

Original Research

# Comparative Evaluation of Biological Activities and Allelopathic Potential of Java Plum Plant Parts

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## Abstract

Plants are vital in maintaining the food chain and have a significant impact on surrounding microbes due to the presence of allelochemicals. The potential allelopathic effects of java plum plant parts were assessed on seed germination and seedling growth of soybean, cucumber, carrot, rice, wheat, and maize. The results indicated that *S. cumini* significantly retarded the germination of soybean, cucumber, and rice while stimulating carrot germination. *S. cumini* had a diverse effect on the seedling growth of soybean, rice, and carrot, with an inhibitory effect on carrot growth. Significant reduction in radicle and hypocotyl growth of soybean and rice was observed. Aqueous extracts of *S. cumini* plant parts were also tested for antibacterial activity against *Escherichia coli* and *Acinetobacter baumannii* and antifungal activity against *Aspergillus niger* and *Candida albicans*. Stem, seed, and fruit extracts of *S. cumini* inhibited the growth of *A. baumannii* at the highest concentration, while leaf extract inhibited the growth of *E. coli*. Stem and fruit extracts had negative effects on the growth of *A. niger*. Knowing the significant allelopathic and antimicrobial potential of *S. cumini*, further studies on its allelochemicals can help in using this plant species for agricultural and medicinal purposes.

**Keywords:** allelochemical, antimicrobial, germination, growth, *Syzygium cumini*

## Introduction

*Syzygium cumini* (L.) Skeels, commonly known as Java Plum/Jamun, are a fruit-bearing crop belonging to the Myrtaceae family. The plant is native to South Asia,

mainly Pakistan, India, Afghanistan and Australia. It is also cultivated in Florida and Kenya. Java plum is an important indigenous minor fruit species that grows under a variety of agro-climatic conditions [1]. During ripening, the fruit is greenish and, at maturity, pink to shining crimson. The harvesting period of the fruit is usually in the monsoon season (June to July) and lasts for 30 to 40 days [2].

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*S. cumini* is a medicinally and nutritionally important fruit crop. Its fruits are rich in carbohydrates, fiber, proteins, vitamins, minerals, fats, and calories [2]. Plants have various secondary metabolites, which are unprocessed by-products, including bioactive substances. Extract of the Jamun fruit has been used to treat a variety of illnesses and ailments and has anti-diabetic properties [3]. Allelochemicals may be present in leaves, flowers, roots, fruits, or stems. They can also be found in the surrounding soil after the decomposition of the allelopathic plant residues. The allelopathic interactions are deleterious to their surrounding plants, but these might also be involved in giving an advantage to selective plants [4]. In allelopathy research, inhibitory substances were often argued to explain the growth pattern, while other substances remained neglected [5].

On the other hand, the antimicrobial activities of some plant species have been widely researched. For example, the crude extracts of cinnamon, garlic, basil, curry, ginger, sage, mustard, and other herbs exhibited antimicrobial properties against a wide range of Gram-positive and Gram-negative bacteria [6-7]. Antimicrobial resistance arises when some of the micro-organisms that cause infection (pathogens) adapt to survive exposure to a medicine that would normally kill them or stop their growth [8].

The present studies were undertaken to investigate the allelopathic potential of various parts of java plum (*S. cumini*) against the growth of economically important crops and to evaluate the antimicrobial potential of various parts of *S. cumini* against pathogenic fungal and bacterial strains.

## Material and Methods

Various parts (leaf, bark, fruit, stem, and seed) of the java plum tree were collected, washed thoroughly with water, and air-dried for 25-30 days. The dried sample was ground into a fine powder and stored in dry condition until used. The aqueous extracts of the plant parts were evaluated for their allelopathic and antimicrobial potential by using the filter paper method and the agar well diffusion method, respectively. Different concentrations, such as 0.5g, 1g, and 2 g of fine powder of each plant part, were weighed separately. To prepare the aqueous extracts, weighed materials were soaked in 100 ml of distilled water overnight, and then the extracts were filtered with the help of a Whatman filter paper. The filtrate was stored in the refrigerator for further use.

### Allelopathic Evaluation

The seeds of carrot (*Daucus carota* L.), cucumber (*Cucumis sativus* L.), rice (*Oryza sativa* L.), soybean (*Glycine max* L.), wheat (*Triticum aestivum* L.), and maize (*Zea mays* L.) were used to investigate the allelopathic potential of all the five parts of java plum

plant against it. The filter paper method is effective and easy to handle. Different extracts of 0.5%, 1%, and 2% concentrations were applied to the test by soaking a double layer of filter papers. For sterilization of medium from dust particles or fungal attack, Petri dishes were cleaned with ethanol-dipped cotton. Ten seeds of the test plant were placed between two folds of filter paper in each petri dish. Extracts of each plant species were applied to seeds. Distilled water was used for the control set instead of extracts. The experiment with each treatment was replicated five times.

Petri dishes of the experiment were set at 25°C. The experiment extended over a period of seven days. The result was determined by counting germinated seeds and measuring the length of the root and shoot. The length of the hypocotyl and radicle of seedlings was measured with a millimeter (mm) scale.

### Antimicrobial Evaluation

*Escherichia coli*, *Acinetobacter baumannii*, *Candida albicans*, and *Aspergillus niger* were used to check the antimicrobial activity of extracts of Java plum plant parts such as root, leaf, stem, bark, and fruit in different concentrations. For the agar well diffusion method, the agar plate surface was inoculated by spreading a volume of the microbial inoculum over the entire agar surface. Then, a hole with a diameter of 6 to 8 mm was punched aseptically with a sterile cork borer, and a volume (20–100 µL) of the antimicrobial agent or extract solution at the desired concentration was introduced into the well. Then, agar plates are incubated under suitable conditions depending on the test microorganism. The antimicrobial agent diffused in the agar medium and inhibited the growth of the microbial strain tested.

### Data Analysis

Data was analyzed using statistical software, SPSS 13.0 (Statistical Package for the Social Sciences) and Microsoft Excel. To determine the significance of the activity, a single-factor ANOVA (analysis of variance) was performed. The level of significance was set at 0.05. To visually represent the percentage germination and growth of the radicle and hypocotyl, a line graph was created.

