

Original Research

Deciphering the Genetic Code of Wheat Genotypes: A Regional Perspective in Pakistan on Morpho-Agronomic Traits and Protein Profiling

**Sadaf Iqbal¹, Bushra Muhammad Fordil², Ayesha Kayani¹, Dunia A Al Farraj³,
Mohamed S Elshikh³, Nadia Khan^{1*}, Abdul Waheed^{4o**}**

¹Department of Genetics, University of Karachi 75270, Pakistan

²School of Basic Medical Sciences, Shenzhen University Medical School, Shenzhen University,
Shenzhen, Guangdong, 518055, China

³Department of Botany and Microbiology, College of Science, King Saud University,
P.O. 2455, Riyadh 11451, Saudi Arabia

⁴Shenzhen Branch, Guangdong Laboratory for Lingnan Modern Agriculture, Genome Analysis Laboratory
of the Ministry of Agriculture, Agricultural Genomics Institute at Shenzhen, Chinese Academy of Agricultural Sciences,
Shenzhen, Guangdong, 518020, China

Received: 25 May 2024

Accepted: 16 February 2025

Abstract

Wheat is a staple diet due to its breadmaking quality, which is governed by seed storage proteins. This research evaluated genetic variability in morphological attributes and profiled gluten proteins in wheat genotypes. Sixty wheat varieties from Pakistan were assessed for agronomic traits, while the diversity of endosperm storage proteins in sixty-six genotypes was evaluated using SDS-PAGE. Moderate heritability for plant height (0.60) and flag leaf area (0.54) was noted in Khyber Pakhtunkhwa, while Punjab showed moderate to high heritability for heading days (0.67), plant height (0.64), flag leaf area (0.81), and flowering days (0.77). Sindh exhibited high heritability for spike length (0.88). A total of 61 alleles were found in 27 genotypes in Punjab, 49 in 15 genotypes in Sindh, and 48 in 24 genotypes in KP. Cluster analysis revealed Bhittai as the most diverse genotype in Sindh, DN lines in KP, and AS-02 in Punjab. These genotypes show significant diversity in gluten proteins, which are crucial for grain quality traits. The study concludes that wheat genotypes have sufficient variation in morpho-agronomic traits and protein profiling, which is useful for developing high-quality genotypes. Future research should explore the genetic basis of gluten protein diversity and its relationship with wheat quality traits using advanced genomic tools.

Keywords: genetic variability, environmental adaptability, regional diversity, sustainable agriculture, agro-ecological zones

*e-mail: nadiakhan@uok.edu.pk

**e-mail: 2021y90100044@caas.cn

°ORCID iD: 0000-0002-9136-3362

Introduction

Wheat has a high nutritional value since it is high in starch (58.2%), has enough sugar and fat, and has a higher percentage of carbohydrates than other crops [1]. In addition to its nutritional importance, wheat is a critical agricultural commodity. Pakistan, for instance, ranks among the top eight wheat-producing countries globally, with an annual production of 25 million tons [2]. Wheat accounts for 10.1% of agricultural value added and 1.7% of Pakistan's gross domestic product (GDP) [3]. Several agronomic traits influence the response of wheat grain yield [4]. Wheat yield improvement has been aided by agronomic parameters such as plant height (PH), harvest index (HI), total biomass (BM), number of productive tillers (PTN), grain number (GN), spike length (SL), and thousand-grain weight (TGW) [5-8]. Trait-based breeding of high-performing, genetically similar wheat genotypes has been essential for yield improvements [4, 9-12]. However, the widespread adoption of elite genotypes has led to reduced genetic diversity, a critical issue for sustainable agriculture. Genetic variety within fields, on the other hand, is a tool for more sustainable production since it allows greater stability and resilience to biotic and abiotic challenges [13]. Grain yield (GY) in wheat is complex and controlled by multiple genes. Effective selection requires understanding population variation, trait correlations, and environmental influences [14]. Correlation coefficient analysis is an important tool for providing information on the proper cause-and-effect relationship between yield and some yield components [15, 16]. Heritability refers to the proportion of phenotypic variation that can be largely attributed to a genetic makeup. Plant breeders can use heritability to estimate the behavior of the next generation and make favorable selections. The higher the heritability, the easier the selection process will become and the stronger the response toward selection [17-19]. Another key selection criterion that helps breeders in the selection procedure is genetic advancement (GA) [19, 20]. Genetic advance is another term for the expected response to selection. The optimum conditions for selection are those with high GA and high heritability estimates [21, 22]. The magnitude of variation in wheat breeding material was estimated using phenotypic (V_p) and genotypic variance (V_g), heritability (h^2_g), and genetic advance (GA) [23, 24]. The protein accumulated during grain development is not only an important reserve for plant germination and early seedling growth, but it is also an important food source for humans. Gluten protein, a mixture of monomeric gliadin and high-molecular-weight glutenin found in wheat starchy endosperm, is a major protein reservoir. The quality of the dough depends on the presence of wheat storage protein (gluten). Gliadins (Mw 30-80 kDa) and glutenins (Mw 30-80 kDa) are the two types of gluten (Mw 12-130 kDa). Glutenins are aggregating proteins that are important for dough strength and quality. Gliadins are nonaggregating

proteins responsible for dough extensibility [25-27]. Glutenin is classified into two classes: high-molecular-weight glutenin (HMW-GS) and low-molecular-weight glutenin (LMW-GS) [28, 29].

A substantial number of germplasm lines can be characterized for biochemical markers in a short period of time. Furthermore, the data more accurately reflect genetic diversity since biochemical markers are direct products of genes, and environmental factors do not affect their expression [30-32]. The variation in storage protein in wheat has been observed to be a beneficial tool to assess variability and improve diversity in germplasm collections [30, 33]. Wheat varieties with higher performance are committed to conventional wheat breeding techniques to develop wheat progeny with improved grain yield and baking quality attributes for end use [34, 35]. Keep in mind that the investigation is to characterize and identify the genetic diversity among wheat varieties grown in Pakistan based on their agronomic traits and differential protein profiles using the technique of SDS-PAGE.

Materials and Methods

Experimental Design

This study was carried out during the crop season (sown in November 2022 and harvested in April 2023) in the screen house. A total of sixty wheat varieties were grown in a screening house of the Department of Genetics, University of Karachi, Pakistan, while sixty-six wheat varieties were used for the qualitative separation of proteins. The plots were prepared by mixing sandy loam soil and manure in a 1:1 ratio. Each plot has a length of 213.36 cm and a width of 243.84 cm. Each variety was germinated in 3 replications. Urea was applied as a fertilizer during the tillering and flowering stages. Weeding was performed manually, and malathion spray was used to keep the plot disease-free. Ten plants were randomly selected and marked for each replication. After collecting the necessary data under field conditions, other observations have been recorded after harvesting.

