Pol. J. Environ. Stud. Vol. 33, No. 4 (2024), 4169-4177
DOI: *10.15244/pjoes/176797* ONLINE PUBLICATION DATE: 2024-04-18

Original Research

Temporal Dynamics and DNA Barcoding of Hymenoptera from Juniper Forest Ecosystem

Qaiser Khan[1], Asmathullah Kakar[1], Shajahan Shabir Ahmed[2], Kashif Kamran[1], Muhammad Amjad Bashir[3*], Munaza Batool[5], Sagheer Atta[3,4], Reem A Alajmi[6]

[1]Department of Zoology, University of Balochistan, Quetta, Pakistan
[2]Department of Biotechnology, Balochistan University of Information technology, Engineering and management Sciences, Quetta, Pakistan.
[3]Department of Plant Protection faculty of Agricultural Sciences Ghazi University Dera Ghazi khan Punjab Pakistan
[4]United States Department of Agriculture Washington DC, USA
[5]Department of Soil & Environmental Sciences faculty of Agricultural Sciences Ghazi University Dera Ghazi khan Punjab Pakistan
[6]Department of Zoology, Faculty of Science, King Saud University, Riyadh, Saudi Arabia

Received: 13 November 2023
Accepted: 11 December 2023

Abstract

The study of Hymenoptera diversity and phenology holds significant importance for basic and applied scientific objectives. The present knowledge on the worldwide fauna diversity of Hymenoptera is good but exact data for Pakistan especially for the Hymenopterous fauna of Balochistan has not yet been updated. This geographical area presents an opportunity for the discovery of previously undocumented insect species. This study represents the first DNA barcode analysis in this research area, for a detailed picture of Hymenoptera. Out of the 8430 collected specimens, 810 insect specimens were morphologically identified as Hymenoptera, representing 50 species belonging to 11 families and 40 genera. These specimens were collected using a Malaise trap for 52 weeks, from 11th December 2018 to 10th December 2019. For further confirmation, the collected insect specimens were analyzed by sequencing cytochrome *c* oxidase subunit 1 (DNA barcode) and BINs were assigned to the Barcode of Life Data Systems. Based on molecular analysis, eight species representing three families and four genera were identified, and respective barcode index numbers (BINs) were assigned to them. Among them, four new species were recorded and two unique BINS (BOLD:AET0858 and BOLD:AET2316) were assigned to *Chrysis castillana* and *Anthophora quadrimaculata,* which had not previously been documented in the DNA barcode database. The most dominant species was *Camponotus compressus* (Fabricius, 1787), which was found throughout the year and had the highest mean population density. We explored the importance of employing harmonizing approaches including adult morphological characteristics and the DNA barcode method to accurately identify wild entomofauna for cryptic

*e-mail: qk.zoology@gmail.com

species. It is recommended that the use of Malaise traps spread over a large area is more beneficial in studying temporal dynamics and species recoveries.

Keywords: hymenoptera, juniper forest, morphological identification, DNA barcoding, malaise trap

Introduction

Hymenoptera represents the second-largest order of insects, with 155,000 known species [1]. In terms of human health concerns, Formicoidea (ants), Apoidea (bees), and Vespoidea (wasps) are the three most prominent superfamilies of this order [2]. The members of this order include parasitoid wasps, which regulate the abundance of host plants; predator wasps, which regulate the abundance of prey populations; and bees, which carry out the process of pollination. Therefore, these members play a significant role in maintaining the structure and function of the forest ecosystem [3]. They also influence the characteristics of modern terrestrial ecosystems [4]. For example, members of Hymenoptera exhibit a wide range of social behaviors, such as the solitary lifestyles of parasitic wasps, the simple family system of bumblebees, and the complex nest networks of supercolonial wood ants [5]. Hymenoptera accounts for the maximum number of parasitoid species, representing 75% of the total [6]. Within the Hymenoptera, a few species of the Terebrantia order use their ovipositor for egg-laying, while species from the Aculeata order use their modified ovipositors as stingers for injecting venom into prey or as defense mechanisms [2]. These features are ideal for understanding the evolutionary dynamics and cohesion of complex social groups of taxa.

The abundance and variety of species are influenced by multiple environmental factors such as climate, interaction among species, anthropogenic effects, etc. [7]. Seasonal variation in insect populations has been documented for various regions of the world, providing insight into the ecological processes that occur in specific ecosystems, particularly in dry tropical or temperate forest ecosystems [8, 9]. The pattern of temporal dynamics reflects the population size and richness of insect species [10] affected by biotic and abiotic factors. Abiotic factors such as temperature, rainfall, and humidity have a significant impact on the distribution of insect populations [11, 12]. For example, insect abundance is controlled by temperature in dry temperate zones [13] and by rainfall in tropical zones [14]. Biotic factors comprise several factors, including the availability of food, predation, parasitism, and the morphological structure of the host plant. All of these strongly influence insect abundance throughout the year [15]. The biodiversity of fauna within an ecosystem provides essential health indicators for that ecosystem [16]. For instance, species diversity ensures the natural sustainability of ecosystems and aids in their recovery from natural disasters [17].

The Malaise trap is a widely used entomological tool for the collection of arthropods on a large scale. This trap has a large tent-like structure made with fine mesh netting. It serves as a non-attractant and static insect trap [18]. This trap is usually used to capture flying insects, particularly Diptera and Hymenoptera, and it can also be used to collect various ground-dwelling species [19]. Several studies have established the effectiveness of these traps in capturing Hymenoptera in different South Asian countries, including India [20], Pakistan [21], and Iran [22].

Traditional morphological classification methods of insects remain relevant in taxonomy for describing the diversity of insects [23]. However, identifying species through this morphological method using a microscope proves challenging because it requires in-depth knowledge [24, 25] to differentiate closely related species [26]. The use of DNA-based methods in the identification of species has several advantages, such as the reproducibility of results [27, 28].