## Results

Experiments were conducted to screen the allelopathic and antimicrobial potential of various parts of the java plum (*Syzygium cumini* (L.) Skeels). Different extracts, i.e., leaf, stem, bark, seed, and fruit, were used for quantitative analysis of allelopathic potential against the germination and growth of carrot (*Daucus carota* L.), cucumber (*Cucumis sativus* L.), rice (*Oryza sativa* L.), soybean (*Glycine max* L.), wheat (*Triticum aestivum* L.), and maize (*Zea mays* L.). These extracts

were also investigated to evaluate their antimicrobial potential against the inhibition of microbes, such as bacterial species (*Escherichia coli* and *Acinetobacter baumannii*) and fungal species (*Candida albicans* and *Aspergillus niger*).

### Evaluation of Allelopathic Potential

For the investigation of allelopathic activity, three different concentrations (0.5%, 1%, and 2%) of five different plant parts, i.e., leaf, stem, bark, seed, and fruit, were applied. Various effects were observed under the treatments of three different concentrations. Percentage germination along with lengths of hypocotyl and radicle of seedlings were recorded by comparing with respective controls. Different extracts of the *S. cumini* plant were used to investigate the seed germination of wheat and maize. Donor plants displayed both stimulatory and inhibitory effects on seed germination of the test plants.

#### *Effect of Syzygium cumini on Seed Germination of Test Plant*

The effects of various extracts of *S. cumini* plant parts (leaf, stem, bark, seed, and fruit) were observed to study the germination percentage of carrot (*Daucus carota* L.), cucumber (*Cucumis sativus* L.), rice (*Oryza sativa* L.) soybean (*Glycine max* L.), wheat (*Triticum aestivum* L.) and maize (*Zea mays* L.). Among all the test species, germination was observed to be increased under the influence of java plum plant extract. Greater effects were shown with bark extract having a 1% concentration. The rest of the crops had non-significant effects on seed germination under the influence of plant part extracts.

#### *Effect on Seed Growth of Test Plants*

All extracts of java plum showed inhibition of wheat at higher concentrations. Leaf extracts showed a negligible effect. Stem and bark extracts inhibited the seed growth of wheat at all of its concentrations. All extracts of seed and fruit showed stimulation of seed growth at all concentrations.

*Syzygium cumini* extracts have diverse effects on the growth of soybean (*Glycine max* L.). Results revealed that the bark extract had the most significant impact, causing maximum retardation of soybean growth across all concentrations (Fig. 1). Leaf extract showed varied effects on the growth of soybean. Stem extract at 1% concentration increased growth, while other concentrations negatively affected hypocotyl and radical length. The seed extract of 0.5% concentration increased the hypocotyl and radical length. Fruit extract overall stimulated soybean growth. Radical length showed the highest elevation against 0.5%, 1%, and 2% fruit extract. ANOVA confirmed the significant effect of *S. cumini* on radical and plumule length at all concentrations (Table 1).

Aqueous extracts of different parts of the *S. cumini* plant were found to inhibit the growth of cucumber (*Cucumis sativus*) at all concentrations. The leaf extract exhibited the maximum reduction in hypocotyl and radical length. Stem extracts at 0.5% showed a significant decrease in both hypocotyl and radical length (Fig. 1). Seed extract of 0.5% concentration caused the highest reduction in radicle length. Bark and fruit extract 2% concentration had the highest reduction in hypocotyl length and caused significant reductions in radicle length as well. However, the ANOVA showed that the effects of *S. cumini* extracts on cucumber hypocotyl and radical length were not statistically significant at all concentrations (Table 1).

*S. cumini* (plant parts) exhibited diverse effects on the growth of carrots at different concentrations. The leaf extract had the highest stimulatory effect, increasing hypocotyl length in a concentration-dependent manner. The highest stimulation of radical length (46%) was observed with 1% leaf extract (Fig. 1). Bark extract at 1% concentration also resulted in a maximal increase in hypocotyl length (30%). However, 0.5% extract had the highest reduction in radical length (51%). Stem extracts initially increased and then decreased hypocotyl length, with 2% concentration inhibiting growth by 15%. Seed extract at 1% increased hypocotyl length by 14%, while other concentrations decreased it. Fruit extract caused reductions in hypocotyl length, with the highest reduction at 2% concentration. The ANOVA indicated that the effects of *S. cumini* on carrot hypocotyl and radical length were not statistically significant at all concentrations (Table 1).

Various concentrations of *Syzygium cumini* extracts had varied effects on the growth of rice (*O. sativa*). Leaf extract showed negligible effects, except for a 19% increase in radical length with 0.5% extract. Stem extract at 2% caused a reduction of 26% in radical length. Bark extracts at 0.5% and 1% concentrations caused reductions in hypocotyl length. Similarly, seed extracts at 1% and 2% concentrations reduced the radical length by 17% and 18%, respectively. Fruit extract decreased hypocotyl and radical length by 35% with 2% concentration (Fig. 1). ANOVA indicated that *Syzygium cumini* extracts had a non-significant effect on rice's radical and hypocotyl length at all concentrations (Table 1).

*S. cumini* (java plum) had a diverse effect on the growth of wheat. The leaf extract stimulated hypocotyl length (65%) at 1% concentration but declined at 2% concentration. The stem extract inhibited the growth of hypocotyl and radicle, with varying degrees of inhibition at different concentrations. The bark extract also showed inhibition at 0.5% and 1% concentrations and a decrease in hypocotyl length at 2% concentration. The radicle length increased with increasing concentration. The seed extract exhibited inhibition at 0.5% concentration and stimulated hypocotyl growth at 1% and 2% concentrations. The radicle length showed inhibitory effects at 1% concentration. The fruit extract showed

