Original Research

Optimizing Sunflower Growth, Nutrient Assimilation, and Biochemical Attributes under Salinity Stress Using a Combination of Sulfur-Treated Biochar and Arbuscular Mycorrhizal Fungi

Asif Minhas^{1#}, Muhammad Ikram^{2, 3}**, Asif Maqbool², Haseeb Ur Rehman², Ahsan Mehmood⁴, Hafiza Samra Younas¹, Abdullah Ehsan¹, Abdul Rauf⁵, Sami Al Obaid⁶, Mohammad Javed Ansari⁷, Ismail Khan⁸**

¹Department of Soil Science, Faculty of Agricultural Science's and Technology Bahauddin Zakariya University Multan, Pakistan
 ²Department of Agronomy, Faculty of Agricultural Science's and Technology Bahauddin Zakariya University Multan, Pakistan
 ³MOA Key Laboratory of Crop Ecophysiology and Farming System in the Middle Reaches of the Yangtze River,
 College of Plant Science & Technology, Huazhong Agricultural University, Wuhan, 430070, China
 ⁴Faculty of Agriculture and Life Sciences, University of Maribor, Slovenia
 ⁵National Key Laboratory of Horticultural Plant Biology, Ministry of Education, Wuhan, Hubei 430070, China
 ⁶Department of Botany and Microbiology, College of Science, King Saud University Riyadh, 11451, Saudi Arabia
 ⁷Department of Botany, Hindu College Moradabad

(Mahatma Jyotiba Phule Rohilkhand University Bareilly)-Uttar Pradesh, 244001, India ⁸Guangdong Provincial Key Laboratory of Applied Botany, and Key Laboratory of Vegetation Restoration and Management of Degraded Ecosystems, South China Botanical Garden, Chinese Academy

> Received: 23 March 2024 Accepted: 5 June 2024

Abstract

Soil salinity is a major issue that lowers the crop yield. Biochar is an organic amendment to the soil that mitigates the drastic effects of salinity. However, high pH is a problem in arid and semi-arid regions due to the use of biochar. Hence, the sulfur-treated biochar covers this problem to some extent. Furthermore, the use of arbuscular mycorrhizae also helps to cope with stressful environments. A limited study is done for the combined application of these strategies. To investigate this, the sulfur-treated biochar and AMF are evaluated for their individual and combined effects on the growth of sunflower plants under saline conditions. The study results revealed that the individual effect proved better, but the combination of BS and AMF showed a remarkable increase in the growth of sunflower plants in a saline environment. The integrated use of AMF and BS increases the plant agronomic attribute,

[#] equal contribution

^{*}e-mail: r4rana88@gmail.com, ikram1996@webmail.hzau.edu.cn;

^{**}e-mail: Ismailagronomist@gmail.com

NPK, and improves gas exchange parameters. The Electrolyte leakage was minimal where integrated application of AMF and BS was applied. The antioxidants that combat the ROS were found to be lower at combined application. This study opens a wide avenue for research exploration for the mitigation of salinity stress in the sunflower crop.

Keywords: sunflower, salinity, arbuscular mycorrhizae fungi, sulphur treated biochar, growth, antioxidants

Introduction

Soil salinity is a major issue facing the arid and semiarid regions, which reduces crop productivity [1]. It has been previously investigated that soil salinity lowers water availability and increases ion toxicity in plants, as well as deteriorates microbial activity in the soil [2]. The loss in production of crops and poor soil health to support plants is severely damaging to the farmer community as related to crop yield. The sunflower is an important edible oil crop. The crop is sensitive to salinity stress and yield reduction is considerable in salt-affected soils [3]. Soil salinity increases as the organic matter level decreases in the soil [4]. Soil microorganisms and organic matter are vital components for soil fertility reduction in microbial activities in saline soils and can decrease the growth of plants [5]. Enhancing the microbial and enzymatic activity of the soil can help plants resist the negative effects of soil salinity [6]. Biochar has traditionally been utilized to enhance soil conditions for improved crop growth. Biochar supplies carbon and nutrients to the microbial population, promoting soil enzymatic activity [7]. Several research efforts have investigated the impact of biochar on soil quality. These studies indicate that biochar enhances soil organic carbon, water-holding capacity, soil aeration, nutrient availability, stimulation of soil microbial and enzymatic activity, and cation exchange capacity (CEC) [8, 9]. Biochar has numerous advantages for soil, but its elevated pH level can pose challenges for soil fertility, particularly affecting the accessibility of soil minerals like phosphorus [10]. Raising the pH of soil enhances microbial nitrification, leading to nitrate losses and reduced availability of ammonium, the primary nitrogen source for plants [11].

Another method to reduce the impact of salinity is by utilizing arbuscular mycorrhizal fungi [12, 13]. Research on salt stress tolerance in mycorrhizal plants indicates that arbuscular mycorrhizal (AM) plants exhibit enhanced growth because of improved mineral nutrition and physiological activities, such as photosynthesis, water use efficiency, and osmoregulation. Arbuscular mycorrhizal fungi (AMF) are commonly found among various soil microorganisms in the rhizosphere [14]. The study [15] showed that inoculating arbuscular mycorrhizae improves nutrition absorption and preserves cell hydration. The AMF boosts the production of proteins and chlorophyll by directly affecting the absorption of magnesium, a crucial component of the chlorophyll molecule [16].

Salinity stress is a significant constraint affecting agricultural productivity, particularly in arid and semi-arid

regions. Sunflower (*Helianthus annuus L*.) is an important oilseed crop with considerable economic value [17]. The synergistic effects of Sulfur-treated biochar and AMF on sunflower growth, nutrient assimilation, and biochemical attributes under salinity stress remain understudied [18–20]. Based on preliminary observations and existing literature, we hypothesize that the combined application of Sulfurtreated biochar and arbuscular mycorrhizal fungi will lead to a significant improvement in sunflower growth, nutrient assimilation, and biochemical attributes under salinity stress conditions.

Material and Methods

Experimental Detail

A pot experiment was conducted in the research area of the Faculty of Agricultural Sciences and Technology, Bahauddin Zakariya University Multan (Punjab) Pakistan, which is situated at geographical coordinates 30°15′49″N &71°30′35″E (Fig. 1). The physiochemical properties of soil and irrigation water were given in (Table. 1). The experimental design followed a completely randomized design (CRD), where each treatment combination was random with three replications assigned to the experimental units (Control (CK), 1% Sulfur-treated biochar (1%BS), arbuscular mycorrhizal fungi (AMF) inoculation. Each treatment group was subjected to two different salinity levels i.e., control having EC 2.58 dSm¹ and saline soils having EC 5.68 dSm¹.

Seeds Collection and Sterilization

Hysun33 (*Helianthus annuus L.*) seeds were obtained from Government of Punjab Pakistan-certified seed dealers. To ensure quality, only sound seeds were chosen for the experiment, excluding broken or fragile ones. Select seeds were surface-sterilized before sowing. Starting with 5% sodium hypochlorite, the seeds were rinsed three times with 95% ethanol. The seeds were triple-washed in sterile deionized water to remove leftover sterilizing chemicals [21].

Fertilizer

A balanced 60:40:25 ratio of nitrogen (N), phosphorus (P), and potassium (K) met plant nutritional needs. This essential nutrition blend came from artificial fertilizers. These nutrients were delivered during sowing, watering, and flowering. These growth phases had different nutritional

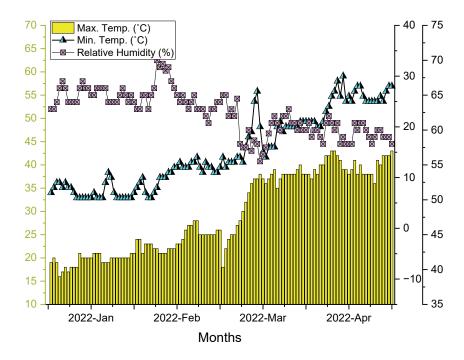


Fig. 1. Climatic data of experimental site.

amounts. Along with primary nutrients, the cultivation regimen included additional micronutrients like Sulphur (S), zinc (Zn) at 33%, and borax (B) at 11%. The micronutrient application was exact, with 5 kg per acre for Sulfur, 6 kg for zinc, and 3 kg for borax.