Quantitative Traits and Measurements

The agronomic traits that were studied were heading (DTH) and flowering days (DTF), flag leaf area (FLA), spike length (SL), plant height (PH), spikelet number (SPKLT), number of grains per spike (GN), and thousand-grain weight (TGW). Heading days were recorded as the number of days from sowing until full exposure of spikes in 50% of the plot. Flowering was recorded as 50% of anthesis emergence in the plot. Flag leaf area (cm²) was measured when the leaves were fully developed and green. The height of the plant (cm) was measured between the base of the plant stand and the tip of the fully emerged spike, excluding awns. The

length of the spike (cm) was measured from the base of the first spike to the tip of the terminal spike, excluding awns at maturity. The spikelets and grain numbers were counted manually. Thousand-grain weights (g) (divide the total weight (g) by the total number of seeds, then multiply by 1000) were recorded on a top-loading balance.

Statistical Analysis

Correlation coefficients were calculated, and the analyses based on ANOVA and Duncan's multiple range test were performed using SPSS software for Windows version 20.0 (SPSS, Inc., Chicago, IL). The graphic presentation for the correlation was constructed using the R-Studio software. Heritability in the broad sense (h^2_B) was computed ($h^2_B = Vg/Vp$) for each trait and province [36]. Genetic advance ($GA = K \times (Vp)^{0.5} \times h^2_B$) was estimated at 5% selection intensity (K), i.e., 2.06 [37].

Protein Extraction and Electrophoresis

Wheat grains were ground to a fine powder, and Tris-HCl buffer (pH 7.5) was added to it in a 4:1 ratio in a microfuge tube (1.5 ml) and thoroughly mixed. Samples were kept at 0°C for 30 minutes, then boiled for 3 minutes and centrifuged at 8,000 rpm for 30 minutes. The supernatant was collected, and sample dilution buffer was added in a 2:1 ratio in the microfuge tube [38]. The variability of total storage proteins was analyzed using sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE), a 12% running gel, and a 4% stacking gel following the instructions in a TV 100 Scie-Plas vertical gel electrophoresis unit. Bromophenol blue (BPB) was used as a tracking dye to observe protein mobility in the gel. DireX Gene, a protein molecular weight marker with a typical range of 11 kDa to 180 kDa, was allowed to run at 100 V until the BPB blue line was visible at the bottom of the gel plates. After removing the gels, excess sodium dodecyl sulfate was removed by incubating the gels in 30% ethanol over a shaking water bath for 30 minutes. The gels were then rinsed twice with double-distilled water and fixed (10% acetic acid in 50% methanol) overnight. The running gel was gently shaken in the staining solution using an EYELA Uni Thermo Shaker NTS-1300 for 2 hours. The gels were stained with a Coomassie brilliant blue (CBB) solution and destained until the background became transparent and the bands sharpened. Gels were photographed after destaining by placing them near a white light.

Estimation of the Molecular Weight of an Unknown Protein

To calculate the molecular weight of an unknown protein, the relative migration distance (R_f) of the protein standards and the unknown protein was

estimated. The migration distance can be determined using the following Equation:

$$R_f = \frac{\text{migration distance of protein}}{\text{migration distance of dye front}}$$

Based on the values obtained for the bands in the standard, the logarithm of the molecular weight of the SDS-denatured polypeptide and its relative migration distance (R_f) are plotted into a graph. A linear plot can be generated for most proteins if the samples are fully denatured and the gel percentage is appropriate for the molecular weight range of the sample. The values of this graph were interpolated, and the molecular weight of the unknown protein band was given (Bio-Rad Bulletin 6040-2).

Using the Equation mentioned below, the molecular weight of an unknown protein was estimated.

$$y = mx + b$$

where y = log molecular weight of the unknown protein; m = slope of the line; $x = R_f$ value; and b = y-intercept of the line.

Data Analysis

For each wheat variety, electropherograms were scored, and the presence (1) and absence (0) of the band were noted. The final basic data structure consisted of a binary (0/1) data matrix, which was analyzed, and cluster analysis measured genetic diversity.

Results and Discussion

Analysis of variance showed a highly significant difference among genotypes recorded for all attributes: i.e., DTH, DTF, FLA, PH, SL, SPKLT, GN, and TGW (Table 1). The diversity implies that the analyzed experimental materials have considerable genetic diversity, which can be used in future wheat breeding efforts.

Agronomic Traits

Quantitative trait data for 60 wheat genotypes showed that AS-02 headed first (46 days) and NARC-11 last (90 days) (Table 2). Early heading allows more time for grain filling. Uqaab-00 had the largest flag leaf area (55 cm²), while Pirsabak-08 had the smallest (16.67 cm²). The flag leaf, crucial for photosynthesis, contributed 41- 43% to grain weight by increasing grain number and weight [39, 40]. Pakistan-13 had the tallest plants, with an average height of 96 cm, while the shortest plants were observed in TD-1, with an average height of 35 cm. Dwarf plants produce thicker stems, helping the plant to direct resources from the stem to the flower and grain filling, leading to an increased yield. Taller wheat

Table 1. ANOVA for quantitative traits in wheat.

Sources of Variance	D.F	DTH	DTF	FLA	PH	SL	SPKLT	GN	TGW
Genotypes	59	220.105***	349.775***	212.205**	617.133***	5.974***	22.621***	127.107***	193.928***
Reps	2	0,556	1,672	1,95	0,906	0,098	0,617	4,272	3,939
Error	120	2,394	3,772	6,479	2,333	0,181	0,761	2,972	4,817

D.F: degrees of freedom; DTH: Days To heading; DTF: Days to flowering; FLA: Flag leaf area; PH: Plant height; SL: Spike length; NS: Spikelet number: of spikelets spike-1; Grain Number: No. of grains spike-1 and TGW: 1000-grain weight; *** and ** Significant at $P \leq 0.001$ and $P \leq 0.05$ probability level.

plants compete better for sunlight but tend to fall under the weight of extra grain – a trait called lodging [41]. Barsaat had the longest spikes (13.39 cm) and most spikelets (23 per spike), while Imdad-05 had the shortest spikes (5.32 cm) and TD-1, Pirsabak-08, and Shalimar-88 had the fewest spikelets (12). Moomal-02 had the most grains (55 per spike), and Pirsabak-08 had the least (26). Marvi-00 had the heaviest 1000 grains (67 g), and Pirsabak-08 had the lightest (22.67 g). Barsaat excelled in spikelet number and spike length, Moomal-02 in grain number, and Marvi-00 in 1000-grain weight, making them valuable for breeding programs. Pirsabak-08 was the weakest variety overall, while Uqaab-00 excelled in spike length and flag leaf area.