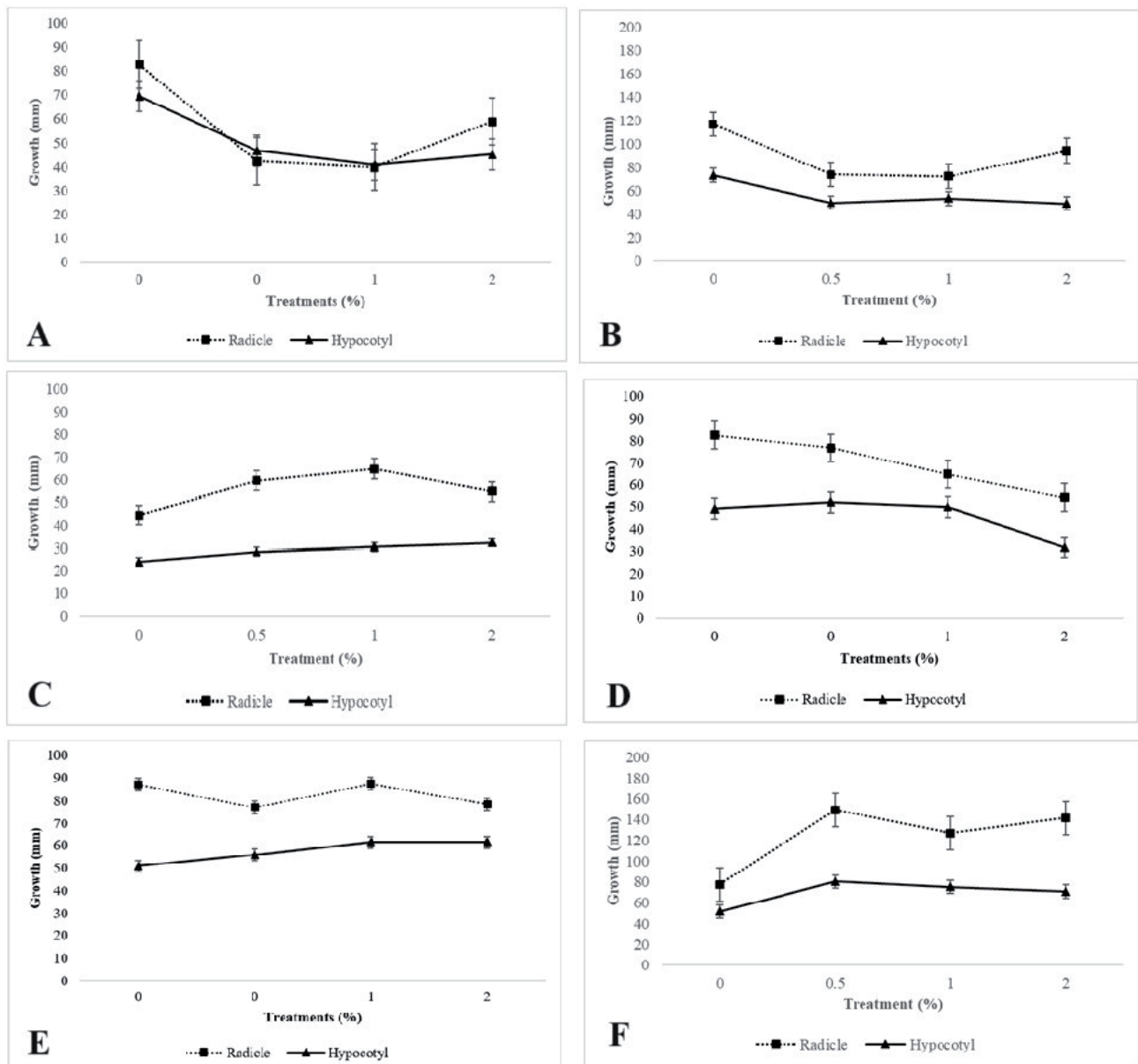


Fig. 1. Allelopathic Effect of Java Plum Plant Parts on various Test Species (A) Bark Extract vs. Soybean (B) Leaf Extract vs. Cucumber (C) Leaf Extract vs. Carrot (D) Fruit Extract vs. Rice (E) Fruit Extract vs. Wheat (F) Stem Extract vs. Maize.

the highest inhibition at 1% and 2% concentrations, particularly affecting hypocotyl length (Fig. 1). The study revealed diverse effects of the extracts on wheat growth, with the fruit extract exhibiting the most significant impact on radicle length (Table 1).

*S. cumini* (java plum) had a diverse effect on the growth of maize. The leaf and bark extracts exhibited inhibitory effects on both hypocotyl and radicle growth of maize, with increasing inhibition at higher concentrations. The stem extract, on the other hand, stimulated the length of maize hypocotyl but showed inhibitory effects on radicle growth (Fig. 1). The seed extract displayed a mix of stimulatory and inhibitory effects on hypocotyl and radicle growth. The fruit extract showed a stimulatory effect on hypocotyl length and increased radicle length with increasing

concentration. Overall, the extracts of *S. cumini* exhibited diverse effects on the growth of maize, with variations depending on the plant part and concentration (Table 1).

#### Evaluation of Antimicrobial Potential

Three different concentrations (0.5%, 1%, and 2%) of *S. cumini* plant parts, i.e., leaf, stem, bark, seed, and fruit, were investigated to evaluate their antibacterial activity against *Escherichia coli* and *Acinetobacter barmannii*, as well as antifungal activity against *Aspergillus niger* and *Candida albicans*. The zone of inhibitions was observed under the treatments of three different concentrations. Results were recorded by measuring the zone of inhibition by comparing them

Table 1. Allelopathic Effect of Java Plum Plant Parts on Growth of Various Test Species.

Source	DF	Test Species (MS Value)											
		Soybean		Cucumber		Carrot		Rice		Wheat		Maize	
		H	R	H	R	H	R	H	R	H	R	H	R
Plant parts	4	1054.56***	3595.10**	64.87 <sup>ns</sup>	185.20 <sup>ns</sup>	994.97***	64.87 <sup>ns</sup>	29.50 <sup>ns</sup>	725.71*	235.55 <sup>ns</sup>	190.12**	81.97 <sup>ns</sup>	660.86 <sup>ns</sup>
Treatments	3	134.25 <sup>ns</sup>	181.50 <sup>ns</sup>	919.05***	2138.24***	95.92 <sup>ns</sup>	919.05***	57.20 <sup>ns</sup>	1417.18***	1014.87***	188.21**	3063.89***	17220.63***
Plant parts * Treatments	12	175.617**	867.79*	47.72 <sup>ns</sup>	295.60 <sup>ns</sup>	181.36 <sup>ns</sup>	47.72 <sup>ns</sup>	73.67 <sup>ns</sup>	127.60 <sup>ns</sup>	130.76 <sup>ns</sup>	42.55 <sup>ns</sup>	116.30 <sup>ns</sup>	247.44 <sup>ns</sup>
Error	40												
Total	59												

DF= Degree of Freedom, MS=Mean Square, H=Hypocotyl, R=Radicle, ns=Non-significant, \*=Significant, \*\*/=\*\*\*=Highly Significant, Level of Significance = 5%

with anti-microbe medicines. For the control treatment, antibacterial amoxil and antifungal axicon were used.