This technology is cost-effective, involves a simple protocol that takes only a few hours to complete, and can easily be applied to small animals [29]. Cytochrome oxidase I (COI) DNA barcoding is widely used as an alternative method for identification compared to the traditional morphological method [27]. In other words, this method is superior to other techniques for discriminating cryptic biodiversity [30]. For example, several cryptic species of butterflies have been identified via DNA barcode analysis that were morphologically similar and could not be distinguished at the species level through traditional morphological identification [31]. The current status of the Hymenopteran fauna is relatively robust in countries like Pakistan [32-35]. In addition, there are almost 5000 known species of insects in Pakistan [36]. However, the presence of Hymenoptera species in the Balochistan province of Pakistan has not been completely explored. A literature survey was carried out, which clearly indicated that only a few publications are available on Hymenoptera (wasp fauna) in the Quetta region of Balochistan, which covers only two species, *Polistes gallicus* and *Vespula germanica*, in the two subfamilies Polistinae and Vespinae [37]. Another two genera (i.e., P*oliste*: Latreille 1802, and *Ropalida*: Guerin-Meneville, 1831) of the subfamily Polistinae and one genus (i.e., V*espa: Linnae Vespinae*) were identified in Killa Saifullah (Northeastern Balochistan) [38].

The Ziarat juniper forests in Balochistan, Pakistan, cover a mountainous area extending from an altitude of 1,181 to 3,488 meters. This forest is the major juniper forest in Pakistan, encompassing an area of 110 thousand hectares [39, 40]. A unique ecosystem of fauna and flora has been observed within our study area, i.e., the juniper forest of Ziarat. This forest has been declared a ‘Ziarat

Juniper Forest Biosphere Reserve' by the United Nations Educational, Scientific, and Cultural Organization (UNESCO, 2013). Certain areas within this ecosystem are safeguarded, thus providing refuge for endangered wildlife and protecting the surrounding variety of flora. This is done by establishing wildlife reserves and game parks that protect the habitats from any potentially anthropogenic activities. The insect biodiversity of this area based on molecular identification has not been fully documented. In this study, we combined traditional morphological and DNA barcoding methods using the mitochondrial marker cytochrome c oxidase subunit I (COI) gene. We also studied the temporal dynamics of the Hymenoptera species population in this region.

Materials and Methods

Study Area

The present study was conducted in the juniper forest of Ziarat District (30°22'51N and 67°43'37S), Balochistan province. This district covers an area of 1487 km2 and is situated at an altitude of an average 2454 m (Government of Balochistan, 2011). It is located 70 km from Quetta, the capital of Balochistan province [41].

Malaise Trap Installation

A single Malaise trap made of knitted polyester mesh was used for the present study. This trap had specific dimensions, i.e., length 165 cm, width 80 cm, and height 180 cm. The size of the mesh opening was 96 cm x 26 cm, and the net weight of this trap was 870 g. One face of the tent was left open, and a single Nalgene® insect-collecting bottle filled with 95% ethanol was placed inside the tent. A funnel was used to channel the insects into the bottle via vertical screens or curtains, which intercepted and facilitated flying insects during their attempts to escape. In December 2018, this trap was installed at Sandaman Tangi (30°24'00.5"N, 67°43'36.5"E), a village of the Union Council of Ziarat with an altitude of 2450 m. Malaise trap installation is shown in Fig. 1.

Sampling and Preservation of Samples

Samples were collected on weekends for 52 weeks, from December 2018 to December 2019. Samples were collected in a 500 mL plastic Nalgene® bottle containing 400 ml ethanol (95%) and then transferred into a Whirl- Pak bag® containing 95% ethanol. The dates of collection were marked on bags, and these specimens were then brought to the Entomology Laboratory, Department of Zoology, at the University of Balochistan for their morphological identification. The Hymenoptera insects were sorted from the samples, and large insects were pinned in an entomological box,

Fig. 1. Malaise trap with an insect-collecting bottle installed at Sundaman Tangi.

while the remaining small insects were left in 95% ethanol for morphological identification using available taxonomic references. From the collected samples, alternate specimens for each species were stored in ethanol at -20ºC before and later shifted to the Center for Biodiversity Ggenomic (CBG) in Canada for DNA barcoding.

DNA Extraction and PCR Amplification and Sequencing

DNA extraction, polymerase chain reaction (PCR), and DNA sequencing were performed at the Canadian Center for DNA Barcoding (CCBD). One or two legs of adult morphologically identified insects were used for DNA extraction using standard protocol, and their respective vouchers were recovered for imaging and curation [27, 42]. The cytochrome c oxidase subunit 1 (CO1) was amplified using forward and reverse primers in PCR which are LepFoIF (ATTCAACCAATCATAAAGATATTGG) and LepFoIR (TAAACTTCTGGATGTCCAAAAAATCA) [43].

Extracted DNA samples and the whole mount of morphologically identified insects were sent to the Canadian Centre for DNA Barcoding (CCDB) (http://ccdb.ca/resources.php) for DNA barcoding following the specified barcoding procedures [44, 45] and then curated at the Center for Biodiversity Genomic for sequencing. The data of each Hymenoptera species regarding DNA sequences, voucher evidence, and taxon information were deposited for public access in the Barcode of Life Data Systems (BOLD) (https://www.boldsystems.org/).