Correlation Coefficient

Significant positive correlations were found between various traits (Fig. 1a)). Heading days correlated with DTF (0.605**), FLA (0.320**), PH (0.200**), SL (0.215**), SPKLT (0.311**), and GN (0.251**). Flowering days correlated with FLA (0.361**), PH (0.333**), and SPKLT (0.299**). Flag leaf area correlated with PH (0.562**) and SPKLT (0.300**). Spike length correlated with SPKLT (0.426**), GN (0.345**), and TGW (0.229**). The number of spikes correlated with GN (0.554**) and TGW (0.155*). Intensive selection of these traits will improve yield [14, 16]. These traits are interrelated; selecting one or more will improve the others. Due to the negative correlation between plant height and yield components, dwarf wheat varieties such as TD-1 should be preferred. They tolerate lodging and are advantageous for direct selection for grain yield [24]. Correlation studies are useful for improving yield by indirectly selecting yield-related traits [16].

Heritability Estimates

Due to their high heritability and relationship with grain yield, agricultural parameters can serve as indirect selection criteria in breeding and cultivar development [42]. Genetic parameters like GCV, PCV, h^2B , and GA are shown in Table 3. In Punjab, the flag leaf area (FLA) has high h^2B (0.81) and GA

(25.45). Flowering had an h^2B of 0.77 and a GA of 24.52. Heading days had moderate h^2B (0.67) and GA (17.68). PH, SL, and TGW had h^2B values of 0.64, 0.38, and 0.30, with GA of 19.55, 1.48, and 8.65, respectively. In Sindh, SL had a high h^2B (0.88) and GA (6.99), making spike length the most important trait. Low to moderate heritability for SL was noted [19, 24]. In Khyber Pakhtunkhwa, PH showed moderate h^2B (0.60) and GA (17.49), indicating important additive gene effects, followed by FLA (h^2B 0.54). Plant height and flag leaf area are the most heritable traits in KP. High h^2B and GA support selection for creating genotypes with desirable traits [19].

Cluster Analysis

To better study the observed variation in protein content in wheat varieties, cluster analysis of different provinces of Pakistan was performed using hierarchical clustering. The data regarding the presence or absence of protein bands was used to construct the dendrograms for wheat varieties from each of the three provinces of Pakistan to find the diversity among the given wheat varieties. The results of the cluster analysis for the Punjab, Sindh, and KP provinces are given in Fig. 2.

Heading

In Punjab, twenty-seven wheat varieties were analyzed and clustered into two main groups. Group G1 included only AS-02, highlighting its unique genetic profile. Group G2 was divided into three sub-clusters: Sub-cluster 1 (Mexipak-65, Millat-11, Faisalabad-08, AS-11, Chakwal-50, Margalla-99, Chenab-00), Sub-cluster 2 (Galaxy-13, Seher-06, Dharabi-11, Punjab-11, Pakistan-13), and Sub-cluster 3 (Ujala-15, Uqaab-00, NARC-09, Punjab-85, Ufaq-02, Bhaktawar-94, Shahkar-13, Zincol-16, NARC-11, Seher, Borloug-16, Wafaaq-01, Shalimar-88). Ujala-15, Uqaab-00, NARC-09, Punjab-85, and Ufaq-02 were closely grouped, indicating a narrow genetic pool, suggesting the need for genetic diversity. In Sindh, fifteen wheat varieties were clustered into three sub-divisions: Sub-division 1 (Bhattai), Sub-division 2 (six varieties), and Sub-division 3 (eight varieties). Kiran-95 and TD-1 were

Table 2. Mean and standard deviation in wheat varieties.

S.NO.	VARIETIES	DTH (days)	SD	DTF (days)	SD	FLA (cm ²)	SD	PH (cm)	SD	SL (cm)	SD	SPKLT (numbers)	SD	GN (numbers)	SD	TGW (g)	SD
1	Aman	76	0	77	3	30,73	0,45	54	3,5	8,5	0,5	16	0,6	44	2,89	45,7	2,2
2	Amber-2010	65	0,9	64	0	30,4	0,15	47	4,8	9,3	0,38	13	0,2	45	2,48	46	2,45
3	Atta Habib	55	1,1	57	2,5	22,1	0,9	44	2,5	8,2	0,87	21	0,9	43	1,01	37	3,45
4	Barsaat	57	3	65	0,6	33,4	1,39	53	1,54	13	2,1	23	2,7	42	0,42	45	2,08
5	Bathoor	78	2,45	87	0,08	25,7	0	44	3,24	9,3	0,25	20	0,9	43	0,99	44,3	3,08
6	Benazir-13	62	1,09	67	0,15	21,13	0,09	55	1,45	9,3	0,25	17	0,4	42	0,42	54	3,56
7	Bhittai	65	0,98	87	0,3	23,64	1,25	64	1,7	10	0,15	20	0,2	43	0,68	43,7	2,45
8	DN-84	77	0,45	83	3	22,37	2,56	53	0,34	11	1,07	15	0,2	32	0,79	42,3	2,1
9	Fakhr -e-Sarhad	69	0,67	73	4,2	23,44	0,4	54	2,46	11	1,57	19	0,7	44	0,42	56	3,4
10	Gomal-08	55	0,57	64	0,9	24,54	0,15	44	3,45	11	0,62	19	2,3	52	0,54	35	2,8
11	Hammal-13	67	1,73	65	0,17	25,63	0,98	54	2,87	9,4	0,29	17	2,6	44	1,09	36,3	2,2
12	Imdad-05	69	1,7	76	1,25	27,67	65	53	1,34	5,8	0,23	18	0,4	49	2,07	52,3	2,1
13	Insaaf	67	0,65	75	1,56	30,13	0,43	65	2,09	12	0,9	17	0,8	43	5,24	46	2,9
14	Kiran-95	75	0,78	84	2,4	21,41	1,24	56	2,54	9,2	0,67	20	2,3	45	0,47	44,7	3,4
15	Lalma	74	0,12	76	0,8	23	2,4	53	0,45	9,7	0,5	17	0,3	42	0,9	44,7	4,08
16	LU-26	67	0,71	71	0,74	23,37	3,2	72	0,65	10	0,34	18	0,5	44	2,34	41	3,24
17	Margalla-99	57	2,06	85	0,2	30,23	0,87	46	0,67	9,4	0,15	22	2,4	45	0,98	45,7	1,45
18	Pakhtunkhwa	59	0,56	66	0,45	23,73	0,5	55	0,13	11	0,56	14	0,8	45	0,9	36,3	2,43
19	Pirsabak-05	55	4,04	71	0,6	25,07	0,74	73	0,2	10	0,25	17	0,9	33	1,01	52,3	2,52
20	Sarang-13	54	0,45	58	0,3	24,33	1,24	55	2,59	8,6	0,97	16	0,3	44	1,24	40,7	4,62
21	Sassui	67	0,8	73	0,9	22,94	2,5	55	2,09	12	0,65	17	0,6	43	1,05	42,7	3,08
22	Scher-06	65	0,46	68	1,15	20,37	0,56	54	3,5	10	0,15	14	0,9	38	2,24	45,7	6
23	Siren	75	0,98	78	2	22,97	0,009	55	2,58	12	0,1	20	0,2	54	0,42	36,3	2,3
24	SKD-1	74	1,8	76	2,9	23,7	0,45	53	1,9	9,3	1,07	14	0,2	34	0,87	43	2,46
25	Sundar-10	57	4,2	65	3,5	22,6	0,35	58	1,67	8,6	0,98	15	0,3	44	0,54	35	1,34
26	Sunheri-10	62	4,8	74	3,8	22,4	0,98	50	1,59	12	0,57	16	2,6	44	0,64	34,3	1,67