Harvesting and Data Collection

125 days after seeding, the plants were mature and ready for harvesting. Each plant's height, achene count, and head diameter were carefully measured. With a calibrated electric balance, 1000 achenes were weighed. We harvested the entire pot and let the plants dry in the sun after this inspection. The plant heads were carefully removed, and the grains were manually threshed. Using a calibrated electric balance, all grains from each pot were weighed.

Determination of Macronutrients

For the determination of nitrogen, phosphorus, and potassium in plant samples, the procedure mentioned in the ICARDA manual 3rd edition was followed by Ryan et al. (2001).

Chlorophyll (Spad) Contents

Chlorophyll content was measured using a portable chlorophyll meter (SPAD meter) by placing the meter on a fully expanded leaf and recording the SPAD reading.

Relative Water Content (%)

Fresh weights of leaf samples were recorded. After soaking in distilled water for several hours, they were wiped dry and reweighed for turgidity. Finally, the samples were oven-dried and weighed again to establish dry weight. The formula for relative water content (RWC) was:

RWC (%) = [(Fresh Weight – Dry Weight) / (Turgid Weight – Dry Weight)] × 100

Electrolyte Leakage (%)

Electrolyte leakage was determined as an indicator of cell membrane damage and stress. Leaf samples were collected and submerged in deionized water (initial conductivity, C1). After 24 hours, the conductivity of the solution (final conductivity, C2) was measured using a conductivity meter. Electrolyte leakage was calculated as the percentage of conductivity increase:

Electrolyte Leakage (%) = $((C2-C1))/C1 \times 100$

Gas Exchange Attributes

Quantifying gas exchange properties required the Infra-Red Gas Analyzer. This sophisticated device measured photosynthesis, intercellular CO2 concentration, transpiration, and stomatal conductance.

Data Analysis

To identify significant treatment differences, the data were analyzed using ANOVA. After comparing means with appropriate Tukey's post-hoc tests, correlation analyses were used to examine parameter connections. To establish treatment effects and significance, R was used for statistical analysis.

Soil	Values	Biochar	Values	Irrigation	Values
pН	8.36	рН	pH 5.11 pH		7.07
ECe (dSm ⁻¹)	2.58	ECe (dSm ⁻¹)	4.95	EC (μS/cm)	195
SOM (%)	0.60	Ash Content (%)	t (%) 35 Carbonates (meq./L)		0.00
TN (%)	0.03	Volatile Matter (%)	r (%) 10 Bicarbonates (meq./L)		5.62
EP (mg/kg)	6.97	Fixed carbon (%)	Fixed carbon (%) 55 Chloride (meq./L)		0.00
AK (mg/kg)	143	TN (%)	(%) 0.45 $Ca + Mg (meq./L)$		4.39
Sand (%)	25	TP (%)	0.85	Sodium (mg/L)	100
Silt (%)	40	TK (%)	0.49	TN = Total Nitrogen	
Clay (%)	35	Surface area (m ² /g)	400	EP = Extractable AK = Available	
Texture	Clay Loam	CEC (meq./100 g)	465	CEC = Cation Exchange Capacity EC = Electrical Conductivity	

Table 1. Pre-experimental biochar and irrigation characteristics.

Results

Plant Height

The study measured sunflower plant height at 2.58 dS/m and 5.68 dS/m to quantify salinity stress. Compared to the control group's 51.62 cm average height, 1% Sulfur-Treated Biochar (BS) and 0.5% Arbuscular Mycorrhizal Fungi (AMF) raised plant height by 11.74% and 18.76%, respectively. The combination of 0.5% AMF and 1% BS increased 31.07%. In the 5.68 dS/m salinity stress condition, controls averaged 49.50 cm tall. Adding 1% Sulfur-Treated Biochar and 0.5% Arbuscular Mycorrhizal Fungi improved plant height by 12.25% and 20.54%. 0.5% AMF and 1% BS increased sunflower crop growth by 30.99%, suggesting they may reduce salt stress (Table 2).

Root Fresh Weight

The sunflower crop's root fresh weight (g/plant) under 2.58 dS/m and 5.68 dS/m salt stress yielded important insights. The control group (CK) averaged 1.99 grams per plant in root fresh weight at 2.58 dS/m. Compared to the control group, 1% Sulfur-Treated Biochar (1%BS) increased yields by 20.50% to 2.40 g/plant. Compared to the control group, 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) increased the average root fresh weight by 66.02% to 3.31 g/plant. The treatment with 0.5% arbuscular mycorrhizal fungus (AMF) and 1% beneficial soil bacteria (BS) produced the highest root fresh weight, 3.84 g per plant. This is 92.59% higher than the control group. The control group had an average root fresh weight of 1.49 g/plant after 5.68 dS/m saline exposure. Compared to the control group, 1% Sulfur-Treated Biochar increased root fresh weight by 41.31% to 2.11 g/plant. Similar to the control group, 0.5% Arbuscular Mycorrhizal Fungi increased root fresh weight by 93.19% to 2.89 g/plant. Significantly, 0.5% arbuscular mycorrhizal fungus (AMF) and 1% bio-stimulant (BS) produced an average root fresh weight of 3.44 g per plant. This is 130.11% higher than the control group (Table 2).

Shoot Fresh Weight

Fresh sunflower shoot weight (g/plant) was measured under 2.58 dS/m and 5.68 dS/m salt stress. The control group (CK) averaged 6.67 g/plant shoot fresh weight at 2.58 dS/m salinity. 1% Sulfur-Treated Biochar (1% BS) boosted growth by 49.32% over the control at 9.96 g/plant. 0.5% AMF raised the average shoot fresh weight by 96.86% to 13.13 g/plant compared with the control. The 0.5% AMF and 1% BS treatment generated the highest average shoot fresh weight at 16.47 g/plant, 146.91% greater than the control group. The control group averaged 4.96 g/plant shoot fresh weight at 5.68 dS/m salinity. Over the control, 1% Sulfur-Treated Biochar raised the shoot fresh weight 67.34% to 8.30 g/plant. The average shoot fresh weight with 0.5% Arbuscular Mycorrhizal Fungi was 11.83 g/plant, 138.61% higher than the control. 0.5% AMF and 1% BS raised shoot fresh weight 206.27% to 15.18 g/plant (Table 2).

Root Dry Weight

In the control group (CK), root dry weight averaged 0.87 g/plant at 2.58 dS/m. With an average of 1.13 g/plant, 1% Sulfur-Treated Biochar (1% BS) increased growth by 29.81% over the control. The treatment of 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) increased root dry weight by 60.67% to 1.39 g/plant. The treatment of 0.5% AMF plus 1% BS (0.5% AMF + 1% BS) produced the greatest average root dry weight at 1.75 g/plant, 102.14% more than the control group. At 5.68 dS/m salinity, the control group had 0.71 g/plant root dry weight. 1% Sulfur-Treated Biochar increased root dry weight by 45.96% to 1.03 g/plant. Arbuscular Mycorrhizal Fungi at 0.5% produced an average root dry weight of 1.25 g/plant, a 77.71% increase over the control. The treatment of 0.5% AMF and 1% BS

C	1		,		,	
Treatments	2.58 dS/m	5.68 dS/m	2.58 dS/m	5.68 dS/m	2.58 dS/m	5.68 dS/m
	Plant height (cm)		Root fresh weight (g plant -1)		Shoot fresh weight (g plant -1)	
CK	51.62f±0.66	49.5f±2.06	1.99e±0.02	1.5f±0.15	6.67ef±0.68	4.96f±0.72
BS	57.68de±0.9	55.56e±0.78	2.4d±0.12	2.11de±0.12	9.96cd±0.8	8.3de±0.77
AMF	61.3bc±0.32	59.67cd±0.99	3.31b±0.05	2.89c±0.08	13.13b±1.18	11.83bc±0.27
AMF + BS	67.66a±1.5	64.84ab±1.81	3.84a±0.12	3.44b±0.12	16.47a±0.45	15.18a±0.54

Table 2. Effect of Sulfur-treated biochar (BS) and arbuscular mycorrhizae (AMF) on plant height, root fresh weight, and shoot fresh weight of sunflower plants cultivated under normal (EC = 2.58 dS/m) and saline (EC = 5.68 dS/m) soil condition.