#### Antibacterial Activity of Java Plum Extracts

The study examined the antibacterial activity of *S. cumini* (java plum) leaf, stem, bark, and fruit extracts against *Escherichia coli* and *Acinetobacter barmannii* (Fig. 2-3). The leaf extract demonstrated significant antibacterial activity against *E. coli*, with an increasing zone of inhibition at higher concentrations. The zone of inhibition rates were 11mm, 15mm, and 18mm at 0.5%, 1%, and 2% concentrations, respectively. However, the leaf extract showed comparatively less effectiveness against *A. barmannii*. The stem extract showed promising results against both *E. coli* and *A. barmannii*, with an increasing zone of inhibition at higher concentrations. The bark extract exhibited variable antibacterial activity, while the seed extract showed a higher zone of inhibition against *E. coli* at higher concentrations. The fruit extract showed significant antibacterial activity against both *E. coli* and *A. baumannii*. Overall, the agar well diffusion method revealed that the plant extracts exhibited better antimicrobial activity against *A. baumannii* compared to *E. coli* in some cases.

#### Antifungal Activity of Java Plum Extract

The study investigated the antifungal activity of *S. cumini* (java plum) leaf, stem, bark, and fruit extracts against *Candida albicans* and *Aspergillus niger* (Fig. 4-5). The leaf extract demonstrated significant antifungal effects against *C. albicans*, with an increasing zone of inhibition at higher concentrations. The zone of inhibition rates were 12mm, 16mm, and 17mm at 0.5%, 1%, and 2% concentrations, respectively. However, the leaf extract showed comparatively less effectiveness against *A. niger*. The stem extract showed promising results against both *C. albicans* and *A. niger*, with an increasing zone of inhibition at higher concentrations. The bark extract exhibited variable antifungal activity, while the seed extract showed a higher zone of inhibition against *A. niger* at higher concentrations. The fruit extract showed significant antifungal activity against both *C. albicans* and *A. niger*. Overall, the agar well diffusion method revealed that the plant extracts exhibited better antifungal activity against *A. niger* compared to *C. albicans* in some cases.

#### Discussion

Allelopathy refers to the phenomenon in which a plant releases certain biochemicals, known as allelochemicals, that can influence the growth, survival, and reproduction of other plant species. These allelochemicals can have both positive and negative effects on the target plant species [9]. It is estimated that higher plants can produce approximately 10,000 distinct allelochemicals, each with



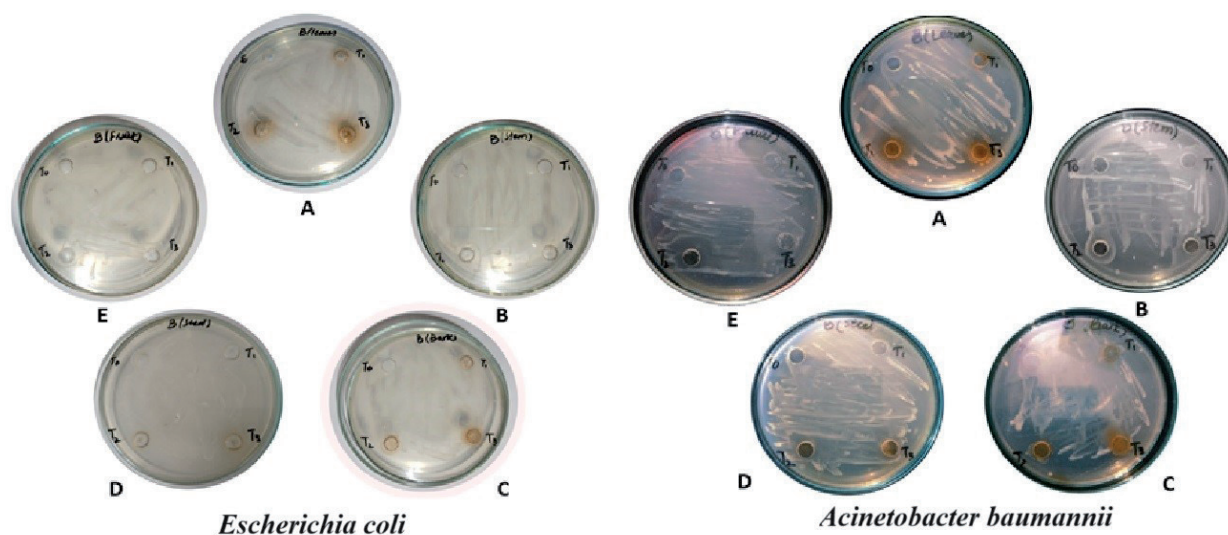


Fig. 2. Antibacterial activity of Java Plum Plant Parts against *Escherichia coli* and *Acinetobacter baumannii*.  
Note: A: Leaf Extract B: Stem Extract C: Bark Extract D: Seed Extract E: Fruit Extract.

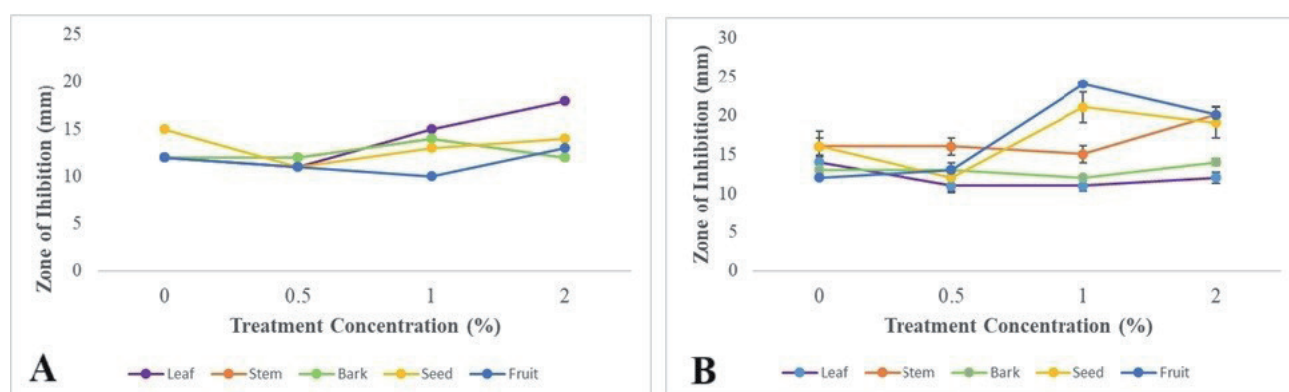


Fig. 3. Effect of Java Plum Plant Parts on (A) *Escherichia coli* (B) *Acinetobacter baumannii*.