27	Tatara	65	0,86	67	4	23,68	0,45	54	2,89	13	0,21	19	0,9	41	4,3	52,7	1,89
28	TD-1	67	1,5	75	0,09	18	1,15	35	2,57	10	0,9	12	0,9	34	5,2	35	2,1
29	TJ-83	66	2,1	73	2,63	24,33	1,39	54	3,45	9,4	0,6	16	0,4	33	2,49	46	3,5
30	Zamindar-04	66	0	67	0	20,7	0,59	45	2,3	11	0,4	14	0,3	44	3,24	36,3	2,5
31	AS-02	46	1,73	54	1,15	24,13	0,42	61	2,04	8,3	0,35	14	0,3	36	3,06	42,7	2,31
32	AS-11	68	3,51	72	0,58	43,9	0,66	76	2,02	11	0,29	18	0,6	37	4,16	43,7	4,93
33	Bhaktawar-94	62	2,06	75	3,06	47,87	0,45	67	2,25	8,5	1,95	15	2,6	32	2,83	43,7	3,04
34	Borloug-16	68	1,73	75	1,15	30,83	1,01	87	1,42	8,9	0,42	16	0,2	31	2,34	22,7	1,65
35	Chakwal-50	78	4,16	83	2,65	38	1,39	78	3,52	9,9	0,51	19	0,8	42	5,18	35,3	1,53
36	Chenab-00	59	0,58	74	4	39,23	0,95	85	0,89	7,7	0,72	15	3,8	37	0,5	32,3	0,55
37	Dera-98	64	0	73	1,15	21,97	0,5	66	1,98	9	0,35	14	0,7	44	0,47	53	2,67
38	Dharabi-11	75	0	79	0,58	37,57	1,43	86	1,12	7,8	0,15	13	0,5	29	0,64	44,7	2,96
39	Faisalabad-08	63	3,00	71	7,51	33,17	2,71	77	3	9,4	0,49	17	0,9	37	2,05	40,3	3,06
40	Galaxy-13	66	3,06	84	1,53	37	3	82	2,53	9,7	0,32	15	0,6	43	3,61	52,3	8,74
41	KPK-15	66	3,21	71	0,54	26,8	0,88	84	4,36	11	0,5	17	1,2	47	0,38	45	2,13
42	Marvi-00	54	0	66	1,62	23,1	2,5	74	4,02	10	0,26	18	0,4	45	2,09	67	3,2
43	Moomal-02	54	0,02	74	1,45	19,3	1,15	75	2,09	8,8	0,8	16	2,1	55	0,9	33,7	2,2
44	Mexipak-65	67	1,15	59	0	29,43	3,14	71	1,18	9,1	1,05	14	0,4	42	5,6	29,3	3,51
45	Millat-11	90	1,73	79	0,58	27,57	0,96	56	2,34	8,5	0,45	13	1,2	31	2,12	36,3	3,61
46	NARC-09	67	0,58	112	8,23	35,5	2,09	89	1,28	10	0,06	18	0,1	43	2	41,3	4,04
47	NARC-11	90	0	98	3,21	43,07	0,38	86	2,88	10	0,35	22	1,3	49	6,56	32	4,62
48	Pakistan-13	70	1,53	79	1	35,3	3,35	96	1,79	10	0,4	21	1,5	47	1,4	43,3	3,61
49	Pirsabak-08	71	2,06	77	0,58	16,67	1,88	60	1,12	6,8	0,15	12	0,3	26	2,05	22,7	3,51
50	Pirsabak-13	72	0,58	86	0,59	42,5	1,51	65	1,36	9,2	0,21	17	0,7	37	3,56	44	1,73
51	Punjab-11	59	5,77	99	3,79	31,43	0,29	76	0,36	8,3	0,7	15	0,8	32	2,19	46,7	5,69
52	Punjab-85	52	4,04	69	0,58	17,37	1,26	60	0,4	8	0,93	17	1,8	34	0,89	37	3,06
53	Shahkar-13	69	3	73	2,52	43,33	3,9	79	1,05	9,2	0,36	18	0,8	37	2,09	31,3	5,86
54	Shalimar-88	51	2,52	52	3,51	22	5,7	62	3,66	7,8	0,96	12	1,7	27	4,02	29,3	4,18

55	Seher-06	53	1,15	59	2,31	35,73	4,33	73	3,01	8,6	0,15	15	0,4	38	0,42	44,7	5,74
56	Ufaaq-02	52	1,15	68	0,58	19,63	3,87	59	4,43	7,7	0,7	11	0,9	31	2,76	33,7	5,13
57	Ujala-15	68	1,53	79	1,15	33,17	3,01	83	2,23	8,6	0,15	17	0,7	43	2,53	42	2,31
58	Uqaab-00	84	0	91	0	55,03	3,3	82	5,24	11	0,64	21	0,2	46	3,4	27,3	4,05
59	Wafaaq-01	67	1	71	0,58	34,03	1,59	83	1,56	9,4	0,25	17	0,9	49	5,71	34	4,36
60	Zincol-16	75	1,53	79	0,58	34,9	2,12	85	2,27	9,4	0,21	17	0,1	46	0,42	36	3,21

most similar, while Bhittai was unique. Kiran-95, TD-1, TJ-83, and Benazir-13 showed the lowest genetic dissimilarity, useful for hybridization programs. In Khyber Pakhtunkhwa, twenty-four wheat varieties were clustered into two main groups. Group G1 included Dera-98, DN-111, DN-102, DN-104, DN-118, DN-93, and DN-84. Group G2 was divided into three sub-clusters: Sub-cluster 1 (Fakhr-e-Sarhad, Zamindar-04, Atta Habib, Siren), Sub-cluster 2 (Pirsabak-05, Pirsabak-08), and Sub-cluster 3 (twelve varieties). Dera-98 and DN-102, as well as Barsaat and Tatara, were more similar. Sub-cluster 2 of G2 and G1 showed the least similarity. The moderate to low genetic diversity suggests the need for a detailed analysis of storage proteins to improve wheat quality [43].