Data Analysis

The sequence generation was possible only for 8 out of 50 insects. The Barcode Index Numbers (BINs) were assigned to seven families of the order Hymenoptera. The sequences, along with BINs and other related taxonomic information, were uploaded to the Barcode of Life Data System (BOLD) (http://www.boldsystems.

org/) following standard protocol [45]. These sequences were then downloaded from BOLD for comparative analysis with NCBI (National Center for Biotechnology Information) (https://www.ncbi.nlm.nih.gov/) using BLAST (Basic Local Alignment Search Tool). These sequences were aligned using the BioEdit alignment editor (version 7.0.5). We provided a direct link to the original sequence used for identification and stored in GenBank (NCBI)

SUB13919971 Anth_cing_1	OR724646
SUB13919971 Ichne_sarc_1	OR724647
SUB13919971 Pol_gal_1	OR724648
SUB13919971 Cata_aen_1	OR724649
SUB13919971 chrys_1	OR724650
SUB13919971 Athoph_1	OR724651
SUB13919971 campo_1	OR724652

Results and Discussion

Temporal Dynamics of Hymenoptera

Out of the total 8430 insects, only 50 were identified morphologically as belonging to Hymenoptera. It is evident from Fig. 2 and Fig. 3 that the maximum mean population of Hymenoptera occurred during July and August, while the minimum mean population was recorded in January, February, March, and

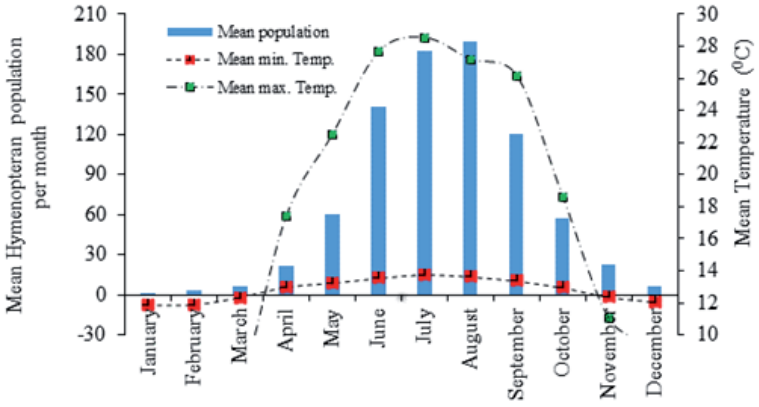

Fig. 2. Mean Hymenoptera population sample.

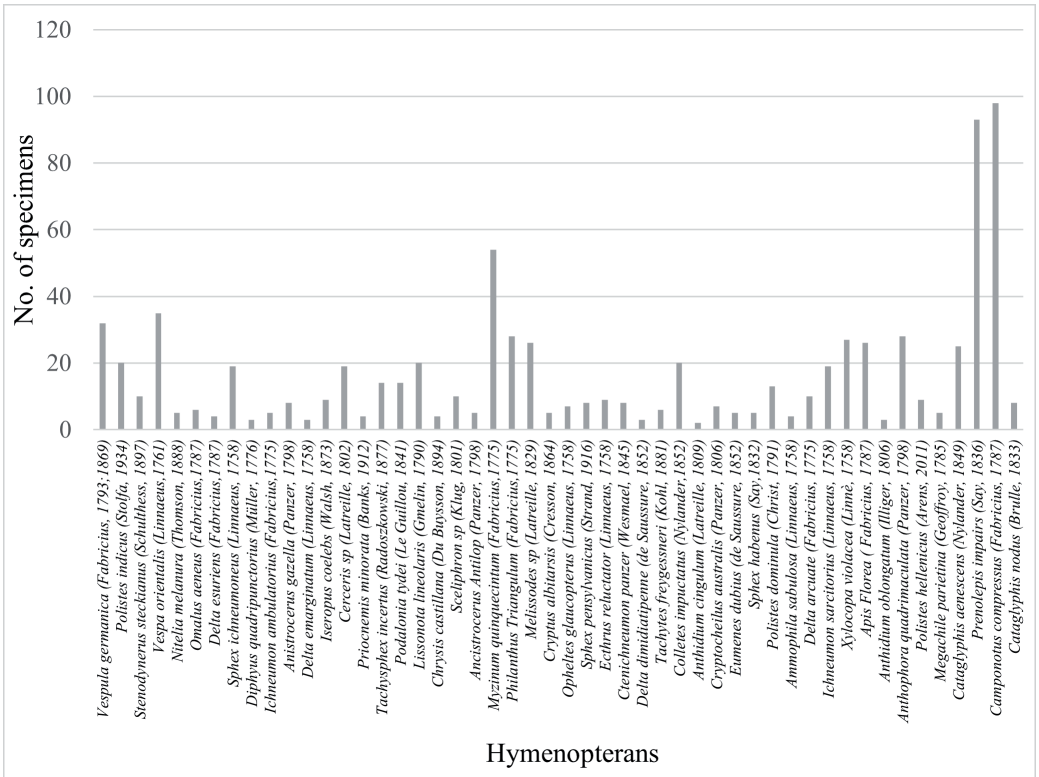

Fig. 3. Temporal dynamics of the Hymenoptera species sample.

Table 1. Identification of Hymenoptera insect species through the DNA barcoding method and their close matching results are presented in the NCBI database (%).

Sample ID	Family	Species	Close match with NCBI database (%)
QBZS 19	Chrysididae	*Chrysis castillana*	88.24
QBZS 33	Megachilidae	*Anthidium cingulum*	98.26
QBZS 40	Ichneumonidae	*Ichneumon sarcitorius*	99.51
QBZS 44	Apidae	*Anthophora quadrimaculata*	93.44
QBZS 45	Vespidae	Polistes hellenicus	98.55
QBZS 46	Megachilidae	*Megachile parietina*	92.24
QBZS 47	Formicidae	*Cataglyphis aenescens*	96.66
QBZS 48	Formicidae	*Camponotus* spp.	92.11