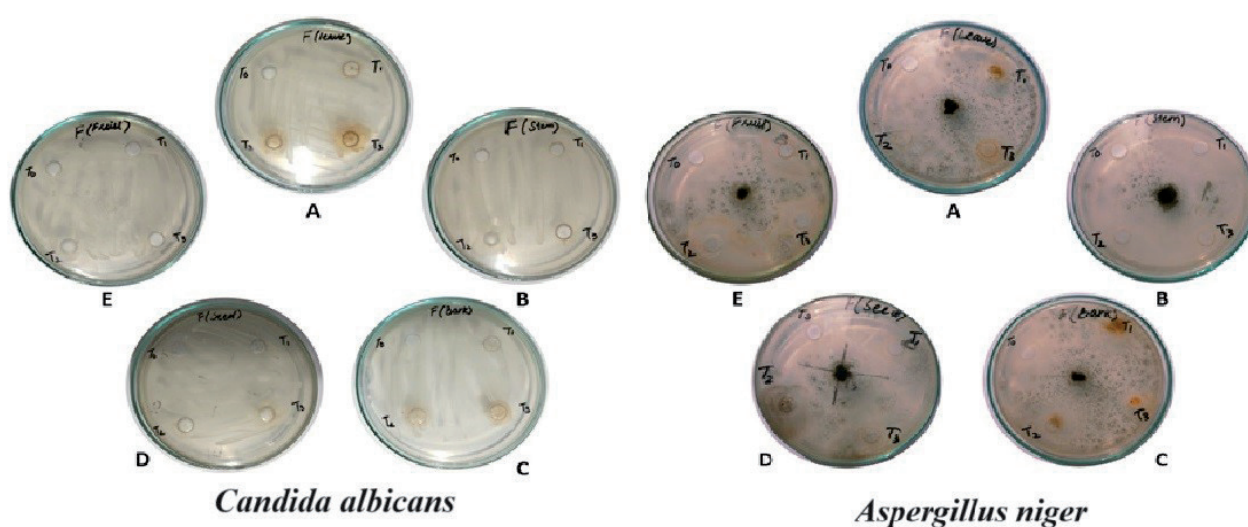


Fig. 4. Antibacterial activity of Java Plum Plant Parts against *Candida albicans* and *Aspergillus niger*.  
Note: A: Leaf Extract B: Stem Extract C: Bark Extract D: Seed Extract E: Fruit Extract.

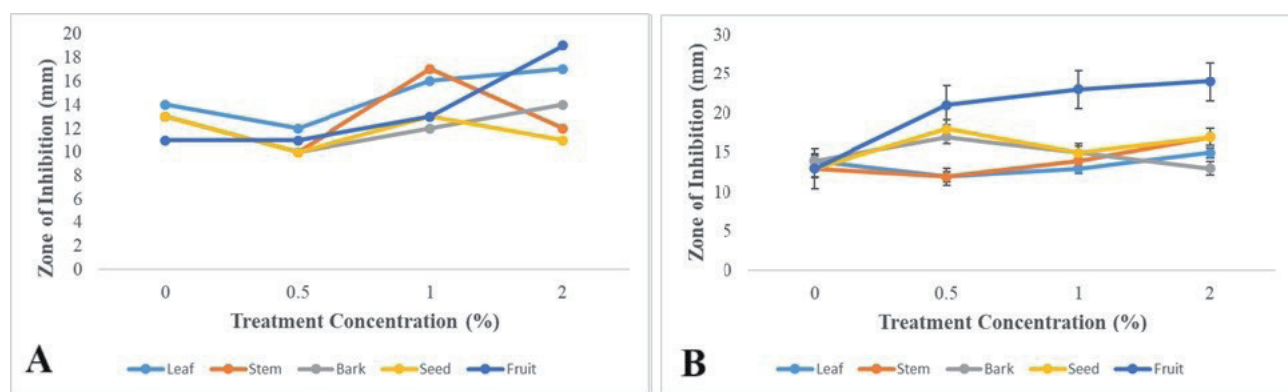


Fig. 5. Effect of Java Plum Plant Parts on (A) *Candida albicans* (B) *Aspergillus niger*.

its own unique activity and mode of action in receptor plants [10-11]. The allelopathic effects of *S. cumini* are likely attributable to the presence of allelochemicals in various parts of the plant, including the leaf, stem, bark, seed, and fruit. The present study aimed to investigate the allelopathic potential of different components of *S. cumini* on the germination and growth of soybean, cucumber, carrot, rice, wheat, and maize seedlings.

The present findings of soybean also agreed with Shi *et al.* [12], who reported the concentration-dependent effect of *Abutilon theophrasti* Medik on the germination and growth of *Glycine max*, *Triticum aestivum* and *Zea mays* L. Antioxidant enzyme and root system activities might be responsible for inhibitory or stimulatory response on test species. Siyar *et al.* [13] reported that a higher concentration of leaf and bark extracts of *Dalbergia sissoo* retarded the germination and growth of wheat. Similarly, Khan *et al.* [14] reported that aqueous leaf extract of rice and mustard restrained the growth of soybean at high extract concentrations and stimulated the growth at low concentrations. The different types of allelochemicals (phenolic acid, terpenes, diterpenoids, flavones, and p-coumaric acid) are present in rice. These chemicals repressed the energy metabolism by decreasing the cell division at the mitosis level (p-coumaric acid) and inhibiting water utilization (ferulic acid). As a result, germination and growth of test species were inhibited depending upon the concentration of allelochemicals [15-18]. Bari *et al.* [19] reported that carrot, garlic, ginger, and turmeric germination and growth are enhanced by aqueous extracts of *Dalbergia sissoo*.