CK = Control, BS = Sulfur-treated biochar, AMF = Arbuscular mycorrhizae, different letters showing the Tukey's HSD results significant at the $p \le 0.05$ along with mean value (n = 3) and \pm standard deviation.

increased root dry weight by 122.04% over the control group to 1.57 g/plant (Table 3).

Shoot Dry Weight

The shoot dry weight (g/plant) of sunflower cultivars under 2.58 dS/m and 5.68 dS/m salinity stress conditions was intriguing. The control group (CK) averaged 1.63 g/plant shoot dry weight at 2.58 dS/m salinity. An average of 2.39 g/plant, 46.25 percent higher than the control, was achieved with 1% Sulfur-Treated Biochar (1% BS). Compared to the control, 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) increased the average shoot dry weight by 113.70% to 3.49 g/plant. 0.5% AMF plus 1% BS (0.5% AMF + 1% BS) produced the highest average shoot dry weight of 3.89 g/plant, 137.80% more than the control group. At 5.68 dS/m salinity, the control group averaged 1.43 g/plant shoot dry weight. Compared to the control, 1% Sulfur-Treated Biochar raised the plant shoot dry weight by 35.79%. The application of 0.5% Arbuscular Mycorrhizal Fungi produced an average shoot dry weight of 2.88 g/plant, a 101.25 percent increase over the control. The combination of 0.5% AMF and 1% BS produced an average shoot dry weight of 3.71 g/plant, 159.25 % more than the control group (Table 3).

Head Diameter

Head diameter averaged 5.32 cm for the control group (CK) at 2.58 dS/m salinity. With an average head diameter

of 5.84 cm, 1% Sulfur-Treated Biochar (1%BS) increased growth by 9.77% over the control. Compared to the control, 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) increased head diameter by 20.87% to 6.43 cm. The treatment of 0.5% AMF and 1% BS (0.5% AMF + 1% BS) produced the largest head diameter at 7.30 cm, 37.17% higher than the control group. An average head diameter of 5.10 cm was observed in the control group at 5.68 dS/m salinity. The average head diameter increased by 7.99% with 1% Sulfur-Treated Biochar to 5.51 cm. Applying 0.5% Arbuscular Mycorrhizal Fungi increased head diameter by 6.20 cm, a 21.50% increase over the control. The average head diameter was 7.02 cm after 0.5% AMF and 1% BS treatment, a 37.62% increase over the control group (Table 3).

Number of Achene Head-1

The control group (CK) averaged 244.50 achenes per head at 2.58 dS/m salinity. Sulfur-treated biochar (1% BS) increased achenes per head by 19.34%, averaging 291.79 per head. Similar to 0.5% AMF, 333.59 achenes per head were produced, a 36.44% increase over the control. The combination treatment of 0.5% AMF and 1% BS (0.5% AMF + 1% BS) caused the greatest achenes per head (372.62), 52.40% more than the control group. After switching to 5.68 dS/m salinity, the control group averaged 219.29 achenes per head. Over the control, 1% Sulfur-Treated Biochar produced 268.81 achenes per head, 22.58% more. Similarly, 0.5% Arbuscular Mycorrhizal Fungi produced 312.14 achenes per head, 42.34%

Table 3. Effects of Sulfur-treated biochar (BS) and arbuscular mycorrhizae (AMF) on root dry weight, shoot dry weight and head diameter of sunflower plants cultivated under normal (EC = 2.58 dS/m) and saline (EC = 5.68 dS/m) soil condition.

	Root dry weight (g plant ⁻¹)		Shoot dry weight (g plant ⁻¹)		Head diameter (cm)	
CK	$0.87 \text{fg} \pm 0.05$	0.7g±0.1	1.64de±0.12	$1.43e\pm0.07$	5.32ef±0.08	5.1f±0.13
BS	1.13de±0.04	1.03ef±0.02	2.39c±0.02	1.94cd±0.2	5.84cd±0.16	5.51de±0.08
AMF	1.4bc±0.04	1.26cd±0.07	3.49a±0.17	2.88b±0.35	6.43b±0.07	6.2bc±0.12
AMF + BS	1.75a±0.11	1.57ab±0.06	3.89a±0.08	3.71a±0.05	7.29a±0.23	7.02a±0.15

CK = Control, BS = Sulfur-treated biochar, AMF = Arbuscular mycorrhizae, different letters showing the Tukey's HSD results significant at the $p \le 0.05$ along with mean value (n = 3) and \pm standard deviation.

weight, and deficitely feld of Salme weight plants cally raised and refinal (De 2500 do m) and Salme (De 500 do m) son condition.								
	Number of a	chene head-1	Thousand achene weight (g)		Achene yield (g plant ⁻¹)			
CK	244.49f±7.21	219.3g±6.47	32.43de±0.16	30.16e±1.6	6.92ef±0.03	6.35f±0.26		
BS	291.79d±12.09	268.81e±10.53	37.81c±0.28	33.5d±0.13	8.46d±0.18	7.49e±0.41		

39.95c±0.68

 $51.98a\pm1.15$

38.66c±0.4

45.02b±2.55

Table 4. Effect of Sulfur-treated biochar (BS) and arbuscular mycorrhizae (AMF) on a number of achene head-1, thousand achene weight, and achene yield of sunflower plants cultivated under normal (EC = 2.58 dS/m) and saline (EC = 5.68 dS/m) soil condition.

CK = Control, BS = Sulfur-treated biochar, AMF = Arbuscular mycorrhizae, different letters showing the Tukey's HSD results significant at the $p \le 0.05$ along with mean value (n = 3) and \pm standard deviation.

more than the control. The combined therapy of 0.5% AMF and 1% BS produced 343.91 achenes per head, 56.83% more than the control group (Table 4).

312.14cd±4.9

343.91b±0.44

333.59bc±5.21

372.62a±9.18

AMF

AMF + BS

Thousand Achene Weight

Under 2.58 dS/m and 5.68 dS/m salinity stress conditions, sunflower crop thousand achene weight (g) was significant. The control group (CK) averaged 32.44 g per thousand achenes at 2.58 dS/m salinity. Adding 1% Sulfur-Treated Biochar (1% BS) increased growth by 16.57%, averaging 37.81 g. Compared to the control, 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) increased the average thousand achene weight by 23.18% to 39.95 g. Most impressively, 0.5% AMF plus 1% BS (0.5% AMF + 1% BS) produced the greatest thousand achene weight at 51.98 g, 60.24% higher than the control group. At 5.68 dS/m salinity, the control group averaged 30.16 g per thousand achenes. With 1% Sulfur-Treated Biochar, the average thousand achene weight was 33.50 g, up 11.05% from the control. The average thousand achene weight was 38.66 g with 0.5% Arbuscular Mycorrhizal Fungi, a 28.16% increase over the control. Achieving an average thousand achene weight of 45.02 g with 0.5% AMF and 1% BS was 49.25% higher than the control group (Table 4).

Achene Yield

Achene yield (g/plant) in sunflower cultivars under 2.58 dS/m and 5.68 dS/m salinity stress conditions provided significant insights. The control group (CK) produced 6.92 g of achenes per plant at 2.58 dS/m salinity. With an average of 8.46 g per plant, 1% Sulphur-Treated Biochar (1% BS) increased achene yield by 22.35%. The application of 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) increased achene yield by 48.71% to 10.29 g/plant. The combo treatment of 0.5% AMF and 1% BS yielded the most achenes at 11.31 g/plant, 63.55% more than the control group. At 5.68 dS/m salinity, the control group yielded 6.35 g per plant in achenes. Adding 1% Sulfur-Treated Biochar increased achene output by 17.97% to 7.49 g/plant. Using 0.5% Arbuscular Mycorrhizal Fungi increased the achene output by 51.45% to 9.62 g/plant. Compared to the control group, 0.5% AMF and 1% BS boosted plant achene yield by 69.58 percent (Table 4).

