., WEERS B.D., KLEIN R.R., PRATT L.H., CORDONNIER-PRATT M.M., KLEIN P.E., MULLET J.E. Sorghum bicolor's transcriptome response to dehydration, high salinity and ABA. Plant molecular biology, 58, 699, 2005.
- 109. GHADIRNEZHAD SHIADE S.R., FATHI A., TAGHAVI GHASEMKHEILI F., AMIRI E., PESSARAKLI M. Plants' responses under drought stress conditions: Effects of strategic management approaches – A review. Journal of Plant Nutrition, 46 (9), 2198, 2023.
- 110. ROBERTSON B.C., HAN Y., LI C. A Comparison of Different Stomatal Density Phenotypes of Hordeum vulgare under Varied Watering Regimes Reveals Superior Genotypes with Enhanced Drought Tolerance. Plants, 12 (15), 2840, 2023.
- 111. HUGHES J., HEPWORTH C., DUTTON C., DUNN J.A., HUNT L., STEPHENS J., WAUGH R., CAMERON D.D., GRAY J.E. Reducing stomatal density in barley improves drought tolerance without impacting on yield. Plant physiology, 174 (2), 776, 2017.
- 112. CAINE R.S., YIN X., SLOAN J., HARRISON E.L., MOHAMMED U., FULTON T., BISWAL A.K., DIONORA J., CHATER C.C., COE R.A., BANDYOPADHYAY A. Rice with reduced stomatal density conserves water and has improved drought tolerance under future climate conditions. New Phytologist, 221 (1), 371, 2019.
- 113. DUNN J., HUNT L., AFSHARINAFAR M., MESELMANI M.A., MITCHELL A., HOWELLS R., WALLINGTON E., FLEMING A.J., GRAY J.E. Reduced stomatal density in bread wheat leads to increased water-use efficiency. Journal of Experimental Botany, 70 (18), 4737, 2019.

- 114. WEI Z., ABDELHAKIM L.O.A., FANG L., PENG X., LIU J., LIU F. Elevated CO₂ effect on the response of stomatal control and water use efficiency in amaranth and maize plants to progressive drought stress. Agricultural Water Management, 266, 107609, 2022.
- 115. CLEMENS M., FARALLI M., LAGREZE J., BONTEMPO L., PIAZZA S., VAROTTO C., MALNOY M., OECHEL W., RIZZOLI A., DALLA COSTA L. VvEPFL9-1 knock-out via CRISPR/Cas9 reduces stomatal density in grapevine. Frontiers in Plant Science, 13, 878001, 2022.
- 116. ZHANG L., ZHANG R., YE X., ZHENG X., TAN B., WANG W., LI Z., LI J., CHENG J., FENG J. Overexpressing VvWRKY18 from grapevine reduces the drought tolerance in Arabidopsis by increasing leaf stomatal density. Journal of Plant Physiology, 275, 153741, 2022.
- 117. YANG J., ZHANG N., BAI J., DUAN X., ZHANG L., LIU S., TANG X., JIN X., LI S. SI H. Stu-miR827-Targeted StWRKY48 Transcription Factor Negatively Regulates Drought Tolerance of Potato by Increasing Leaf Stomatal Density. International Journal of Molecular Sciences, 23 (23), 14805, 2022.
- 118. WANG Y., WAN L., ZHANG L., ZHANG Z., ZHANG H., QUAN R., ZHOU S., HUANG R. An ethylene response factor OsWR1 responsive to drought stress transcriptionally activates wax synthesis related genes and increases wax production in rice. Plant molecular biology, 78, 275, 2012.
- 119. ABDULLAH H.M., RODRIGUEZ J., SALACUP J.M., CASTAÑEDA I.S., SCHNELL D.J., PAREEK A., DHANKHER O.P. Increased cuticle waxes by overexpression of WSD1 improves osmotic stress tolerance in *Arabidopsis thaliana* and *Camelina sativa*. International journal of molecular sciences, 22 (10), 5173, 2021.
- 120. LEI Z.Y., HAN J.M., YI X.P., ZHANG W.F., ZHANG Y.L. Coordinated variation between veins and stomata in cotton and its relationship with water-use efficiency under drought stress. Photosynthetica, **56** (4), 1326, **2018**.
- 121. YU C., YAN C., LIU Y., LIU Y., JIA Y., LAVELLE D., AN G., ZHANG W., ZHANG L., HAN R., LARKIN R.M. Upregulation of a KN1 homolog by transposon insertion promotes leafy head development in lettuce. Proceedings of the National Academy of Sciences, 117(52), 33668, 2020.
- 122. ZHANG J., ZHANG H., SRIVASTAVA A.K., PAN Y., BAI J., FANG J., SHI H., ZHU J.K. Knockdown of rice microRNA166 confers drought resistance by causing leaf rolling and altering stem xylem development. Plant physiology, 176 (3), 2082, 2018.
- 123. ZHU X., WANG L., YANG R., HAN Y., HAO J., LIU C., FAN S. Effects of exogenous putrescine on the ultrastructure of and calcium ion flow rate in lettuce leaf epidermal cells under drought stress. Horticulture, Environment, and Biotechnology, 60, 479, 2019.
- 124. MCADAM S.A., SUSSMILCH F.C., BRODRIBB T.J. Stomatal responses to vapour pressure deficit are regulated by high speed gene expression in angiosperms. Plant, cell & environment, 39 (3), 485, 2016.
- 125. BUCKLEY T.N., JOHN G.P., SCOFFONI C., SACK L. The sites of evaporation within leaves. Plant Physiology, 173 (3), 1763, 2017.
- 126. NOLAN R.H., TARIN T., SANTINI N.S., MCADAM S.A., RUMAN R., EAMUS D. Differences in osmotic adjustment, foliar abscisic acid dynamics, and stomatal regulation between an isohydric and anisohydric woody