The present results were consistent with the findings of Mondal *et al.* [20], who reported a reduction in germination and growth of rice at higher concentrations of leaf extract of *S. cumini*. Saeed *et al.* [21] also reported *S. cumini* exhibited an inhibitory effect on the growth of wheat at higher aqueous extract concentrations. The phytochemical analysis of *S. cumini* revealed that a greater number of alkaloids, flavonoids, glycosides, steroids, steroid phenols, tannins, and saponins were present in the leaves [22]. The different effect of allelochemicals depends upon the plant source from

which chemicals are liberated, the level of concentration and the recipient species [23-24].

The present study investigated the effects of aqueous extracts of *S. cumini* (java plum) on the growth and germination of wheat seedlings. The leaf extract stimulated wheat hypocotyl growth at all tested concentrations, consistent with Mali and Kanade [25] who found a similar stimulatory effect with *Cynodon dactylon* extract. However, the leaf extract showed inhibition of wheat radicle growth at the highest concentration, in line with Tripathy [26] and Al-Wakeel *et al.* [27], who observed inhibition with higher concentrations of *Acacia nilotica* extract.

Sisodia and Siddique [28] also reported inhibitory effects at higher concentrations of leaf extracts. It was observed that by increasing the aqueous extract concentrations, pH, total phenolic contents and osmotic potential were also increased. It may retard the growth of test species. The fruit extract exhibited inhibition of wheat hypocotyl growth at lower concentrations but stimulation at the highest concentration, as observed by Idu and Oghale [29] with *Ageratum conyzoides* extract. Mulatu *et al.* [30] reported inhibitory effects of *Parthenium hysterophorus* and *A. auriculiformis* extracts, respectively, supporting the present findings. The seed extract of *S. cumini* showed inhibition in wheat seed germination across all tested concentrations, similar to Arslan and Uremis [31], who observed inhibition with rapeseed extract. These findings suggest the presence of allelochemicals in *S. cumini* extracts that predominantly exert inhibitory effects on the growth and germination of wheat. Several types of research proved that allelopathic effects might depend on the plant tissues from which chemicals are liberated [32-33].

In the present study, the bark extract of *S. cumini* demonstrated a stimulating effect on maize growth at lower concentrations but showed inhibition at the highest concentrations. Similar effects were reported by Iqbal [34] with *Moringa oleifera* and *Brassica extracts*. Seigler *et al.* [35] highlighted the allelopathic effects of certain *Acacia* species on other crops, mainly attributed to phenolic compounds in their litter. These

allelochemical retarded the early growth of weed due to the inhibition of metabolic processes.

Gomaa *et al.* [36] reported negative effects of *Sonchus oleraceus* on crops and weeds, associated with phenols and alkaloids that can inhibit cell division and alter cell ultrastructure. The aqueous seed extracts of *S. cumini* significantly inhibited maize seed germination, with the seed extract exhibiting the highest inhibitory effect, consistent with Phuwiwat *et al.* [37], who observed inhibition with *Melia azedarach* leaf extract. Inhibitory compounds were reported by Carpinella *et al.* [38] and Harborne *et al.* [39] in *M. azedarach*, while Kato-Noguchi and Macias [40] suggested  $\alpha$ -amylase inhibition as a potential mode of action. On the other hand, the fruit extract of *S. cumini* stimulated maize growth, with higher concentrations showing greater stimulation in hypocotyl growth, aligning with Sisodia and Siddiqui [28], who found a stimulatory effect with *Croton bonplandianum* stem extract. Antibodies stimulating plant growth regulators and increased amino acid incorporation into proteins were proposed as mechanisms by Bouwmeester *et al.* [41] and Tang and Eaton [42].

In the present study, the seed extract of java plum (*S. cumini*) showed inhibitory effects against *E. coli*, with a zone of inhibition of 14 mm. The antimicrobial activity of the seed extracts varied depending on the concentration, with inhibitory effects ranging from 11 mm to 15 mm against *E. coli*. The stem and bark extracts of *S. cumini* displayed a 14 mm zone of inhibition against *E. coli* at a concentration of 1%, while the fruit extract showed a 14 mm zone of inhibition against *E. coli* at a concentration of 2%. These results are consistent with the antibacterial activity reported for *Allium sativum* [43] and *Mentha piperita* [44]. Both the stem and fruit extracts of *S. cumini* demonstrated inhibitory effects against *Acinetobacter baumannii*, with a maximum zone of inhibition of 20 mm observed at a concentration of 2%. However, at low concentrations, the extracts showed minimal or no inhibitory effects. It is important to note that the antimicrobial activity of the extracts may vary depending on the specific microorganism tested and the concentration of the extracts used. Discrepancies in results between different studies can be attributed to variations in the sources of microorganisms and other factors. Similar inhibitory effects of *S. cumini* extracts against *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Escherichia coli* were reported by Keskin and Toroglu [45].

In the present study, the extracts of different parts of *S. cumini* exhibited varying levels of antifungal activity against the tested fungal species. The aqueous leaf extract at a concentration of 1% showed a significant zone of inhibition of 17 mm for *Candida albicans* and 15 mm for *Aspergillus niger*. The antifungal activity of the leaf extract of *S. cumini* is likely attributed to the presence of tannins and other phenolic constituents. Previous research has reported that *S. cumini* is rich in gallic and ellagic acid polyphenol derivatives [46].

Acylated flavonol glycosides, kaempferol, myricetin, and other polyphenols have also been isolated from *S. cumini* leaves, which could contribute to the observed antifungal activity [47]. The significant antifungal effects of the aqueous leaf extract indicate the presence of specific antifungal chemicals in the leaves of *S. cumini*.

In the present study, the fruit extracts of *S. cumini* exhibited significant antifungal activity against *Aspergillus niger*. The aqueous fruit extracts showed clear zones of inhibition of 21 mm, 23 mm, and 24 mm at concentrations of 0.5%, 1%, and 2%, respectively. However, the fruit extracts were found to be comparatively less effective against *Candida albicans*. Previous research by Chandrasekaran and Venkatesalu [48] also reported the effectiveness of aqueous fruit extract of *S. cumini* against various fungal species, including *C. albicans*, *Aspergillus flavus*, *Aspergillus fumigatus*, and *Aspergillus niger*. This suggests that the chemicals present in the fruit of *S. cumini* possess specific antifungal activity. Anthocyanins, such as glucosides of delphinidin, petunidin, and malvidin found in the fruit peel, as well as phenolics present in the kernel and seed coatings, could be responsible for the observed antifungal activity of the fruit extracts [47].