Root/Shoot (N:P:K) Assimilation

 $10.29bc\pm0.13$

11.31a±0.3

 $9.62c\pm0.19$

10.77ab±0.29

The impact of various treatments on nitrogen, phosphorus, and potassium content in sunflower shoots and roots under different salinity stress levels is evident. Generally, treatments such as Sulfur-Treated Biochar (1% BS) and Arbuscular Mycorrhizal Fungi (AMF) individually and in combination significantly enhanced nutrient uptake compared to the control groups across both salinity levels. For nitrogen concentration in shoots and roots, the application of 0.5% AMF and 1% BS (0.5% AMF + 1% BS) produced the highest increases, with a 28.90% rise in shoot nitrogen and a remarkable 47.31% increase in root nitrogen compared to respective controls. Regarding phosphorus, the combined treatment of 0.5% AMF and 1% BS consistently resulted in the highest increments, with shoot phosphorus increasing by 54.68% and root phosphorus by 60.13% compared to controls across salinity levels. Similarly, for potassium content, the combined treatment of 0.5% AMF and 1% BS showed the most significant enhancements, with shoot potassium increasing by 34.71% and root potassium by 63.37% compared with controls under 2.58 dS/m salinity. Overall, these findings suggest that combined applications of AMF and BS can effectively mitigate salinity stress and promote nutrient uptake in sunflower crops (Fig. 2, Fig. 3A, B).

Chlorophyll Contents (SPAD)

The control group averaged 23.90 Chlorophyll at 2.58 dS/m. Sulfur-treated Biochar (1% BS) boosted Chlorophyll content by 15.46%, whereas 0.5% Arbuscular Mycorrhizal Fungi (0.5%) enhanced it by 28.82%. 0.5% AMF and 1% BS produced the greatest average Chlorophyll content at 34.40, a 43.92% increase. At 5.68 dS/m, the control group had 22.37 chlorophyll, whereas 1% Sulfur-Treated Biochar increased it by 11.18%. Chlorophyll concentration averaged 28.88 with 0.5% Arbuscular Mycorrhizal Fungi, a 29.14% increase. The combination of 0.5% AMF and 1% BS increased by 46.51% (Fig. 3C).

Relative Water Contents

The control group's (CK) relative water content was 68.68% at 2.58 dS/m salinity. 1% Sulfur-Treated Biochar

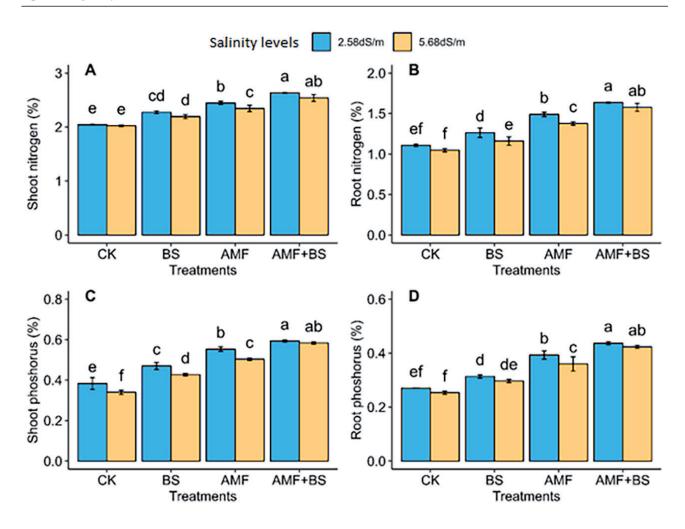


Fig. 2. Effect of Sulfur-treated biochar (BS), arbuscular mycorrhizae (AMF) and their combination on the shoot and root, nitrogen and phosphorus percentage in sunflower plants. Different letters on bars (means of 3 replicates \pm SD) showing significant changes at the $p \le 0.05$ compared by using Tukey's HSD.

(1% BS) boosted growth by 5.12% over the control with an average relative water content of 72.19%. 0.5% Arbuscular Mycorrhizal Fungi (AMF) raised relative water content by 9.26% to 75.04% compared to the control. The treatment of 0.5% AMF and 1% BS (0.5% AMF + 1% BS) yielded the highest average relative water content at 79.20%, 15.32% higher than the control group. The control group's relative water content was 67.47% at 5.68 dS/m salinity. Compared to the control, 1% Sulfur-Treated Biochar raised the relative water content to 70.02%. With 0.5% Arbuscular Mycorrhizal Fungi, the relative water content averaged 73.64%, up 9.14% from the control. To 77.14%, 0.5% AMF and 1% BS increased relative water content by 14.32% above the control group (Fig. 3D).

Electrolyte Leakage

Under CK had an average EL of 65.27% during 2.58 dS/m salt stress. The addition of 1% Sulfur-Treated Biochar (1% BS) reduced EL by 17.3%. EL dropped 26.5% with 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF). The therapy of 0.5% AMF plus 1% BS (0.5% AMF + 1% BS) demonstrated

the greatest reduction, 38.7%. At a salinity of 5.68 dS/m, the control group had an average EL of 67.11%. According to the previous trend, 1% Sulfur-Treated Biochar reduced EL by 9.6%. Applying 0.5% Arbuscular Mycorrhizal Fungi reduced EL by 25.5%. The highest significant reduction was 34.9% with 0.5% AMF and 1% BS (Fig. 4A).

Net Photosynthetic Rate

At 2.58 dS/m salinity, the control group (CK) had an average net photosynthetic rate of 15.67 μmolm-2s-1. The addition of 1% Sulfur-Treated Biochar (1% BS) led to an average growth rate of 17.50 μmolm-2s-1, an 11.67% increase over the control. The addition of 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) resulted in an average rate of 20.51 μmolm-2s-1, a significant increase of 30.84% compared to the control. The treatment with 0.5% AMF and 1% BS resulted in the highest Net Photosynthetic Rate of 23.63 μmolm-2s-1, a 50.79% increase over the control group. After adjusting to 5.68 dS/m salinity, the control group achieved an average Net Photosynthetic Rate of 15.26 μmolm-2s-1. Adding 1% Sulfur-Treated Biochar

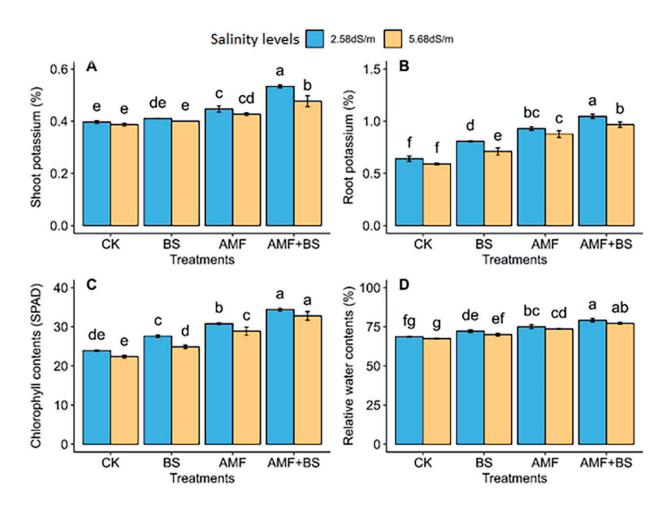


Fig. 3. Effect of Sulfur-treated biochar (BS), arbuscular mycorrhizae (AMF), and their combination on the shoot and root potassium percentage, chlorophyll contents, and relative water contents in sunflower plants. Different letters on bars (means of 3 replicates \pm SD) showing significant changes at the $p \le 0.05$ compared by using Tukey's HSD.

resulted in an average rate of 17.30 μ molm-2s-1, a 13.39% increase over the control. Applying 0.5% Arbuscular Mycorrhizal Fungi resulted in an average growth rate of 19.26 μ molm-2s-1, a significant 26.21% increase over the control. In the treatment of 0.5% AMF and 1% BS, the average Net Photosynthetic Rate increased by 45.94% over the control group (Fig. 4B).