December (Table 1). It has been observed that the average population of Hymenoptera follows a Gaussian distribution pattern. The average monthly atmospheric temperature increases from May to September, leading to an increase in the insect population. A decline in the population of Hymenoptera between January-April and October-December is linked to a fall in temperature during these periods (Fig. 2). The monthly Hymenoptera population plotted as a function of temperature is shown in Fig. 3, which indicates exponential growth. A total of 50 insects of Hymenoptera were identified from the samples, and these belonged to 11 families and 40 genera (Fig. 3). These families included Apidae, Carabornidae, Chrysididae, Colletidae, Ichneumonidae, Formicidae, Megachilidae, Pompilidae, Sphecidae, Thynnidae, and Vespidae. The Ichneumonidae family was the most dominant and accounted for 10 genera and 11 species, followed by the Vespidae family, representing 7 genera and 13 species. The Thynnidae and Colletidae families were represented only by single genera and species. The species population was the highest during June-September and January-May while the population was extremely low from October-December due to low atmospheric temperatures. The most common species was *Camponotus compressus* (Fabricius, 1787), which reached its peak population during June-July, respectively, followed by Prenolepis impairs (Say, 1836), which was observed in all months except January with a maximum population in July. Anthidium cingulum (Latreille, 1809) had its lowest mean population during July-August while Delta dimidiatipenne (de Saussure, 1852) had its lowest mean population during August-September.

DNA Barcoding

The DNA barcodes were generated only for eight species out of the 50 morphologically identified specimens (Fig. 4 and Fig. 5). *Ichneumon sarcitorius* showed the highest close match (99.51%) within the NCBI database, followed by *Polistes hellenicus* (98.55%), *Anthidium cingulum* (98.26%), *Cataglyphis aenescens* (96.66%), *Anthophora quadrimaculata* (93.44%), *Megachile parietina* (92.24%), *Camponotus* spp. (92.11%) and *Chrysis castillana* (88.24%) as shown in Table 1. Barcodes were assigned to three families, four genera, and four species based on their sequence divergence between Hymenoptera taxa and their barcode index numbers (BINs) (Table 2). Two unique BINs were assigned to the Chrysididae family (BOLD:AET0858), one to the Formicidae family (BOLD:AET2316) and one to *Anthophora quadrimaculata* (BOLD:AET5134). These unique BINS can offer a structured process of categorizing groups of genetically identical taxa, allowing the same taxa featured in different examinations to be labeled with a common identifier. There were no DNA barcodes in the rest of the specimens, possibly due to the contamination of samples and resulting degradation of DNA.

In this study, the Hymenoptera population was found to be most abundant during June-September due to the relatively high atmospheric temperature. Such results have also been reported in earlier studies [46, 47]. The abundance and species richness of bees have been

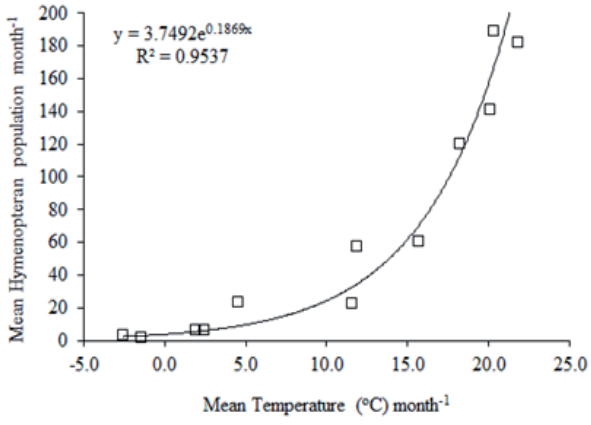

Fig. 4. Exponential correlation between mean population month-1 and mean temperature month-1 of Hymenoptera.

Table 2. Sample ID of eight Hymenoptera, taxa, p-distance (%), member count, barcode index numbers (BINs) assigned, and their comparison to the nearest BINs in the BOLD database.

Sample ID	Taxa	*p*-distance (%)			Member count	BINs
		Mean	Maximum	Nearest neighbor (NN)		
QBZS 19	Chrysididae	N/A	N/A	10.57	1	BOLD:AET0858
QBZS 33	Megachilidae	0.43	0.49	1.61	3	BOLD:AEH2998
QBZS 40	Ichneumon sarcitorius	0.45	1.92	3.41	19	BOLD:AAN3371
QBZS 44	*Anthophora quadrimaculata*	N/A	N/A	6.04	1	BOLD:AET2316
QBZS 45	*Polistes*	0.92	2.73	2.62	75	BOLD:AAN3303
QBZS 46	*Megachile*	0.8	0.8	7.06	2	BOLD:AAK7027
QBZS 47	Formicidae	N/A	N/A	3.53	1	BOLD:AES3790
QBZS 48	Formicidae	N/A	N/A	3.05	1	BOLD:AET5134

* Bold caption indicates the unique BIN id.

reported during June-August [48]. Temperature, rainfall, and humidity are the main meteorological drivers that cause seasonal variations, which affect the population of insects [49]. Insect abundance is also influenced by temporal changes, which is a common spectacle in any ecosystem [50]. Other factors that control insect populations during a particular season include the unavailability of food, the dominance of insect predators and parasites, the competition for resources, and habitat destruction [51, 52].