## Conclusions

The results concluded that *Syzygium cumini* (L.) Skeels exhibit significant allelopathic and antimicrobial activity. It was observed that aqueous extract of *S. cumini* plant parts had diverse effects on carrot, rice, maize, and soybean growth. All extracts inhibited cucumber growth at all concentrations as compared to the control. The antimicrobial findings showed that the plant extracts exhibited stronger antimicrobial activity than commercially available antibiotics. For example, the fruit extract had the largest zone of inhibition (24 mm) against *A. niger* and *A. baumannii*. The study also revealed the potential antifungal effects of java plum seed and fruit extract on tested fungal species, particularly *Aspergillus niger*, as evidenced by the size of the inhibition zones. Further studies on the allelochemical of java plum plant parts can help in understanding its use for agricultural and medicinal purposes.

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## Conflict of Interest

The authors declare no conflict of interest.



## References

- ISHARTATI E., ROHMAN S. Bioactive properties and anti-diabetic potential of black jamun (*Syzygium cumini* (L.) Skeels) pulp and seed extracts. *Medicinal Plants-International Journal of Phytomedicines and Related Industries*, **14** (3), 421, **20**
- QAMAR M., AKHTAR S., ISMAIL T., WAHID M., ABBAS M.W., MUBARAK M.S., YUAN Y, BARNARD R.T., ZIORA M.Z., ESATBEYOGLU T. Phytochemical Profile, Biological Properties, and Food Applications of the Medicinal Plant *Syzygium cumini*. *Foods*, **11** (3), 378, **2022**.
- SINGH Y., BHATNAGAR P., KUMAR S. A review on bioactive compounds and medicinal strength of Jamun (*Syzygium cumini* Skeels). *International Journal of Chemical Study*, **7** (4), **2019**.
- OTMAINNA M., JURAIMI A.S., AHMAD-HAMDANI M.S., HASAN M., YEASMIN S., ANWAR M.P., ISLAM A.M. Allelopathic potential of tropical plants – a review. *Agronomy*, **13** (8), 2063, **2023**.
- MUSHTAQ W., SIDDIQUI M.B., HAKEEM K.R. Allelopathic control of native weeds. *In Allelopathy: Potential for Green Agriculture*, pp. 53, **2020**.
- CHASSAGNE F., SAMARAKOON T., PORRAS G., LYLES J.T., DETTWEILER M., MARQUEZ L., SALAM A.M., SHABIH S., FARROKHI D.R., QUAVE C.L. A systematic review of plants with antibacterial activities: A taxonomic and phylogenetic perspective. *Frontiers in Pharmacology*, **11**, 586548, **2021**.
- SOLIMAN A.M., YOON T., WANG J., STAFFORD J.L., BARREDA D.R. Isolation of skin leukocytes uncovers phagocyte inflammatory responses during induction and resolution of cutaneous inflammation in fish. *Frontiers in Immunology*, **12**, 725063, **2021**.
- AICH N., AHMED N., PAUL A. Issues of antibiotic resistance in aquaculture industry and its way forward. *International Journal of Current Microbiology and Applied Sciences*, **7** (8), 26, **2018**
- WANG R., DENG X., GAO Q., WU X., HAN L., GAO X., ZHAO S., CHEN W., ZHOU R., LI Z. *Sophora alopecuroides* L.: An ethnopharmacological, phytochemical, and pharmacological review. *Journal of Ethnopharmacology*, **248**, 112172, **2020**.
- KHAMARE Y., CHEN J., MARBLE S.C. Allelopathy and its application as a weed management tool: A review. *Frontiers in Plant Science*, **13**, 1034649, **2022**.
- SCAVO A., MAUROMICALE G. Crop allelopathy for sustainable weed management in agroecosystems: Knowing the present with a view to the future. *Agronomy*, **11** (11), 2104, **2021**.
- SHI S., CHENG J., AHMAD N., ZHAO W., TIAN M., YUAN Z., LI C, ZHAO C. Effects of potential allelochemicals in a water extract of *Abutilon theophrasti* Medik. on germination and growth of *Glycine max* L., *Triticum aestivum* L., and *Zea mays* L. *Journal of the Science of Food and Agriculture*, **103** (4), 2155, **2023**.
- SIYAR S., SAMI S., HUSSAIN F., HUSSAIN Z. Allelopathic effects of sheesham extracts on germination and seedling growth of common wheat. *Cercetări Agronomice în Moldova*, **4** (276), 17, **2019**.
- KHAN R.M.I., NAEEM M., ALI H.H., SHAHZAD M.A. Performance of Soybean against allelopathic leaf aqueous extracts and soil incorporated Residues. *Pakistan Journal of Botany*, **53** (4), 1441, **2021**.
- FAROOQ N., ABBAS T., TANVEER A., JABRAN K. Allelopathy for weed management. *In Co-evolution of Secondary Metabolites*, pp. 505, **2020**.
- HUSSAIN W.S., ABBAS M.M. Application of allelopathy in crop production. *In Agricultural Development in Asia-Potential Use of Nano-Materials and Nano-Technology*, **2021**.
- CHOUDHARY C.S., BEHERA B., RAZA M.B., MRUNALINI K., BHOI T.K., LAL M.K., NONGMAITHEM D., PRADHAN S., SONG B., DAS T.K. Mechanisms of allelopathic interactions for sustainable weed management. *Rhizosphere*, **25**, 100667, **2023**.
- PERVEEN A., RASHID M., MOKAL M.N., HASSAN N. Allelopathic Potential of Dominating Weed Species on Germination and Growth of Selected Winter Crops. *Journal Plantarum*, **6** (1), 19, **2024**.
- BARI M.S., RAHIM M.A., MIAN M.M.H. Allelopathic proclivities of *Dalbergia sissoo* on agricultural crops. *Journal of Agroforestry and Environment*, **4** (1), 59, **2010**.
- MONDAL S., DUARY B., MONDAL B., PANDA D. Allelopathic effect of aqueous leaf extracts of selected trees on germination and seedling growth of rice. *Journal of Environmental Biology*, **41** (2), 255, **2020**.
- SAEED H.S., RASUL F.S.M., MUBEEN M., NASIM W. Allelopathic Potential Assessment of Jaman (*Syzygium cumini* L.) on Wheat. *International Poster Journal of Science & Technology* **3** (1), 09-14, **2013**.
- TAMBE B.D., PEDHEKAR P., HARSHALI P. Phytochemical screening and antibacterial activity of *Syzygium cumini* (L.) (Myrtaceae) leave extracts. *Asian Journal of Pharmaceutical Research and Development*, **9** (5), 50, **2021**.
- ZAREEN S., FAWAD M., HAROON M., AHMAD I., ZAMAN A. Allelopathic potential of summer weeds on germination and growth performance of wheat and chickpea. *Journal of Natural Pesticide Research*, **1**, 100002, **2022**.
- KATO-NOGUCHI H., KATO M. Evolution of the secondary metabolites in invasive plant species *Chromolaena odorata* for the defense and allelopathic functions. *Plants*, **12** (3), 521, **2023**.
- MALI A.A., KANADE M.B. Allelopathic effect of two common weeds on seed germination, root-shoot length, biomass and protein content of jowar. *Annals of Biological Research*, **5** (3), 89, **2014**.
- TRIPATHY S. Effect of tree leaves' aqueous extracts on germination and seedling growth of soybean. *Allelopathy Journal*, **5**, 75, **1998**.
- AL-WAKEEL S.A.M., GABR M.A., HAMID A.A., ABUEL-SOUD W.M. Allelopathic effects of *Acacia nilotica* leaf residue on *Pisum sativum* L. *Allelopathy Journal*, **19** (2), 411, **2007**.
- SISODIA S., SIDDIQUI M.B. Allelopathic effect by aqueous extracts of different parts of *Croton bonplandianum* Baill. on some crop and weed plants. *Journal of Agricultural Extension and Rural Development*, **2** (1), 22, **2010**.
- IDU M., OGHAE O.U. Studies on the allelopathic effect of aqueous extract of *Ageratum conyzoides* Asteraceae on seedling growth of *Sesamum indicum* L. (Pedaliaceae). *Scitec Nutrition*, **2**, 1185, **2013**.
- MULATU B., APPLEBAUM S.W., KEREM Z., COLL M. Tomato fruit size, maturity and  $\alpha$ -tomatine content influence the performance of larvae of potato