Protein Profiling in Wheat Genotypes

Different varieties of wheat showed variable banding patterns. Electrophoregrams for Punjab, Sindh, and KP are given in Fig. 1b), c), and d), respectively. The grain storage protein patterns were determined by examining the molecular weight of the bands, with a total of 63 bands obtained. In Punjab, the most frequent HMW protein band was 104 kDa, and the most frequent LMW band was 18 kDa (Supplementary Table S1). Seher-06 had the maximum bands (17). In Sindh, the most frequent HMW band was 104 kDa, and the most frequent LMW band was 17 kDa (Supplementary Table S1). Sunehri had the maximum bands (17). In KP, the most frequent HMW bands were 66 kDa and 79 kDa, and the most frequent LMW bands were 36 kDa and 23 kDa. DN-93, DN-102, and DN-111 had the maximum bands (15). Higher polymorphism was observed with HMW-GS proteins. The diversity among wheat varieties results from various allele combinations [44]. Specific alleles of HMW-GS determine breadmaking quality, and certain HMW-GS genes can enhance this quality [45]. SDS-PAGE analysis showed maximum diversity for HMW-GS compared to LMW-GS [46]. This method is reliable, simple, and quick, widely used to estimate genetic diversity in crops like wheat [47]. HMW-GS plays a crucial role in defining bread quality [48]. A strong positive association exists between the HMW-GS fraction and breadmaking parameters [49]. Seed protein patterns obtained by electrophoresis are used to evaluate genetic diversity in wheat due to their simplicity and effectiveness [50]. Wheat protein analysis helps identify genetic diversity and maximize variations in germplasm collections, breeding wheat genotypes with improved breadmaking qualities [51]. Flours with higher glutenin content yield more stable dough, function better in breadmaking, increase dough during fermentation, and increase loaf capacity [52]. Higher HMW-GS/LMW-GS ratio wheat genotypes are associated with better rheological and breadmaking properties [27]. Future research should delve deeper into the genetic basis of gluten protein diversity and its correlation with wheat quality traits. Utilizing advanced genomic tools

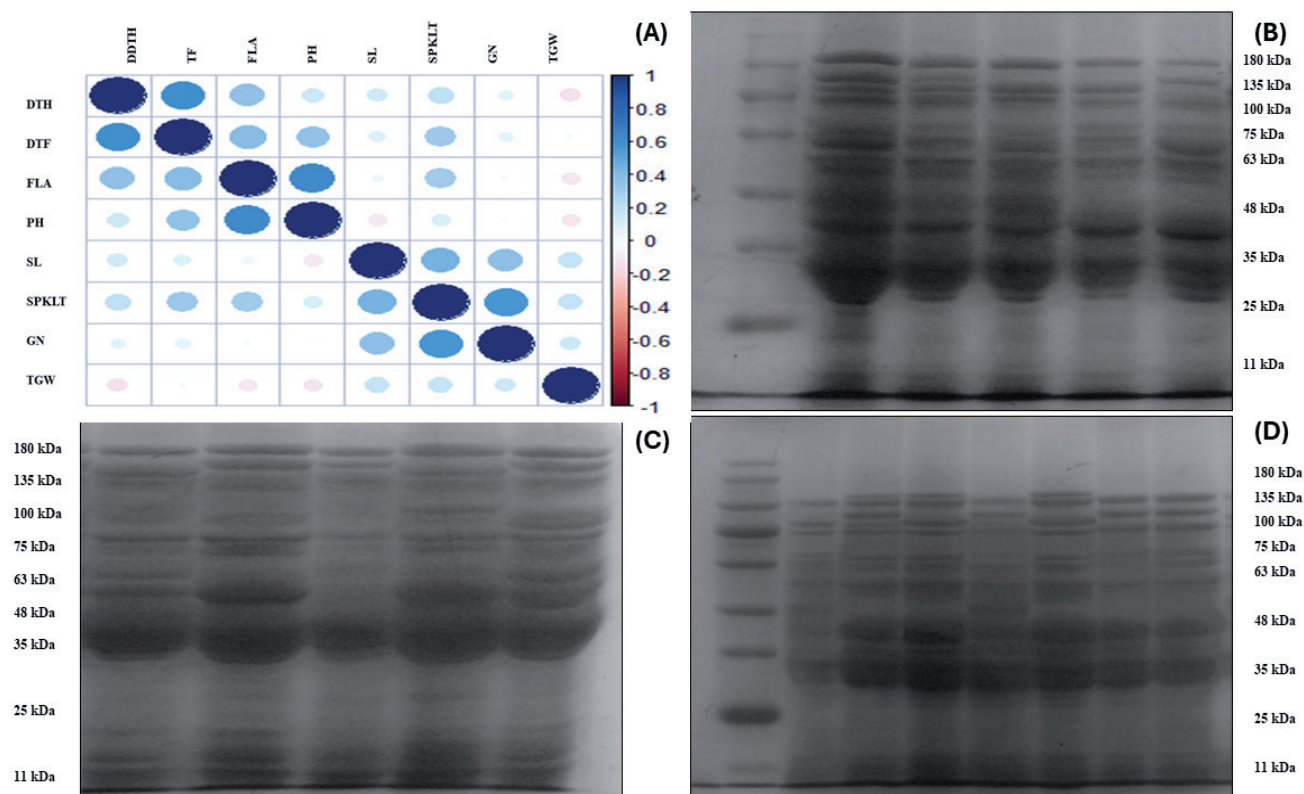


Fig. 1. a) correlation in all varieties, b) SDS-PAGE profile of some Punjab wheat, c) SDS-PAGE profile of some Sindh wheat Varieties, d) SDS-PAGE profile of some Khyber Pakhtunkhwa wheat varieties.