Among the morphologically identified 50 species, two species, *Camponotus compressus* (Fabricius, 1787) and *Prenolepis impairs* (Say, 1836), were recorded throughout the year. Both species showed the highest individual abundance over the entire study period [53]. *Camponotus compressus* is considered a significant economic pest [54] and widely known as a 'carpenter's pest' [55, 56] because of its nesting behavior, which causes damage to buildings. [57] also studied the temporal dynamics of ants and found that

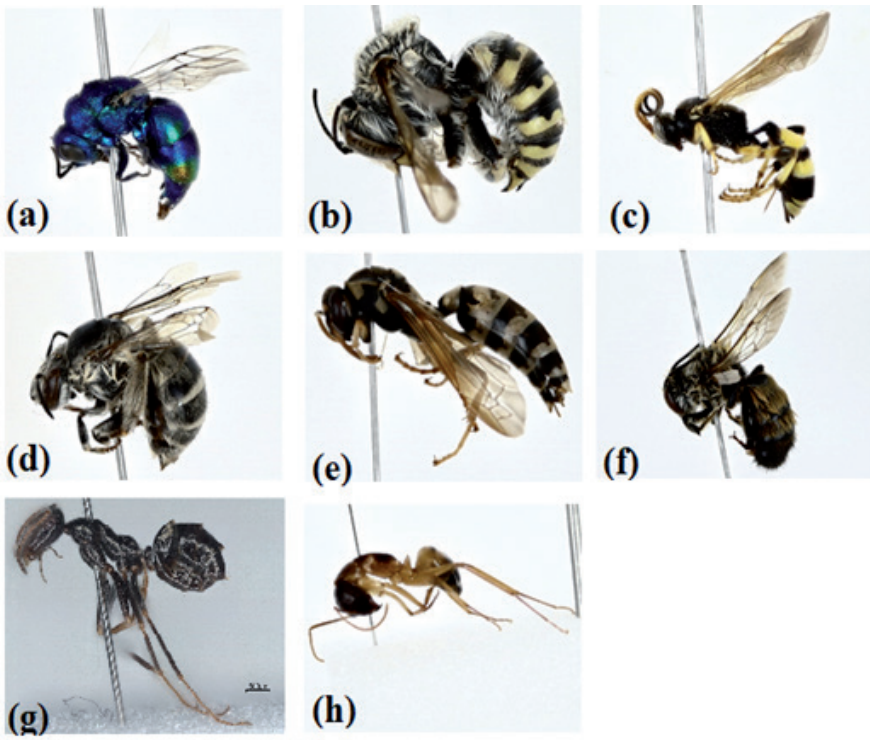

Fig. 5. Identified species of Hymenoptera through DNA barcoding. a) *Chrysis castillana*, b) *Anthidium cingulum,* c) *Ichneumon sarcitorius*, d) *Anthophora quadrimaculata*, e) *Polistes hellenicus*, f) *Megachile parietina,* g) *Cataglyphis aenescens* and h) *Camponotus* spp.

seasonal behavior was evident in most of their species. On the contrary, *Prenolepis impairs* (Say, 1836), known as the 'winter ant', remains active, foraging across the year, even in the lowest temperature period of the year. This species is more tolerant at low temperatures than other ant species [58, 59]. Our findings are consistent with those of [60], who have also reported the abundances of temporally dynamic of species of these insects.

Malaise traps are mostly preferred for the sampling of flying insects, particularly the Diptera and Hymenoptera, due to their effectiveness [19]. It has been a common practice in insect surveillance studies to use the Malaise trap as a standardized method for species collection and to investigate both the temporal and spatial dynamics of insect populations, as well as the identification of species using DNA barcoding methods [18, 61, 62]. It was reported that *Psilochalcis minuta* in the Juniper ecosystem using a Malaise trap was the most abundant species during July-August [63].

The taxonomical identification of species relies heavily on DNA sequencing. Only eight sequences and their relevant barcode index numbers (BINs) were generated. Among the assigned BINs, four species were unique, for which there was a lack of barcode databases, which indicates the possibility of a new species. In the IBOL (International Barcode of Life) database, all species codes (from QBZS1 to QBZS50) are displayed along with their photos of 50 morphologically identified species. Only eight species have a standard barcode, while barcodes for 42 species were not possible either due to non-amplification or produced sequences that were difficult to interpret, which might be due to contamination or damage to DNA after the extraction process. Another reason was that those specimens nucleotide sequences higher than 500bp obscured the others showing lower nucleotide sequence values. [64] barcoded 50,094 out of 60,273 specimens collected from Pakistan for DNA sequencing of insect biodiversity in a surveillance study, where DNA barcodes of 17% specimens were not generated. Various research groups have also reported such differences in the recovery of DNA sequences across varied insect taxa [65, 66]. Among the assigned BINs, four species were unique, indicating the possibility of new species.

The earlier published literature has shown that operational taxonomic units (OTUs) assigned to morphologically unidentifiable species are less frequently criticized and accepted to some extent [67, 68]. The BIN system was developed to overcome these constraints [69]. This system has been applied to diverse groups of animals to discriminate between species as well as discover new species [70]. However, it is not an easy tool for morphological analysis and requires further scrutiny and thorough understanding before it could be applied to evolutionary and lineage studies [71].

Conclusion

We have concluded from our study that the Juniper Forest ecosystem is rich in Hymenoptera species. The overall population of Hymenoptera varies with temperature fluctuations throughout the year and exhibits a Gaussian distribution. Hymenoptera abundance showed an increase with a rise in mean temperature, as indicated by the exponential correlation value. Furthermore, we concluded that the combination of traditional morphological and DNA barcoding procedures is useful for investigating Hymenopteran insect species. Malaise traps spread over large areas have proved to be more beneficial and should be installed to study temporal dynamics and species recoveries in Juniper ecosystems.

Acknowledgments

The authors especially Reem Atalla Alajmi extend appreciation to Researchers Supporting Project (RSP-2024/R99) King Saud University, Riyadh, Saudi Arabia for the study. We are thankful to Prof. Dr. Muhammad Ashfaq Ahmed, University of Balochistan Quetta for revision of this manuscript.

Conflict of Interest

All authors have no conflicting interests.