Transpiration Rate

Transpiration Rate averaged 2.86 mmolm-2s-1 at 2.58 dS/m for the control group (CK). With 1% Sulfur-Treated Biochar (1% BS), growth averaged 3.86 mmolm-2s-1, 35.22% higher than the control. The treatment of 0.5% Arbuscular Mycorrhizal Fungi increased the average rate by 69.26% to 4.84 mmolm-2s-1. 0.5% AMF and 1% BS (0.55% AMF + 1% BS) had the highest average Transpiration Rate at 5.69 mmolm-2s-1, 99.11% higher than the control group. Transpiration Rate averaged 2.24 mmolm-2s-1 in the control group under 5.68 dS/m. Averaging 4.36 mmolm-2s-1, 1% Sulfur-Treated Biochar and 0.5%

Arbuscular Mycorrhizal Fungi increased growth by 94.47% over the control. Treatment with 0.5% AMF and 1% BS increased by 143.63% over the control group (Fig. 4C).

Enzymatic Activities (SOD, POD, CAT)

The enzymatic activities of Superoxide Dismutase (SOD), Peroxidase (POD), and Catalase (CAT) in sunflower crops exhibited significant alterations under varying salinity stress levels. For SOD activity, at 2.58 dS/m salinity, the introduction of treatments led to reductions compared to the control, with the combined treatment of 0.5% AMF and 1% BS resulting in the most substantial decrease, around 33.7%. Similarly, at 5.68 dS/m, treatments also reduced SOD activity, with the combined treatment showing the most significant reduction, approximately 33.2%. Regarding POD activity, at both salinity levels, treatments caused decreases compared to the control, with the combined treatment of 0.5% AMF and 1% BS demonstrating the most prominent reduction, approximately 38.3% at 2.58 dS/m and 28.4% at 5.68 dS/m (Fig. 4D, 5A, B).

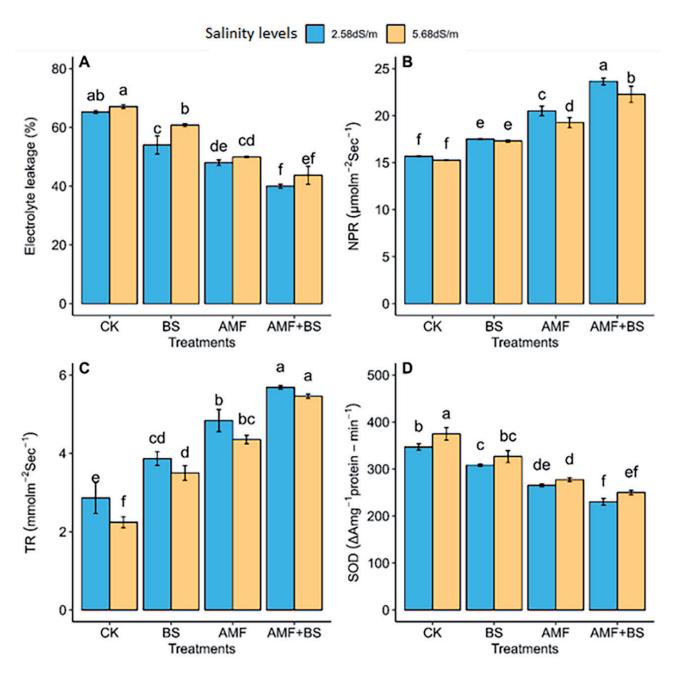


Fig. 4. Effect of Sulfur-treated biochar (BS), arbuscular mycorrhizae (AMF), and their combination on the shoot and root potassium percentage, chlorophyll contents, and relative water contents in sunflower plants. Different letters on bars (means of 3 replicates \pm SD) showing significant changes at the $p \le 0.05$ compared by using Tukey's HSD.

For CAT activity, reductions were observed under both salinity conditions with treatment application. The combined treatment consistently resulted in the greatest reduction in CAT activity compared to individual treatments and the control group. Overall, these findings indicate that the combined application of 0.5% AMF and 1% BS consistently led to the most significant reductions in enzymatic activities across all three enzymes under study, suggesting potential implications for mitigating salinity stress in sunflower crops.

Proline Contents

The Proline levels in sunflower (Helianthus annuus) crops exposed to 2.58 dS/m and 5.68 dS/m salt stress showed different percentage changes. Under 2.58 dS/m salinity stress, the control group (CK) had an average Proline level of 14.04 µmol/g. Adding 1% Sulfur-Treated Biochar (1% BS) reduced Proline by 15.1%. Similarly, 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) reduced Proline levels by 32.5%. The therapy with 0.5% AMF

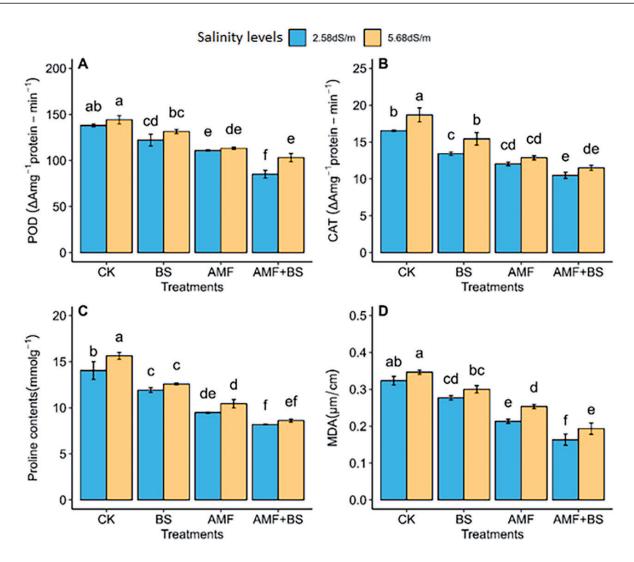


Fig. 5. Effect of Sulfur-treated biochar (BS), arbuscular mycorrhizae (AMF), and their combination on the shoot and root potassium percentage, chlorophyll contents, and relative water contents in sunflower plants. Different letters on bars (means of 3 replicates \pm SD) showing significant changes at the $p \le 0.05$ compared by using Tukey's HSD.

and 1% BS (0.5% AMF + 1% BS) showed the greatest reduction, 41.7%. At a salinity level of 5.68 dS/m, the control group had an average Proline content of 15.63 μmol/g. As expected, 1% Sulfur-Treated Biochar reduced Proline levels by 19.5%. Similarly, 0.5% Arbuscular Mycorrhizal Fungi reduced Proline by 33.2%. The largest significant decrease was 44.8% with 0.5% AMF and 1% BS (Fig. 5C).

Malondialdehyde (MDA)

At salinity stress of 2.58 dS/m, the control group (CK) had an average MDA level of 0.33μm cm-1. The addition of 1% Sulfur-Treated Biochar (1% BS) reduced MDA levels by 15.2%. Applying 0.5% Arbuscular Mycorrhizal Fungi (0.5% AMF) resulted in a significant drop in MDA levels, around 36.4%. The therapy of 0.5% AMF plus 1% BS (0.5% AMF +1% BS) showed the greatest reduction, 48.5%. Upon adjusting to 5.68 dS/m salinity, the control group had an average MDA level of 0.34 μm cm-1. In line with the previous trend, 1% Sulfur-Treated Biochar reduced MDA

levels by 11.8%. Similarly, 0.5% Arbuscular Mycorrhizal Fungi reduced MDA by 11.8%. The largest significant decrease was 44.1% with 0.5% AMF and 1% BS (Fig. 5D).