- angiosperm during drought. Plant, cell & environment, **40** (12), 3122, **2017**.
- 127. SANKAR B., KARTHISHWARAN K., SOMASUNDARAM R. Leaf anatomical changes in peanut plants in relation to drought stress with or without paclobutrazol and ABA. Journal of Phytology, **5**, 25, **2016**.
- 128. HASSIOTOU F., RENTON M., LUDWIG M., EVANS J.R., VENEKLAAS E.J. Photosynthesis at an extreme end of the leaf trait spectrum: how does it relate to high leaf dry mass per area and associated structural parameters?. Journal of Experimental Botany, **61** (11), 3015, **2010**.
- 129. TOMÁS M., FLEXAS, J., COPOLOVICI L., GALMÉS J., HALLIK L., MEDRANO H., RIBAS-CARBÓ M., TOSENS T., VISLAP V., NIINEMETS U. Importance of leaf anatomy in determining mesophyll diffusion conductance to CO₂ across species: quantitative limitations and scaling up by models. Journal of experimental botany, 64 (8), 2269, 2013.
- 130. MUIR C.D., HANGARTER R.P., MOYLE L.C., DAVIS P.A. Morphological and anatomical determinants of mesophyll conductance in wild relatives of tomato (S olanum sect. 1 ycopersicon, sect. 1 ycopersicoides; s olanaceae). Plant, Cell & Environment, 37 (6), 1415, 2014.
- 131. TOSENS T., NIINEMETS U., VISLAP V., EICHELMANN H., CASTRO DIEZ P. Developmental changes in mesophyll diffusion conductance and photosynthetic capacity under different light and water availabilities in Populus tremula: how structure constrains function. Plant, cell & environment, 35 (5),839, 2012.
- 132. HAN J., LEI Z., ZHANG Y., YI X., ZHANG W., ZHANG Y. Drought-introduced variability of mesophyll conductance in Gossypium and its relationship with leaf anatomy. Physiologia plantarum, 166 (3), 873, 2019.
- 133. ZHU J., CAI D., WANG J., CAO J., WEN Y., HE J., ZHAO L., WANG D., ZHANG S. Physiological and anatomical changes in two rapeseeds (*Brassica napus* L.) genotypes under drought stress conditions. Oil Crop Science, 6 (2), 97. 2021.
- 134. VOLLENWEIDER P., MENARD T., AREND M., KUSTER T.M., GÜNTHARDT-GOERG M.S. Structural changes associated with drought stress symptoms in foliage of Central European oaks. Trees, 30, 883, 2016.
- 135. FALCHI R., PETRUSSA E., BRAIDOT E., SIVILOTTI P., BOSCUTTI F., VUERICH M., CALLIGARO C., FILIPPI A., HERRERA J.C., SABBATINI P., ZANCANI M. Analysis of non-structural carbohydrates and xylem anatomy of leaf petioles offers new insights in the drought response of two grapevine cultivars. International Journal of Molecular Sciences, 21 (4), 1457, 2020.
- 136. HARAYAMA H., KITAO M., AGATHOKLEOUS E., ISHIDA A. Effects of major vein blockage and aquaporin inhibition on leaf hydraulics and stomatal conductance. Proceedings of the Royal Society B, 286 (1904), 20190799, 2019.
- 137. ABDELGAWAD H., AVRAMOVA V., BAGGERMAN G., VAN RAEMDONCK G., VALKENBORG D., VAN OSTADE X., GUISEZ Y., PRINSEN E., ASARD H., VAN DEN ENDE W., BEEMSTER G.T. Starch biosynthesis contributes to the maintenance of photosynthesis and leaf growth under drought stress in maize. Plant Cell & Environment, 43 (9), 2254. 2020.
- 138. ENNAJEH M., VADEL A. M., COCHARD H., KHEMIRA H. Comparative impacts of water stress on the leaf anatomy of a drought-resistant and a drought-sensitive

- olive cultivar. The Journal of Horticultural Science and Biotechnology, **85** (4), 289, **2010**.
- 139. EL-AFRY M. M. Anatomical studies on drought-stressed wheat plants (*Triticum aestivum* L.) treated with some bacterial strains. Acta Biologica Szegediensis, **56** (2), 165, **2012**.
- 140. MAKBUL S., GÜLER N.S., DURMUŞ N., GÜVEN S. Changes in anatomical and physiological parameters of soybean under drought stress. Turkish Journal of Botany, **35** (4), 369, **2011**.