References

1. AGUIAR A.P., DEANS A.R., ENGEL M.S., FORSHAGE M., HUBER J.T., JENNINGS J.T., JOHNSON N.F., LELEJ A.S., LONGINO J.T., LOHRMANN V. Order Hymenoptera. In: Zhang, Z.-Q.(Ed.) Animal biodiversity: an outline of higher-level classification and survey of taxonomic richness (Addenda 2013). Zootaxa, **3703** (1), 51, **2013**.
2. SHORTER J., RUEPPELL O. A review on self-destructive defense behaviors in social insects. Insectes sociaux, **59** (1), 1, **2012**.
3. ARCHIBALD S., RASNITSYN A.P., BROTHERS D.J., MATHEWES R.W. Modernisation of the Hymenoptera: ants, bees, wasps, and sawflies of the early Eocene Okanagan Highlands of western North America. The Canadian Entomologist, **150** (2), 205, **2018**.
4. RESH V.H., CARDÉ R.T. Encyclopedia of insects. Academic press, **2009**.
5. SCHULTNER E., SARAMÄKI J., HELANTERÄ H. Genetic structure of native ant supercolonies varies in space and time. Molecular Ecology, **25** (24), 6196, **2016**.
6. BELSHAW R. Life history characteristics of Tachinidae (Diptera) and their effect on polyphagy. Parasitoid community ecology, 145, **1994**.
7. RAHMAN A.U., KHAN S.M., AHMAD Z., ALAMRI S., HASHEM M., ILYAS M., AKSOY A., DÜLGEROĞLU C., SHAHAB ALI G.K. -Impact of multiple environmental factors on species abundance in various forest layers using an integrative modeling approach. Global Ecology and Conservation, **29** e01712, **2021**.

8. GILLIAM F.S. Forest ecosystems of temperate climatic regions: from ancient use to climate change. New Phytologist, **212** (4), 871, **2016**.
9. HALSCH C.A., SHAPIRO A.M., FORDYCE J.A., NICE C.C., THORNE J.H., WAETJEN D.P., FORISTER M.L. Insects and recent climate change. Proceedings of the national academy of sciences, **118** (2), e2002543117, **2021**.
10. PINHEIRO F., DINIZ I., COELHO D., BANDEIRA M. Seasonal pattern of insect abundance in the Brazilian cerrado. Austral Ecology, **27** (2), 132, **2002**.
11. AMJAD BASHIR M., BATOOL M., KHAN H., SHAHID NISAR M., FAROOQ H., HASHEM M., ALAMRI S., A. EL-ZOHRI M., ALAJMI R.A., TAHIR M., JAWAD R. Effect of temperature & humdity on population dynamics of insects' pest complex of cotton crop. PLOS ONE, **17** (5), e0263260, **2022**.
12. CHEN C., HARVEY J.A., BIERE A., GOLS R. Rain downpours affect survival and development of insect herbivores: the specter of climate change? Ecology, **100** (11), e02819, **2019**.
13. SKENDŽIĆ S., ZOVKO M., ŽIVKOVIĆ I.P., LEŠIĆ V., LEMIĆ D. The impact of climate change on agricultural insect pests. Insects, **12** (5), 440, **2021**.
14. MORRIS W.F., EHRLÉN J., DAHLGREN J.P., LOOMIS A.K., LOUTHAN A.M. Biotic and anthropogenic forces rival climatic/abiotic factors in determining global plant population growth and fitness. Proceedings of the National Academy of Sciences, **117** (2), 1107, **2020**.
15. RICHARDS L.A., COLEY P.D. Seasonal and habitat differences affect the impact of food and predation on herbivores: a comparison between gaps and understory of a tropical forest. Oikos, **116** (1), 31, **2007**.
16. HILTY J., MERENLENDER A. Faunal indicator taxa selection for monitoring ecosystem health. Biological conservation, **92** (2), 185, **2000**.
17. SEDDON N., TURNER B., BERRY P., CHAUSSON A., GIRARDIN C.A. Grounding nature-based climate solutions in sound biodiversity science. Nature Climate Change, **9** (2), 84, **2019**.
18. SKVARLA M.J., LARSON J.L., FISHER J.R., DOWLING A.P. A review of terrestrial and canopy malaise traps. Annals of Entomological Society of America, **114** (1), 27, **2021**.
19. VAN ACHTERBERG K. Can Townes type Malaise traps be improved? Some recent developments. Entomologische berichten, **69** (4), 129, **2009**.
20. SHWETA M., RAJMOHANA K. A comparison of sweep net, yellow pan trap and malaise trap for sampling parasitic Hymenoptera in a backyard habitat in Kerala. Entomon, **43** (1), 43, **2018**.