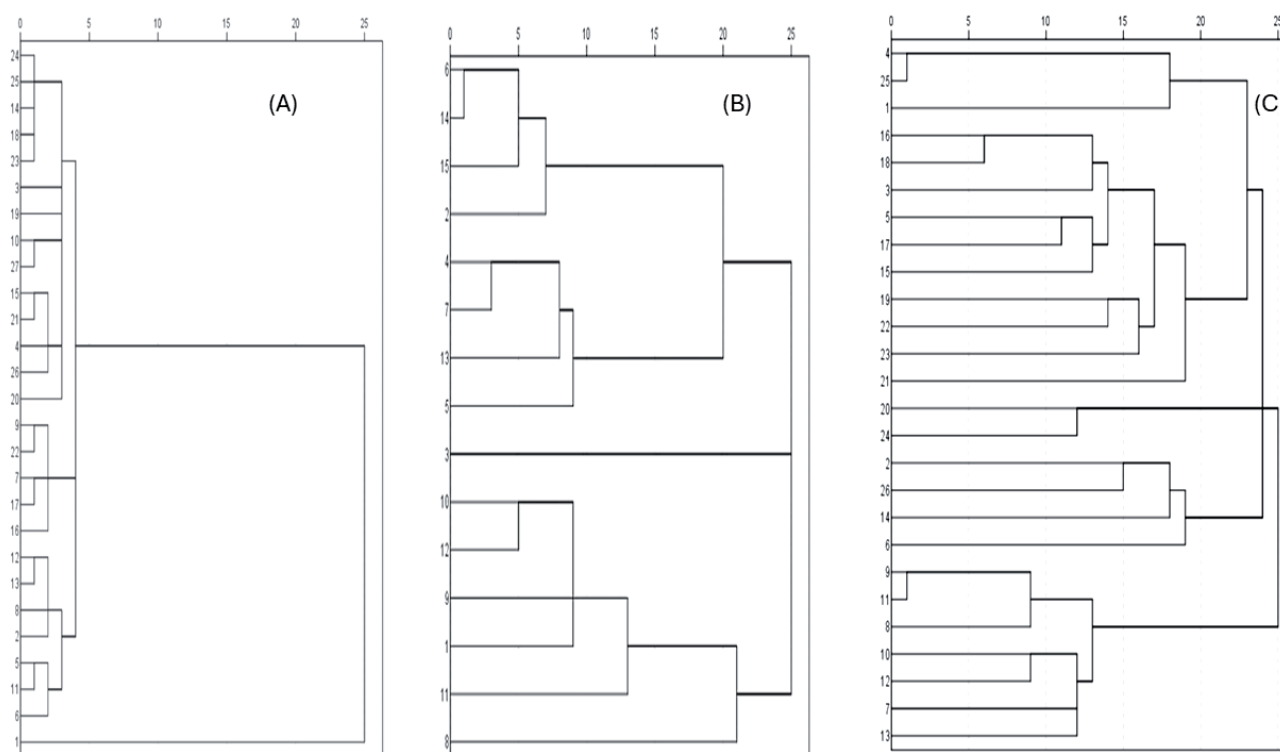


Fig. 2. Cluster analysis of wheat varieties (A) Punjab, (B) Sindh, (C) Khyber Pakhtunkhwa.

Table 3. Estimates of broad sense heritability, genetic advance, genotypic and phenotypic coefficients of variation for some quantitative traits in wheat in the Punjab, Sindh and Khyber Pakhtunkhwa.

Characters	h^2_B	GA	GCV	PCV	h^2_B	GA	GCV	PCV	h^2_B	GA	GCV	PCV
Days to heading	0,67	17,68	169,5	253	0,29	6,26	41	169,2	0,25	7,29	78,2	309,1
Days to flowering	0,77	24,52	24,5	323	0,21	6,7	70,9	324,9	0,18	6,13	67,4	368,9
Flag leaf area (cm ²)	0,81	25,45	676,2	831	0,05	1,09	22,7	402,1	0,54	17,5	484,1	888,1
Plant height (cm)	0,64	26,45	222	344	0,36	11,35	134	366,2	0,6	17,5	201,6	311,3
Spike length (cm)	0,38	27,45	15,5	40	0,88	6,99	19,3	165,2	0,28	1,7	27,2	97,2
Number of spikelets spike ⁻¹	0,18	28,45	45,6	250	0,15	1,52	23,3	152,4	0,15	2,04	73,1	273,1
Number of grains spike ⁻¹	0,27	29,45	93,5	340	0,35	6,49	73,3	200,5	0,16	5,15	102,2	610,6
1000-grain weight (g)	0,3	30,45	148,4	490	0,22	7,8	163,6	740	0,18	5,83	111,9	618,5

to pinpoint specific genes linked to desirable traits and developing new wheat varieties with enhanced nutritional value and breadmaking qualities will be crucial. This research could also explore the potential impact of climate change on wheat quality and yield, paving the way for more resilient wheat genotypes.

Conclusions

There was a significant difference between wheat varieties for all evaluated traits. Barsaat, Uqaab-00, Moomal-02, and Marvi-00 excelled in various agronomic traits, indicating their potential for breeding programs to develop improved bread varieties. High heritability estimates and moderate genetic advance were observed for traits such as heading and flowering days, plant height, spike length, and flag leaf area, suggesting these should be prioritized in future breeding efforts. The quality of wheat protein is genetically determined, varying by variety. Recent research on seed storage proteins aims to improve crops' nutritional and processing properties. This study shows that wheat varieties from different regions of Pakistan have significant nutritional potential in terms of protein. Gluten protein profiles can serve as markers in genotyping, cataloging new varieties, evaluating pedigrees and genetic diversity studies, and enhancing the efficiency of wheat breeding programs. Future research should focus on advanced genomic selection techniques, developing climate-resilient wheat varieties, incorporating genetic markers for improved nutritional content, enhancing resistance to biotic and abiotic stresses, integrating sustainable agricultural practices, and utilizing precision agriculture technologies.

Acknowledgments

The authors extend their appreciation to the researchers supporting project number (RSP2024R190), King Saud University, Riyadh, Saudi Arabia.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

1. AHMED M., QAMAR M., WAQAR S., NAEEM A., JAVAID R., TANVEER S., HUSSAIN I. Estimation of genetic components, heterosis and combining ability of elite Pakistani wheat varieties for yield attributing traits and stripe rust response. *Vavilov Journal of Genetics and Breeding*. **27** (6), 609, **2023**.
2. BERES B.L., RAHMANI E., CLARKE J.M., GRASSINI P., POZNIAK C.J., GEDDES C.M., PORKER K.D., MAY W.E., RANSOM J.K. A systematic review of durum wheat: Enhancing production systems by exploring genotype, environment, and management (G× E× M) synergies. *Frontiers in Plant Science*. **11**, 568657, **2020**.
3. ISHAQUE W., SHELIA V., ANOTHAI J., ZAMAN M., HOOGENBOOM G. Determining optimum nitrogen management as a function of planting date for spring wheat (*Triticum aestivum* L.) under semi-arid conditions using a modeling approach. *Journal of Arid Environments*. **182**, 104256, **2020**.
4. KATAMADZE A., VERGARA-DÍAZ O., UBEREGUI E., YOLDI-ACHALANDABASO A., ARAUS J.L., VICENTE R. Evolution of wheat architecture, physiology, and metabolism during domestication and further cultivation: Lessons for crop improvement. *The Crop Journal*. **11** (4), 1080, **2023**.
5. SLAFER G.A., GARCÍA G.A., SERRAGO R.A., MIRALLES D.J. Physiological drivers of responses of grains per m² to environmental and genetic factors in wheat. *Field Crops Research*. **285**, 108593, **2022**.
6. GAO F., MA D., YIN G., RASHEED A., DONG Y., XIAO Y., XIA X., WU X., HE Z. Genetic progress in grain yield and physiological traits in Chinese wheat cultivars of southern yellow and Huai Valley since 1950. *Crop Science*. **57** (2), 760, **2017**.
7. AMINIZADEH S., POURSEYEDI S., MOHAMMADI-NEJAD G., MOHAYEJI M., FARAHBAKHSH H., SALARPOUR M., ABDOLSHAH R. Comprehensive morpho-physiological criteria for screening bread wheat (*Triticum aestivum* L.) genotypes under drought stress