Discussion

Salinity reduces sunflower growth and biochemistry, according to the study. As salt-sensitive plants, sunflowers grew and yielded less at higher salinities. Salinity stress did not stop the AMF and BS from surviving. AMF and BS help plants absorb NPK, as shown by their increased NPK content. The treatments significantly improved plant growth (Table 2). Arbuscular mycorrhizae and sulfur treatment biochar increase plant height, root and shoot dry and fresh weight, and achene yield. The study showed improved nutrient uptake, which may explain this. The arbuscular mycorrhizae with Sulfur-treated biochar strategy outperformed the individual treatments. Similar results were seen with saline-irrigated sunflowers [22]. [23] found that AMF and PGBP increased fungi

colonization and nutrient uptake in salinity-treated lucerne. The current experiment showed that AMF increased NPK concentration more than BS [24, 25]. The best results came from AMF and BS. BS may control nutrient toxicity, and AMF increases plant NPK, which boosts yield and agronomic qualities [26]. AMF increases potassium uptake, which maintains turgor pressure and osmotic balance, preventing enzymatic disruption and protein synthesis inhibition [27–29].

Arbuscular mycorrhizae and BS improved chlorophyll, relative water, and photosynthetic rate, but the combination of AMF and BS yielded remarkable results. Electrolyte leakage (EL) indicates plant injury in abiotic stress. AMF-BS combination had the lowest EL under both salinity stress levels. The study confirmed that AMF is a biological method for mitigating salinity effects and improving plant morphological, physiological, and biochemical changes that cope with salinity [30–33]. This boosts nutrient absorption. Since magnesium is an essential part of chlorophyll, AMF boosts de novo protein and chlorophyll synthesis [34]. [35] found that AMF enhanced chlorophyll pigment, supporting the study results.

Plants under high salinity stress had lower NPK levels. These findings were consistent with [36, 37], who found that soil-increasing salinity decreased plant N concentration. Microbial treatments raised root N levels. AMF may have increased N by assimilation of nitrate in extraradical mycelium and increased enzyme production for primary nitrogen fixation [38, 39]. Plants under saline stress absorb less P [40]. AMF increases P concentration in salt-affected soil plants [41]. AMF and BS reduced stress-induced increases in antioxidants like SOD, POD, CAT, and proline. Our study supported Ebrahimian et al. [42], who found that salinity stress greatly increased enzyme activity. Similar results were obtained by [43] on stressed millet leaves.

Salt stress alters plant metabolism. Ion toxicity, osmotic stress, and ROS production are the main ones [44]. Antioxidant enzymes like SOD, APX, and CAT reduce ROS production [45]. In sunflowers, cadmiumstressed plants accumulate more proline, which increases tolerance [46]. Higher proline levels maintain cell osmotic balance and water content. Instead of interfering with metabolic pathways, proline replaces water [47–50]. AMFinoculated plants accumulate more proline under normal and stressed conditions [51, 52]. Our study found lower proline concentrations, contradicting [49]. The BS may increase NPK and preserve soil conditions for plant growth by reducing stress, while the AMF may increase NPK uptake and reduce proline production. Previous research showed that sulfur-modified biochar with sulfate ions increased CEC. The number of functional groups in sulfurmodified biochar changes, causing this CEC change. Musa al-Reza Taheri found that B and BS treatments increased RWC, with BS enhancing RWC at a lower level than B. The relationship between RWC and soil water content is positive [53]. Sulfur-enriched biochar and effective microorganisms improved Capsicum annuum growth and yield under salt stress [52, 53]. Ion toxicity, osmotic stress, and ROS production are the main effects of salt stress on plant metabolism [53, 54]. SOD, APX, and CAT reduce ROS production [53]. The current study also found low antioxidant production in the integrated effect of AMF and BS, suggesting favorable plant growth conditions. Salinity stress increased antioxidant enzyme activity, as found in [54]. Similar results were obtained by [55] on stressed millet leaves.

The latest study [56] found that 0.50AMF-BC reduces antioxidants and drought stress at the highest dose. They also release glomalin, a glycoprotein that helps soil aggregation and water retention. In times of osmotic stress, it helps the soil retain moisture for longer, providing plant roots with water. The increased water retention capacity helps maintain soil moisture and provides plants with more consistent water, reducing osmotic stress [57]. A porous carbon-based substance called biochar improves soil structure and water retention, improving treatment effectiveness. Due to its large surface area and micropores, the material stores water and nutrients, maintaining soil moisture and facilitating root penetration [58, 59]. Biochar prevents soil compaction and improves water infiltration [60]. The study examined biochar and [61, 62] synergy. This study examines how AMF-Biochar affects spinach plant growth, nutrient concentrations, and antioxidant enzymes to reduce drought stress. Sulfur-treated biochar and arbuscular mycorrhizal fungi (AMF) reduced salinity-induced stress in sunflower cultivation, improving plant height, shoot biomass, and root development [63]. This treatment regimen also improved nutrient assimilation, as plant tissues had higher nitrogen, phosphorus, and potassium levels. The combined application also increased chlorophyll content, antioxidant enzyme activities (e.g., superoxide dismutase, catalase), and osmolytes like proline, reducing salinity stress-induced oxidative damage. Biochar and AMF symbiosis improve soil structure and nutrient retention, which may explain the synergistic effects. Future research could investigate the complex molecular mechanisms behind these synergies and conduct field trials to evaluate this approach's longterm efficacy and scalability in various agroecosystems. These findings demonstrate that integrating sulfur-treated biochar and AMF can sustainably improve sunflower salinity stress resilience and crop productivity.

Conclusion

The results of the current pilot research study conclude that treatments with arbuscular mycorrhizae fungi and Sulfur-treated biochar proved beneficial for the growth of the sunflower plant. However, the combination of 0.5% AMF and 1% Sulfur-treated biochar gives remarkable results to cope with the salinity problem. Furthermore, field experimentation is preferred to verify the results and the investigation of the mechanisms involves combating the salinity problem in agriculture.

Author Contributions

A.M,& M.I. Contributed equally; Conceptualization: A.M, I.K, & M.I.; Methodology: A.M., M.I., I.K., M.J.A., S.A.O & A.M; Data Curation: A.M., M.I, & H.S.U.; Writing-original draft preparation: A.M., M.I. & A.E.; Writing-Review and Editing: M.A., & M.I., AR.

Acknowledgments

This project was supported by Researchers Supporting Project number (RSP2025R315) King Saud University, Riyadh, Saudi Arabia.

Conflicts of Interest

The author declares no conflicts of interest regarding the publication of this paper.

References

- HASSANI A., AZAPAGIC A., SHOKRI N. Global predictions of primary soil salinization under changing climate in the 21st century. Nature Communications, 12 (1), 6663, 2021.
- SAFDAR H., AMIN A., SHAFIQ Y., ALI A., YASIN R., SHOUKAT A., SARWAR M. I. A review: Impact of salinity on plant growth. Nature and Science, 17 (40), 2019.
- WICHERN F., ISLAM M.R., HEMKEMEYER M., WATSON C., JOERGENSEN R.G. Organic amendments alleviate salinity effects on soil microorganisms and mineralisation processes in aerobic and anaerobic paddy rice soils. Frontiers in Sustainable Food Systems, 4 (30), 2020.
- SRITONGON N., SARIN P., THEERAKULPISUT P., RIDDECH N. The effect of salinity on soil chemical characteristics, enzyme activity and bacterial community composition in rice rhizospheres in Northeastern Thailand. Scientific Reports, 12 (1), 20360, 2022.
- PALANSOORIYA K.N., WONG J.T.F., HASHIMOTO Y., HUANG L., RINKLEBE J., CHANG S.X., OK Y.S. Response of microbial communities to biochar-amended soils: a critical review. Biochar, 3 (22), 2019.
- KHAN I., CE L., WU Q., XUANMING W., BINHANG Y., ABDUL R., ASIF A.K., JAVID K., WANG L.-X. The residual impact of straw mulch and biochar amendments on grain quality and amino acid contents of rainfed maize crop. Journal of Plant Nutrition, 46 (7), 1283, 2023.
- KHAN I., TARIQ M., ALABBOSH K.F., REHMAN A., JALAL A., KHAN A.A., FAROOQ M., LI G., IQBAL B., AHMAD N., KHAN K.A. Soil microplastics: Impacts on greenhouse gasses emissions, carbon cycling, microbial diversity, and soil characteristics. Applied Soil Ecology, 105343, 2024.
- 8. TAHERI M.A.R., ASTARAEI A.R., LAKZIAN A., EMAMI H. Sorbitol and biochar have key roles in microbial and enzymatic activity of saline-sodic and calcareous soil in millet cropping. Rhizosphere, 24 (1), 2022.
- 9. WICHERN F., ISLAM M.R., HEMKEMEYER M., WATSON C., JOERGENSEN R.G. Organic amendments