- tuber moth *Phthorimaea operculella* (Lepidoptera: Gelechiidae). Bulletin of Entomological Research, **96** (2), 173, **2006**.
31. ARSLAN M., UREMIS İ. Allelopathic Potential of Rapeseed (*Brassica napus* L.) Cultivars on Weeds. In: Science Conference, pp. 593, **2018**.
  32. OYUN M.B. Allelopathic potentialities of *Gliricidia sepium* and *Acacia auriculiformis* on the germination and seedling vigour of maize (*Zea mays* L.). American Journal of Agricultural and Biological Science, **1** (3), 44, **2006**.
  33. EL-KHAWAS S.A., SHEHATA M.M. The allelopathic potentialities of *Acacia nilotica* and *Eucalyptus rostrata* on monocot (*Zea mays* L.) and dicot (*Phaseolus vulgaris* L.) plants. Biotechnology, **4** (1), 23, **2005**.
  34. IQBAL M.A. Role of moringa, brassica and sorghum water extracts in increasing crops growth and yield: A Review. American-Eurasian Journal of Agricultural & Environmental Sciences, **14**, 1150, **2014**.
  35. SEIGLER D.S. Phytochemistry of *Acacia* – sensu lato. Biochemical Systematics and Ecology, **31** (8), 845, **2003**.
  36. GOMAA A.A., KLUMPE H.E., LUO M.L., SELLE K., BARRANGOU R., BEISEL C.L. Programmable removal of bacterial strains by use of genome-targeting CRISPR-Cas systems. Molecular Biology & Microbiology, **5** (1), 10, **2014**.
  37. PHUWIWAT W., WICHITTRAKARN W., LAOSINWATTANA C., TEERARAK M. Inhibitory effects of *Melia azedarach* L. leaf extracts on seed germination and seedling growth of two weed species. Pakistan Journal of Weed Science Research, **18** (Special Issue), 485, **2012**.
  38. CARPINELLA M.C., FERRAYOLI C.G., PALACIOS S.M. Antifungal synergistic effect of scopoletin, a hydroxycoumarin isolated from *Melia azedarach* L. fruits. Journal of Agricultural and Food Chemistry, **53** (8), 2922, **2005**.
  39. HARBORNE J.B., SAITO N., DETONI C.H. Anthocyanins of cephaelis, cynomorium, euterpe, lavatera and pinanga. Biochemical Systematics and Ecology, **22** (8), 835, **1994**.
  40. KATO-NOGUCHI H., MACÍAS F.A. Effects of 6-methoxy-2-benzoxazolinone on the germination and  $\alpha$ -amylase activity in lettuce seeds. Journal of Plant Physiology, **162** (12), 1304, **2005**.
  41. BOUWMEESTER H.J., MATUSOVA R. ZHONGKUI S., BEALE M.H. Secondary metabolite signaling in host–parasitic plant interactions. Current Opinion in Plant Biology, **6** (4), 358 **2003**.
  42. TANG L., EATON J.W. Inflammatory responses to biomaterials. American Journal of Clinical Pathology, **103** (4), 466, **1995**.
  43. PATHMANATHAN M.K., UTHAYARASA K., JEYADEVAN J.P., JEYASEELAN E.C. In vitro antibacterial activity and phytochemical analysis of some selected medicinal plants. International Journal of Pharmaceutical & Biological Archive, **1** (3), 291, **2010**.
  44. WALTER C., SHINWARI Z.K., AFZAL I., MALIK R.N. Antibacterial activity in herbal products used in Pakistan. Pakistan Journal of Botany, **43**, 155, **2011**.
  45. KESKIN D., TOROGLU S. Studies on antimicrobial activities of solvent extracts of different spices. Journal of Environmental Biology, **32** (2), 251, **2011**.
  46. JAVAID A., SAMAD S. Screening of allelopathic trees for their antifungal potential against *Alternaria alternata* strains isolated from dying-back *Eucalyptus* spp. Natural Product Research, **26** (18), 1697, **2012**.
  47. JABEEN K., JAVAID A. Antifungal activity of *Syzygium cumini* against *Ascochyta rabiei*–the cause of chickpea blight. Natural Product Research, **24** (12), 1158, **2010**.
  48. CHANDRASEKARAN M., VENKATESALU V. Antibacterial and antifungal activity of *Syzygium jambolanum* seeds. Journal of Ethnopharmacology, **91** (1), 105, **2004**.