- condition. Cereal Research Communications. **52** (1), 287, **2024**.
8. KHAN H., KRISHNAPPA G., KUMAR S., MISHRA C.N., KRISHNA H., DEVATE N.B., RATHAN N.D., PARKASH O., YADAV S.S., SRIVASTAVA P. Genome-wide association study for grain yield and component traits in bread wheat (*Triticum aestivum* L.). Frontiers in Genetics. **13**, 982589, **2022**.
 9. XIAO J., LIU B., YAO Y., GUO Z., JIA H., KONG L., ZHANG A., MA W., NI Z., XU S. Wheat genomic study for genetic improvement of traits in China. Science China Life Sciences. **65** (9), 1718, **2022**.
 10. LANGRIDGE P., REYNOLDS M. Breeding for drought and heat tolerance in wheat. Theoretical and Applied Genetics. **134**, 1753, **2021**.
 11. VAN FRANK G., RIVIÈRE P., PIN S., BALTASSAT R., BERTHELLOT J.-F., CAIZERGUES F., DALMASSO C., GASCUEL J.-S., HYACINTHE A., MERCIER F. Genetic diversity and stability of performance of wheat population varieties developed by participatory breeding. Sustainability. **12** (1), 384, **2020**.
 12. SABOURI H., ALEGH S.M., SAHRANAVARD N., SANCHOULI S. SSR linkage maps and identification of QTL controlling morpho-phenological traits in two Iranian wheat RIL populations. BioTech. **11** (3), 32, **2022**.
 13. GHAFFAR Y., ASHRAF W., AKHTAR N., ZESHAN M.A., GHANI M.U., FATIMA S., ANSARI M.J., ALFARRAJ S., MAQBOOL A. Estimation of statistical parameters in candidate wheat genotypes for yield-related traits. Journal of King Saud University-Science. **34** (8), 102364, **2022**.
 14. OUMATA S., MONNEVEUX P., ZAHARIEVA M., MEKLICHE-HANIFI L., DAVID J. Variation of morphological traits among wheat (*Triticum aestivum* L.) landraces from two regions of the algerian sahara. Potential interest for wheat breeding. Genetic Resources and Crop Evolution. **70** (1), 235, **2023**.
 15. THAPA R.S., SHARMA P.K., PRATAP D., SINGH T., KUMAR A. Assessment of genetic variability, heritability and genetic advance in wheat (*Triticum aestivum* L.) genotypes under normal and heat stress environment. Indian Journal of Agricultural Research. **53** (1), 51, **2019**.
 16. BAYE A., BERIHUN B., BANTAYEHU M., DEREBE B. Genotypic and phenotypic correlation and path coefficient analysis for yield and yield-related traits in advanced bread wheat (*Triticum aestivum* L.) lines. Cogent Food & Agriculture. **6** (1), 1752603, **2020**.
 17. EL-ESAWI M.A., ELASHTOKHY M.M., SHAMSELDIN S.A., EL-BALLAT E.M., ZAYED E.M., HEIKAL Y.M. Analysis of genetic diversity and phylogenetic relationships of wheat (*Triticum aestivum* L.) genotypes using phenological, molecular and DNA barcoding markers. Genes. **14** (1), 34, **2022**.
 18. MAHMOUD S.A., EL-SHARKAWY E.S., EMAM M. Breeding sesame for resistance to charcoal rot caused by *Macrophomina phaseolina*. SVU-International Journal of Agricultural Sciences. **6** (2), 18, **2024**.
 19. AHMED H.G.M.-D., FATIMA N., FAISAL A., ULLAH A., ALI M., AMEEN M., IRFAN M., IMRAN M. Characterization of Bread Wheat Genotypes Using Spike Related Traits for Sustainable Yield Potential. Journal of Applied Research in Plant Sciences. **4** (1), 469, **2023**.
 20. KHAN M.J., ALI M.F., SHAHWANI S.A., WAHEED A., AZIZ I. Exploring the Role of Foliar Application of Aspirin on Growth and Productivity of Wheat under Control and Rainfed Conditions. Journal of Plant and Environment. **3** (1), 01, **2021**.
 21. BUKHARI M.A., SHAH A.N., FAHAD S., IQBAL J., NAWAZ F., MANAN A., BALOCH M.S. Screening of wheat (*Triticum aestivum* L.) genotypes for drought tolerance using polyethylene glycol. Arabian Journal of Geosciences. **14** (24), 2808, **2021**.
 22. ANWAR T., MUNWWAR F., QURESHI H., SIDDIQI E.H., HANIF A., ANWAAR S., GUL S., WAHEED A., ALWAHIBI M.S., KAMAL A. Synergistic effect of biochar-based compounds from vegetable wastes and gibberellic acid on wheat growth under salinity stress. Scientific Reports. **13** (1), 19024, **2023**.
 23. ANWAR T., SHEHZADI A., QURESHI H., SHAH M.N., DANISH S., SALMEN S.H., ANSARI M.J. Alleviation of cadmium and drought stress in wheat by improving growth and chlorophyll contents amended with GA3 enriched deashed biochar. Scientific Reports. **13** (1), 18503, **2023**.
 24. TCHERKEZ G., BEN MARIEM S., LARRAYA L., GARCÍA-MINA J.M., ZAMARREÑO A.M., PARADELA A., CUI J., BADECK F.-W., MEZA D., RIZZA F. Elevated CO₂ has concurrent effects on leaf and grain metabolism but minimal effects on yield in wheat. Journal of Experimental Botany. **71** (19), 5990, **2020**.
 25. DE SOUSA T., RIBEIRO M., SABENÇA C., IGREJAS G. The 10,000-year success story of wheat! Foods. **10** (9), 2124, **2021**.
 26. XUE C., MATROS A., MOCK H.-P., MÜHLING K.-H. Protein composition and baking quality of wheat flour as affected by split nitrogen application. Frontiers in plant science. **10**, 452714, **2019**.
 27. SIDDIQI R.A., SINGH T.P., RANI M., SOGI D.S., BHAT M.A. Diversity in grain, flour, amino acid composition, protein profiling, and proportion of total flour proteins of different wheat cultivars of North India. Frontiers in Nutrition. **7**, 141, **2020**.
 28. ZHANG H., TANG S., WANG H., WANG Y., ZHI H., LIU B., ZHANG R., MA Q., JIA G., FENG B. Genetic diversity of grain yield traits and identification of a grain weight gene *StGW6* in foxtail millet. Theoretical and Applied Genetics. **137** (4), 84, **2024**.
 29. WIESER H., KOEHLER P., SCHERF K.A. Chemistry of wheat gluten proteins: Qualitative composition. Cereal Chemistry. **100** (1), 23, **2023**.
 30. NAGY A.E.-F., ABD-ELSAYED S., MAHGOUB E., KOMBER R. Genetical studies on wheat drought tolerance using molecular and biochemical markers. Zagazig Journal of Agricultural Research. **43** (6), 2411, **2016**.
 31. WIESER H., KOEHLER P., SCHERF K.A. Chemistry of wheat gluten proteins: Quantitative composition. Cereal Chemistry. **100** (1), 36, **2023**.
 32. GEISLITZ S., PRONIN D., NEERUKONDA M., CURELLA V., NEUFANG S., KOCH S., WEICHERT H., WEBER H., BÖRNER A., SCHUPPAN D. Breeding from 1891 to 2010 did not increase the content of amylase/trypsin-inhibitors in wheat (*Triticum aestivum*). npj Science of Food. **7** (1), 43, **2023**.
 33. PRONIN D., GEISLITZ S., BÖRNER A., SCHERF K.A. Fingerprinting of wheat protein profiles for improved distinction between wheat cultivars and species. Cereal Chemistry. **97** (5), 999, **2020**.
 34. WANG Q., SUN G., REN X., DU B., CHENG Y., WANG Y., LI C., SUN D. Dissecting the genetic basis of grain size and weight in barley (*Hordeum vulgare* L.) by QTL and comparative genetic analyses. Frontiers in plant science. **10**, 469, **2019**.