- alleviate salinity effects on soil microorganisms and mineralisation processes in aerobic and anaerobic paddy rice soils. Frontiers in Sustainable Food Systems 4 (30), 2020.
- SRITONGON N., SARIN P., THEERAKULPISUT P., RIDDECH N. The effect of salinity on soil chemical characteristics, enzyme activity and bacterial community composition in rice rhizospheres in Northeastern Thailand. Scientific Reports, 12 (1), 20360, 2020.
- 11. PALANSOORIYA K.N., WONG J.T.F., HASHIMOTO Y., HUANG L., RINKLEBE J., CHANG S.X., OK Y.S. Response of microbial communities to biochar-amended soils: a critical review. Biochar, 3 (22), 2019.
- 12. TAHERI M.A.R., ASTARAEI A.R., LAKZIAN A., EMAMI H. Sorbitol and biochar have key roles in microbial and enzymatic activity of saline-sodic and calcareous soil in millet cropping. Rhizosphere, **24** (1), 100598, **2022.**
- 13. RAMZANI P.M.A., SHAN L., ANJUM S., RONGGUI H., IQBAL M., VIRK Z.A., KAUSAR S. Improved quinoa growth, physiological response, and seed nutritional quality in three soils having different stresses by the application of acidified biochar and compost. Plant Physiology and Biochemistry, 116 (138), 2017.
- AGBNA G.H., DONGLI S., ZHIPENG L., ELSHAIKH N.A., GUANGCHENG S., TIMM L.C. Effects of deficit irrigation and biochar addition on the growth, yield, and quality of tomato. Scientia Horticulturae, 222 (101), 2017.
- 15. BAMMINGER C., POLL C., SIXT C., HÖGY P., WÜST D., KANDELER E., MARHAN S. Short-term response of soil microorganisms to biochar addition in a temperate agroecosystem under soil warming. Agriculture, Ecosystems & Environment, 233 (317), 2016.
- HAIDER G., KOYRO H.W., AZAM F., STEFFENS D., MÜLLER C., KAMMANN C. Biochar but not humic acid product amendment affected maize yields via improving plant-soil moisture relations. Plant and Soil, 395 (157), 2015.
- CHENG H., JONES D.L., HILL P., BASTAMI M.S., TU C.L. Influence of biochar produced from different pyrolysis temperature on nutrient retention and leaching. Archives of Agronomy and Soil Science, 64 (6), 850, 2018.
- 18. XIAO L., MENG F. Evaluating the effect of biochar on salt leaching and nutrient retention of Yellow River Delta soil. Soil Use and Management, **36** (4), 740, **2020**.
- BACHMANN H.J., BUCHELI T.D., DIEGUEZ-ALONSO A., FABBRI D., KNICKER H., SCHMIDT H.P., ZEHETNER F. Toward the standardization of biochar analysis: the COST action TD1107 interlaboratory comparison. Journal of Agricultural and Food Chemistry, 64 (2), 513, 2016.
- 20. HAFEEZ A., PAN T., TIAN J., CAI K. Modified biochars and their effects on soil quality: a review. Environments, 9 (5), 60, 2022.
- 21. NGUYEN T.H., PHAM T.H., NGUYEN T.H.T., NGUYEN T.N., NGUYEN M.V., TRAN DINH T., THI V.H.T. Synthesis of iron-modified biochar derived from rice straw and its application to arsenic removal. Journal of Chemistry, 1 (8), 2019.
- 22. JANU R., MRLIK V., RIBITSCH D., HOFMAN J., SEDLÁČEK P., BIELSKÁ L., SOJA G. Biochar surface functional groups as affected by biomass feedstock, biochar composition and pyrolysis temperature. Carbon Resources Conversion, 4 (46), 2021.
- 23. ZHANG H., VORONEY R.P., PRICE G.W., WHITE A.J. Sulfur-enriched biochar as a potential soil amendment and fertiliser. Soil Research, 55 (1), 93, 2016.

- EL-SHARKAWY M., EL-NAGGAR A.H., AL-HUQAIL A.A., GHONEIM A.M. Acid-modified biochar impacts on soil properties and biochemical characteristics of crops grown in saline-sodic soils. Sustainability, 14 (13), 8190, 2022.
- 25. SAHIN O., TASKIN M.B., KAYA E.C., ATAKOL O.R.H.A.N., EMIR E., INAL A., GUNES A.Y.D.I.N. Effect of acid modification of biochar on nutrient availability and maize growth in a calcareous soil. Soil Use and Management, 33 (3), 447, 2017.
- PEREIRA S.I., MOREIRA H., ARGYRAS K., CASTRO P.M., MARQUES A.P. Promotion of sunflower growth under saline water irrigation by the inoculation of beneficial microorganisms. Applied Soil Ecology, 105 (47), 2016.
- HUANG Z., LIU Y., TIAN F.P., WU G.L. Soil water availability threshold indicator was determined by using plant physiological responses under drought conditions. Ecological Indicators, 118, 106740, 2020.
- 28. ABD EL-MAGEED T.A., RADY M.M., TAHA R.S., ABD EL AZEAM S., SIMPSON C.R., SEMIDA W.M. Effects of integrated use of residual sulfur-enhanced biochar with effective microorganisms on soil properties, plant growth and short-term productivity of Capsicum annuum under salt stress. Scientia Horticulturae, 261, 108930, 2020.
- 29. ALOTAIBI M.O., IKRAM M., ALOTAIBI N.M., HUSSAIN G.S., GHONEIM A.M., YOUNIS U., DANISH S. Examining the role of AMF-Biochar in the regulation of spinach growth attributes, nutrients concentrations, and antioxidant enzymes in mitigating drought stress. Plant Stress, 10, 100205, 2023.
- 30. HUANG S., HUANG P., HAREEM M., TAHZEEB-UL-HASSAN M., YOUNIS U., DAWAR K. Evaluating the hidden potential of deashed biochar in mitigating salinity stress for cultivation of fenugreek. Scientific Report, 14, 141, 2024.
- GUPTA A., BANO A., RAI S., MISHRA R., SINGH M., SHARMA S. Mechanistic insights of plant-microbe interaction towards drought and salinity stress in plants for enhancing the agriculture productivity. Plant Stress, 4, 100073, 2022.
- 32. ADIL M., NOOR SHAH A., KHAN AN., YOUNAS T., SAJID MEHMOOD M., MAHMOOD A. Amelioration of harmful effects of soil salinity on plants through silicon application: a review. Pakistan Journal of Botany, 55 (10), 2023.
- 33. BINTI ABD RAHMAN S.N., ABDUL RAZAK M.S.F., BIN B., OTHMAN M.R., ABD RAHIM N.I.B. Screening of Mardi rice accessions for salinity tolerance at seedling stage based on growth performance and molecular analysis. Pakistan Journal of Botany, 55 (902), 2023.
- 34. MA Y., HUANG P., HUANG S., YOUNIS U., HUSSAIN G.S., FAHAD S. γ-Aminobutyric acid (GABA) and ectoine (ECT) impacts with and without AMF on antioxidants, gas exchange attributes and nutrients of cotton cultivated in salt affected soil. BMC Plant Biology, 23 (476), 2023.
- 35. ABU QAOUD H., ABU ALI I., AL-FARES H., QUBBAJ T., SHTAYA J.Y.M. Effect of salinity on the growth and some morphological traits of pearl millet. Pakistan Journal of Botany, 55 (1), 2023.
- 36. KHALIQ M., NAWAZ K., HUSSAIN K., JAVERIA M., IQBAL I., ARSHAD N. Foliar application of sorbitol is a shotgun approach to alleviate the adverse effects of salinity stress on two varieties of wheat (Triticum aestivum L). Pakistan Journal of Botany, 55 (1243), 56, 2023.