21. ASHFAQ M., SABIR J.S., EL-ANSARY H.O., PEREZ K., LEVESQUE-BEAUDIN V., KHAN A.M., RASOOL A., GALLANT C., ADDESI J., HEBERT P.D. Insect diversity in the Saharo-Arabian region: Revealing a little-studied fauna by DNA barcoding. PLoS. ONE, **13** (7), e0199965, **2018**.
22. ABDOLI P., TALEBI A.A., FARAHANI S., FERNANDEZ-TRIANA J. Three new species of the genus Choeras Mason, 1981 (Hymenoptera: Braconidae, Microgastrinae) from Iran. Zootaxa, **4545** (1), 77, **2019**.
23. MIDGLEY J.M., VILLET M.H. Metrological framework for selecting morphological characters to identify species and estimate developmental maturity of forensically significant insect specimens. Forensic Sciences Research, **6** (1), 75, **2021**.
24. BAILET B., BOUCHEZ A., FRANC A., FRIGERIO J.-M., KECK F., KARJALAINEN S.-M., RIMET F., SCHNEIDER S., KAHLERT M. Molecular versus morphological data for benthic diatoms biomonitoring in Northern Europe freshwater and consequences for ecological status. Metabarcoding and Metagenomics, **2019**.
25. ZIMMERMANN J., GLÖCKNER G., JAHN R., ENKE N., GEMEINHOLZER B. Metabarcoding vs. morphological identification to assess diatom diversity in environmental studies. Molecular ecology resources, **15** (3), 526, **2015**.
26. ZHAN J., ZHENG Y., XIA Q., WANG J., LIU S., YANG Z. Diversity investigation by application of DNA barcoding: A case study of lepidopteran insects in Xinjiang wild fruit forests, China. Ecology and evolution, **12** (3), e8678, **2022**.
27. HEBERT P.D., CYWINSKA A., BALL S.L., DEWAARD J.R. Biological identifications through DNA barcodes. Proceedings of the Royal Society of London. Series B: Biological Sciences, **270** (1512), 313, **2003**.
28. LIN X.-L., STUR E., EKREM T. DNA barcodes and morphology reveal unrecognized species in Chironomidae (Diptera). Insect Systematics & Evolution, **49** (4), 329, **2018**.
29. ROUGERIE R., DECAËNS T., DEHARVENG L., PORCO D., JAMES S.W., CHANG C.-H., RICHARD B., POTAPOV M., SUHARDJONO Y., HEBERT P.D. DNA barcodes for soil animal taxonomy. Pesquisa Agropecuária Brasileira, **44** 789, **2009**.
30. HEBERT P.D., RATNASINGHAM S., ZAKHAROV E.V., TELFER A.C., LEVESQUE-BEAUDIN V., MILTON M.A., PEDERSEN S., JANNETTA P., DEWAARD J.R. Counting animal species with DNA barcodes: Canadian insects. Philosophical Transactions of the Royal Society B: Biological Sciences, **371** (1702), 20150333, **2016**.
31. BURNS J.M., JANZEN D.H., HAJIBABAEI M., HALLWACHS W., HEBERT P.D. DNA barcodes and cryptic species of skipper butterflies in the genus Perichares in Area de Conservacion Guanacaste, Costa Rica. Proceedings of the National Academy of Sciences, **105** (17), 6350, **2008**.
32. INAYATULLAH M. To the knowledge of Vespidae (Hymenoptera) of Pakistan. Zootaxa, **3318** 26, **2012**.
33. SIDDIQUI J.A., BODLAH I., CARPENTER J.M., NAEEM M., AHMAD M., BODLAH M.A. Vespidae (Hymenoptera) of the Pothwar region of Punjab, Pakistan. Zootaxa, **3914** (5), 501, **2015**.
34. ARSALAN M., ABBAS A., GUL S.U., REHMAN H.U., JAWAD S.M., SHAH W., MEHMOOD A. Biodiversity of wasps species collected from district Karak, KP, Pakistan. Journal of Entomology and Zoology Studies, **6** (2), 21, **2018**.
35. KHAN A.K., KHAN M.R., RAFI M.A., QASIM M. Wasp fauna of (Eumeninae, vespinae and polistinae) of district Poonch, Azad Jammu and Kashmir (Pakistan). Journal of Entomology and Zoology Studies, **5** (6), 1587, **2017**.
36. GOVERNMENT OF PAKISTAN Government of Pakistan Biodiversity action plan for Pakistan. **2000**.
37. DVOŘÁK L. The social wasp fauna of Pakistan (Hymenoptera, Vespidae). Linzer Biologische Beiträge, **39**, **2007**.
38. NAZ F., KAKAR A., NASIM M., MOHAMMAD W., KHAN A., NASEEB N. 109. Faunal diversity of paper wasp species and oriental hornet from tehsil Killa Saifullah of Zhob division, Pakistan. Pure and Applied Biology (PAB), **9** (1), 1148, **2020**.
39. JALLAT H., KHOKHAR M.F., KUDUS K.A., NAZRE M., SAQIB N.U., TAHIR U., KHAN W.R. Monitoring carbon stock and land-use change in 5000-year-old juniper