35. SINGH S.P., CHAND P., SINGH V., SINGH A., TIWARI A., SINGH A., KUMAR M. Morphological Characterization and Assessment of Genetic Variability, Heritability and Genetic Advance in Bread Wheat (*Triticum aestivum* L.). *Plant Cell Biotechnology and Molecular Biology*. **25** (1-2), 120, **2024**.
36. GORDEEVA E., SHAMANIN V., SHOEVA O., KUKOEVA T., MORGOUNOV A., KHLESTKINA E. The strategy for marker-assisted breeding of anthocyanin-rich spring bread wheat (*Triticum aestivum* L.) cultivars in Western Siberia. *Agronomy*. **10** (10), 1603, **2020**.
37. MAHARAJAN T., ROCH G.V., CEASAR S.A. Recent advancements of molecular breeding and functional genomics for improving nitrogen-phosphorus and potassium-use efficiencies in wheat. In: *Molecular breeding in wheat, maize and sorghum: Strategies for improving abiotic stress tolerance and yield*, pp. 170, CABI Digital Library. **2021**.
38. JONES B.H., BLAKE N.K., HEO H.-Y., MARTIN J.M., TORRION J.A., TALBERT L.E. Allelic response of yield component traits to resource availability in spring wheat. *Theoretical and Applied Genetics*. **134**, 603, **2021**.
39. KUMAR A., MANTOVANI E.E., SIMSEK S., JAIN S., ELIAS E.M., MERGOUM M. Genome wide genetic dissection of wheat quality and yield related traits and their relationship with grain shape and size traits in an elite× non-adapted bread wheat cross. *Plos One*. **14** (9), e0221826, **2019**.
40. KUMAR D., KUMAR A., CHHOKAR V., GANGWAR O.P., BHARDWAJ S.C., SIVASAMY M., PRASAD S.S., PRAKASHA T., KHAN H., SINGH R. Genome-wide association studies in diverse spring wheat panel for stripe, stem, and leaf rust resistance. *Frontiers in Plant Science*. **11**, 748, **2020**.
41. CHAUDHARY S., DEVI P., BHARDWAJ A., JHA U.C., SHARMA K.D., PRASAD P.V., SIDDIQUE K.H., BINDUMADHAVA H., KUMAR S., NAYYAR H. Identification and characterization of contrasting genotypes/cultivars for developing heat tolerance in agricultural crops: Current status and prospects. *Frontiers in Plant Science*. **11**, 587264, **2020**.
42. FISCHER R., REBETZKE G. Indirect selection for potential yield in early-generation, spaced plantings of wheat and other small-grain cereals: a review. *Crop and Pasture Science*. **69** (5), 439, **2018**.
43. AMIRI R., SASANI S., JALALI-HONARMAND S., RASAEI A., SEIFOLAHPOUR B., BAHRAMINEJAD S. Genetic diversity of bread wheat genotypes in Iran for some nutritional value and baking quality traits. *Physiology and Molecular Biology of Plants*. **24**, 147, **2018**.
44. WANG X., SONG R., AN Y., PEI H., GAO S., SUN D., REN X. Allelic variation and genetic diversity of HMW glutenin subunits in Chinese wheat (*Triticum aestivum* L.) landraces and commercial cultivars. *Breeding Science*. **72** (2), 169, **2022**.
45. GIANCASPRO A., GIOVE S.L., ZACHEO S.A., BLANCO A., GADALETA A. Genetic variation for protein content and yield-related traits in a durum population derived from an inter-specific cross between hexaploid and tetraploid wheat cultivars. *Frontiers in Plant Science*. **10**, 1509, **2019**.
46. LI W., WAN Y., LIU Z., LIU K., LIU X., LI B., LI Z., ZHANG X., DONG Y., WANG D. Molecular characterization of HMW glutenin subunit allele 1Bx14: further insights into the evolution of Glu-B1-1 alleles in wheat and related species. *Theoretical and Applied Genetics*. **109**, 1093, **2004**.
47. DAI Y., LI J., SHI J., GAO Y., MA H., WANG Y., MA H. Molecular Characterization and Marker Development of the HMW-GS Gene from *Thinopyrum elongatum* for Improving Wheat Quality. *International Journal of Molecular Sciences*. **24** (13), 11072, **2023**.
48. ZHAO L., LI L., SONG L., LIU Z., LI X., LI X. HMW-GS at Glu-B1 locus affects gluten quality possibly regulated by the expression of nitrogen metabolism enzymes and glutenin-related genes in wheat. *Journal of Agricultural and Food Chemistry*. **68** (19), 5426, **2020**.
49. YANG T., WANG P., WANG F., ZHOU Q., WANG X., CAI J., HUANG M., JIANG D. Influence of starch physicochemical properties on biscuit-making quality of wheat lines with high-molecular-weight glutenin subunit (HMW-GS) absence. *LWT- Food Science and Technology*. **158**, 113166, **2022**.
50. CHEN X., YIN G., BÖRNER A., XIN X., HE J., NAGEL M., LIU X., LU X. Comparative physiology and proteomics of two wheat genotypes differing in seed storage tolerance. *Plant Physiology and Biochemistry*. **130**, 455, **2018**.
51. MICHEL S., LÖSCHENBERGER F., AMETZ C., PACHLER B., SPARRY E., BÜRSTMAYR H. Combining grain yield, protein content and protein quality by multi-trait genomic selection in bread wheat. *Theoretical and Applied Genetics*. **132**, 2767, **2019**.
52. PARENTI O., GUERRINI L., CANUTI V., ANGELONI G., MASELLA P., ZANONI B. The effect of the addition of gelatinized flour on dough rheology and quality of bread made from brown wheat flour. *LWT- Food Science and Technology*. **106**, 240, **2019**.

Supplementary Materials

Supplementary materials can be found at the link <https://www.pjoes.com/SuppFile/201946/1/>