- 37. SHEREEN A., ASMA A., SHIRAZI M.U., KHAN M.A., ALI M., ARIF M. Physio-biochemical analysis of salinity tolerance in sodium contrasting rice (Oryza sativa L.) genotypes Pakistan Journal of Botany, **54** (787), 94, **2022**.
- 38. NAWAZ M.S., SAMI S.A., BANO M., KHAN M.R.Q., ANWAR Z., IJAZ A. Impact of salt stress on cotton. International Journal of Agriculture and Biosciences, 12 (98), 103, 2023.
- 39. BOUMAAZA B., GACEMI A., BENZOHRANE I.E., BENADA M., BOUDALIA S., BELAIDI H. Impact of salinity on the behavior of fungi. International Journal of Agriculture and Biosciences, 11 (139), 47, 2022.
- 40. RAZA A., SALEHI H., RAHMAN M.A., ZAHID Z., MADADKAR HAGHJOU M., NAJAFI-KAKAVAND S. Plant hormones and neurotransmitter interactions mediate antioxidant defenses under induced oxidative stress in plants. Frontier Plant Science, 13, 961872, 2022.
- ZAFAR-UL-HYE M., YASEEN R., ABID M., ABBAS M., AHMAD M., RAHI A.A. Rhizobacteria having ACCdeaminase and biogas slurry can mitigate salinity adverse effects in wheat. Pakistan Journal of Botany, 54 (297), 303, 2022.
- 42. IKRAM M., RAUF A., RAO M.J., MAQSOOD M.F.K., BAKHSH M.Z.M., ULLAH M. CRISPR-Cas9 based molecular breeding in crop plants: a review. Molecular Biology Reports, **51** (1), 227, **2024.**
- 43. SABAGH A.E., HOSSAIN A., ISLAM M.S., IQBAL M.A., FAHAD S., RATNASEKERA D. Consequences and mitigation strategies of heat stress for sustainability of soybean (Glycine max L. Merr.) production under the changing climate. In Plant stress physiology, IntechOpen, 2020.
- 44. PATEL M.R., PANWAR N.L. Biochar from Agricultural Crop residues: Environmental, Production, and Life Cycle Assessment Overview. Resources, Conservation and Recycling Advances, 200173, 2023.
- 45. HAYAT K., KHAN W.M., KHAN M.N., SHAH S.N. Pollen morphological investigation of selected species of family Asteraceae from Pakistan by using light and scanning electron microscopy. Microscopy Research and Technique, 86, 2023.
- 46. ABBAS N., ZAFAR M., AHMAD M., ALTHOBAITI A.T., RAMADAN M.F., MAKHKAMOV T., GAFFOROV Y., KHAYDAROV K., KABIR M., SULTANA S. Tendril anatomy: a tool for correct identification among Cucurbitaceous taxa. Plants, 11 (23), 3273, 2022.
- 47. LLAURADÓ MAURY G., MÉNDEZ RODRÍGUEZ D., HENDRIX S., ESCALONA ARRANZ J.C., FUNG BOIX Y., PACHECO A.O., GARCÍA DÍAZ J., MORRISQUEVEDO H.J., FERRER DUBOIS A., ALEMAN E.I. Antioxidants in plants: A valorization potential emphasizing the need for the conservation of plant biodiversity in Cuba. Antioxidants, 9 (11), 1048, 2020.
- 48. SHAHEEN H., QURESHI R., QASEEM M.F., BRUSCHI P. The fodder grass resources for ruminants: A indigenous treasure of local communities of Thal desert Punjab, Pakistan. PloS one, 15 (3), e0224061, 2020.
- 49. MAJEED M., BHATTI K.H., AMJAD M.S., ABBASI A.M., BUSSMANN R.W., NAWAZ F., RASHID A., MEHMOOD A., MAHMOOD M., KHAN W.M. Ethnoveterinary uses of Poaceae in Punjab, Pakistan. PloS one, 15 (11), e0241705, 2020.
- 50. RAHMAN S.U., ULLAH Z., ALI A., AHMAD M., SHER H., SHINWARI Z.K., NAZIR A. Ethnoecological knowledge of wild fodder plant resources of district Buner Pakistan. Pakistan Journal of Botany, 54 (2), 645, 2022.

- 51. MAROYI A. Traditional uses of wild and tended plants in maintaining ecosystem services in agricultural landscapes of the Eastern Cape Province in South Africa. Journal of Ethnobiology and Ethnomedicine, 18 (1), 17, 2022.
- 52. SINGH K., SHARMA R., SAHARE H. Implications of synthetic chemicals and natural plant extracts in improving vase life of flowers. Scientia Horticulturae, 302 (111), 133, 2022.
- IKRAM M., UR REHMAN H., SOYSAL S., AAMIR M., ISLAM M.S., KUMARI A. Impact of climate change on cotton growth and yield. Cotton Production Under Abiotic Stress, 3, 22, 2021.
- 54. GEBASHE F., AREMU A.O., GRUZ J., FINNIE J.F., VAN STADEN J. Phytochemical profiles and antioxidant activity of grasses used in South African traditional medicine. Plants, 9 (3), 371, 2020.
- 55. COTAS J., LEANDRO A., MONTEIRO P., PACHECO D., FIGUEIRINHA A., GONÇALVES A.M., DA SILVA G.J., PEREIRA L. Seaweed phenolics: From extraction to applications. Marine drugs, 18 (8), 384, 2020.
- 56. RAFIQ S., KAUL R., SOFI S., BASHIR N., NAZIR F., NAYIK G.A. Citrus peel as a source of functional ingredient: A review. Journal of the Saudi Society of Agricultural Sciences, 17 (4), 351, 2018.
- 57. SHARIFI-RAD M., VARONI E.M., IRITI M., MARTORELL M., SETZER W.N., DEL MAR CONTRERAS M., SALEHI B., SOLTANI-NEJAD A., RAJABI S., TAJBAKHSH M. Carvacrol and human health: A comprehensive review. Phytotherapy Research, 32 (9), 1675, 2018.

- 58. FEKRI A., TORBATI M., KHOSROWSHAHI A.Y., SHAMLOO H.B., AZADMARD-DAMIRCHI S. Functional effects of phytate-degrading, probiotic lactic acid bacteria and yeast strains isolated from Iranian traditional sourdough on the technological and nutritional properties of whole wheat bread. Food Chemistry, 306, 125620, 2020.
- SCARANO P., TARTAGLIA M., ZUZOLO D., PRIGIONIERO A., GUARINO C., SCIARRILLO R. Recovery and valorization of bioactive and functional compounds from the discarded of Opuntia ficus-indica (L.) mill. fruit peel. Agronomy, 12 (2), 388, 2022.
- 60. CHENG Z., LU X., LIN F., NAEEM A., LONG C. Ethnobotanical study on wild edible plants used by Dulong people in northwestern Yunnan, China. Journal of Ethnobiology and Ethnomedicine, 18 (1), 1, 2022.
- 61. BHAT M.N., SINGH B., SURMAL O., SINGH B., SHIVGOTRA V., MUSARELLA C.M. Ethnobotany of the Himalayas: Safeguarding medical practices and traditional uses of Kashmir regions. Biology, 10 (9), 851, 2021.
- 62. MINHAS A., SAEED M.S., EHSAN A., MURTAZA G., AKRAM M., MEHRAN M., SYED R.K.I.A. Response of integrated use of humic acid and chemical fertilizer on growth and yield of rice crop (Oryza sativa L.) in calcareous soil. Pakistan Journal of Botany, 56, 3, 2024.
- 63. HAFEZ M., MOHAMED A.E., RASHAD M., POPOVA I. The efficiency of application of bacterial and humic preparations to enhance of wheat (Triticum aestivum L.) plant productivity in the arid regions of Egypt. Biotechnology Reports, 29, e00584, 2021.