forest stand of Ziarat, Balochistan, through a synergistic approach. Forests, **12** (1), 51, **2021**.
40. GOVERNMENT OF PAKISTAN D.G.O.A. Ziarat Juniper Forest. **2016**.
41. KAMRAN K., AKBAR A., NASEEM M., SAMAD A., ACHAKZAI J.K., REHMAN Z.U., SAJID M.S., ALI A. Participatory appraisal for healthcare and welfare management strategies of donkeys (*Equus ascinus*) in Balochistan, Pakistan. Frontiers in Veterinary Science, **9 2022**.
42. PORCO D., ROUGERIE R., DEHARVENG L., HEBERT P. Coupling non-destructive DNA extraction and voucher retrieval for small soft-bodied Arthropods in a high-throughput context: the example of Collembola. Molecular Ecology Resources, **10** (6), 942, **2010**.
43. SMITH M.A., FISHER B.L. Invasions, DNA barcodes, and rapid biodiversity assessment using ants of Mauritius. Frontiers in zoology, **6** (1), 1, **2009**.
44. DEWAARD J.R., LEVESQUE-BEAUDIN V., DEWAARD S.L., IVANOVA N.V., MCKEOWN J.T., MISKIE R., NAIK S., PEREZ K.H., RATNASINGHAM S., SOBEL C.N. Expedited assessment of terrestrial arthropod diversity by coupling Malaise traps with DNA barcoding. Genome, **62** (3), 85, **2019**.
45. DEWAARD J.R., RATNASINGHAM S., ZAKHAROV E.V., BORISENKO A.V., STEINKE D., TELFER A.C., PEREZ K.H., SONES J.E., YOUNG M.R., LEVESQUE-BEAUDIN V. A reference library for Canadian invertebrates with 1.5 million barcodes, voucher specimens, and DNA samples. Scientific data, **6** (1), 1, **2019**.
46. YADAV N., SINGH P. Seasonal abundance of insect pests on mung bean and its correlation with abiotic factors. Journal of Entomological Research, **37** (4), 297, **2013**.
47. SCHWARZ B., VÁZQUEZ D.P., CARADONNA P.J., KNIGHT T.M., BENADI G., DORMANN C.F., GAUZENS B., MOTIVANS E., RESASCO J., BLÜTHGEN N. Temporal scale-dependence of plant–pollinator networks. Oikos, **129** (9), 1289, **2020**.
48. OERTLI S., MÜLLER A., DORN S. Ecological and seasonal patterns in the diversity of a species-rich bee assemblage (Hymenoptera: Apoidea: Apiformes). European Journal of Entomology, **102** (1), 53, **2005**.
49. SILVA N.A.P.D., FRIZZAS M.R., OLIVEIRA C.M.D. Seasonality in insect abundance in the" Cerrado" of Goiás State, Brazil. Revista Brasileira de Entomologia, **55**, 79, **2011**.
50. ADAMS B.J., LI E., BAHLAI C.A., MEINEKE E.K., MCGLYNN T.P., BROWN B.V. Local- and landscape-scale variables shape insect diversity in an urban biodiversity hot spot. Ecological Applications, **30** (4), e02089, **2020**.
51. MARCHIORO C.A., FOERSTER L.A. Biotic factors are more important than abiotic factors in regulating the abundance of Plutella xylostella L., in Southern Brazil. Revista Brasileira de Entomologia, **60** (4), 328, **2016**.
52. DAR S.A., ANSARI M.J., AL NAGGAR Y., HASSAN S., NIGHAT S., ZEHRA S.B., RASHID R., HASSAN M., HUSSAIN B. Causes and Reasons of Insect Decline and the Way Forward. IntechOpen, **2021**.
53. ABDAR M. Seasonal Abundance and Commonly Occurring Household Ants Species in Sangli District Maharashtra. Research Journal of Agricultural Sciences, **11** (6), 1413, **2020**.
54. KHOT K., QUADROS G., SOMANI V. Ant diversity in an urban garden at Mumbai, Maharashtra. **2013**.
55. LEE C.Y., TAN E.K. Guide to urban pest ants in Singapore. Singapore Pest Management Association, **2004**.
56. CHAVHAN A., PAWAR S. Distribution and diversity of ant species (Hymenoptera: Formicidae) in and around Amravati City of Maharashtra, India. World journal of Zoology, **6** (4), 395, **2011**.
57. BASU P. Seasonal and spatial patterns in ground foraging ants in a rain forest in the Western Ghats, India 1. Biotropica, **29** (4), 489, **1997**.
58. TSCHINKEL W.R. Seasonal life history and nest architecture of a winter-active ant, Prenolepis imparis. Insectes Sociaux, **34** (3), 143, **1987**.
59. WILLIAMS J.L., LAPOLLA J.S. Taxonomic revision and phylogeny of the ant genus Prenolepis (Hymenoptera: Formicidae). Zootaxa, **4200** (2), 1, **2016**.
60. D'SOUZA M.L., HEBERT P.D. Stable baselines of temporal turnover underlie high beta diversity in tropical arthropod communities. Molecular ecology, **27** (10), 2447, **2018**.
61. AGUIAR A.P., SANTOS B.F. Discovery of potent, unsuspected sampling disparities for Malaise and Möricke traps, as shown for Neotropical Cryptini (Hymenoptera, Ichneumonidae). Journal of Insect Conservation, **14** (2), 199, **2010**.
62. BUCKLEY L.B., TEWKSBURY J.J., DEUTSCH C.A. Can terrestrial ectotherms escape the heat of climate change by moving? Proceedings of the Royal Society B: Biological Sciences, **280** (1765), 20131149, **2013**.
63. PETERSEN M.J., ANDERSON V.J., JOHNSON R.L., EGGETT D.L. Seasonal Trap Abundance of Two Species of Psilochalcis Kieffer (Hymenoptera: Chalcididae) in Rangelands of the Eastern Great Basin of Utah, USA. Land, **12** (1), 54, **2023**.
64. ASHFAQ M., KHAN A.M., RASOOL A., AKHTAR S., NAZIR N., AHMED N., MANZOOR F., SONES J., PEREZ K., SARWAR G. A DNA barcode survey of insect biodiversity in Pakistan. PeerJ, **10** e13267, **2022**.
65. GEIGER M.F., MORINIERE J., HAUSMANN A., HASZPRUNAR G., WÄGELE W., HEBERT P.D., RULIK B. Testing the Global Malaise Trap Program – How well does the current barcode reference library identify flying insects in Germany? Biodiversity data journal, (4), **2016**.
66. PENTINSAARI M., BLAGOEV G.A., HOGG I.D., LEVESQUE-BEAUDIN V., PEREZ K., SOBEL C.N., VANDENBRINK B., BORISENKO A. A DNA barcoding survey of an Arctic arthropod community: implications for future monitoring. Insects, **11** (1), 46, **2020**.
67. STACKEBRANDT E., GOEBEL B.M. Taxonomic note: a place for DNA-DNA reassociation and 16S rRNA sequence analysis in the present species definition in bacteriology. International journal of systematic and evolutionary microbiology, **44** (4), 846, **1994**.
68. KAUSERUD H., MATHIESEN C., OHLSON M. High diversity of fungi associated with living parts of boreal forest bryophytes. Botany, **86** (11), 1326, **2008**.
69. RATNASINGHAM S., HEBERT P.D. A DNA-based registry for all animal species: the Barcode Index Number (BIN) system. PLoS ONE, **8** (7), e66213, **2013**.
70. LANDRY J.-F., HEBERT P.D. Plutella australiana (Lepidoptera, Plutellidae), an overlooked diamondback moth revealed by DNA barcodes. ZooKeys, (327), 43, **2013**.
71. BORKENT A. The state of phylogenetic analysis: narrow visions and simple answers – examples from the *Diptera* (flies). Zootaxa, **4374** (1), 107, **2018**.