Original Research

# Studies of the Biochemistry Behavior of Herbal Butterfly Pea Blue Pea Flower with some Antimicrobial and Antioxidant Potentials

Huda Aljumayi<sup>1</sup>, Rokayya Sami<sup>1\*</sup>, Helal F. Al-Harthi<sup>2</sup>, Ruqaiah I. Bedaiwi<sup>3</sup>, Naseh A. Algehainy<sup>3</sup>, Awatif M. Almehmadi<sup>4</sup>, Suzan A. Abushal<sup>5</sup>, Roqayah H. Kadi<sup>6</sup>, Tasahil S. Albishi<sup>7</sup>, Alaa T. Qumsani<sup>8</sup>, Sameer H. Qari<sup>8</sup>

<sup>1</sup>Department of Food Science and Nutrition, College of Sciences, Taif University,
P.O. Box 11099, Taif 21944, Saudi Arabia

<sup>2</sup>Department of Biology, Turabah University College, Taif University, 21995, Saudi Arabia

<sup>3</sup>Department of Medical Laboratory Technology, Faculty of Applied Medical Sciences, University of Tabuk,
P.O. Box 741, Tabuk 71491, Saudi Arabia

<sup>4</sup>Department of Clinical Nutrition, Faculty of Applied Medical Sciences, Umm Al-Qura University,

P.O. Box 715, Makkah 24382, Makkah 24382, Saudi Arabia

<sup>5</sup>Program of Food Sciences and Nutrition, Turabah University College, Taif University,
P.O. 11099, Taif 21944, Saudi Arabia

<sup>6</sup>Department of Biological Sciences, College of Science, University of Jeddah, Jeddah 21959, Saudi Arabia

<sup>7</sup>Biology Department, College of Sciences, Umm Al-Qura University, Makkah, Saudi Arabia

<sup>8</sup>Department of Biology, Aljumum University College, Umm Al-Qura University, Makkah 21955, Saudi Arabia

Received: 17 August 2024 Accepted: 22 February 2025

#### **Abstract**

Butterfly pea flowers are commonly used in food manufacturing as coloring agents. The current research aimed to study some phytochemical, antimicrobial, and antioxidant potentials for their extracts. Several assays have been applied to prepare five extracts coded as ME, WE, EE, GE, and GWE for methanol, water, ethanol, glycerol, and glycerol/water, especially for phenolic determination. At the same time, ultrasound processing has been achieved for both fresh and dried flowers, especially for the anthocyanin determination. The antibacterial potential for *Staphylococcus aureus* reported a 1.2 mm zone of inhibition. *C. ternatea* flower extract reported 6.4 and 10.29°Brix for acidity and total soluble sugar, respectively. The *C. ternatea* flower extract had a transparent, dark, and more opaque blue color due to a higher *L\** value (29.52) than lower *a\** and *b\** values (2.5 and 40.32-), respectively. *C. ternatea* flower extract exhibited a high total antioxidant capacity (59.11%), while ORAC reported (109.22 mg TE/L). ABTS results were IC<sub>50</sub> = 35.27 μL/mL and 189.68 mg TE/L. The total phenolic contents presented various results on a blue color, GE had the deepest blue color and reported (18.76 mg GAEg¹), followed by EE (18.55 mg GAEg¹), GWE (18.53 mg GAEg¹), and ME

...

<sup>\*</sup>e-mail: rokayya.d@tu.edu.sa

(15.17 mg GAEg<sup>-1</sup>). WE had the lightest blue color and reported the lowest value (9.79 mg GAEg<sup>-1</sup>). Four extractions were prepared for the total anthocyanin contents, which showed the efficacy of the ultrasound process to get the highest anthocyanin content of 4.21 mg Cyanidin-3-glucoside/g on fresh petals. Butterfly pea flowers might be used safely in food, beverage, and cosmeceutical products.

**Keywords:** Clitoria ternatea, antibacterial agent, antioxidants, phenols, anthocyanin

#### Introduction

The butterfly blue pea flower is considered one of the medical plants with ethnic medicinal uses. Clitoria ternatea is widely grown within the tropical belt, while some species are grown in temperate areas as ornamentals, fodders, or even medicinal plants [1]. It contains anthocyanins alkaloid, tannin, resin, steroid, saponin, phenol, ternatins, and flavonol glycoside [2]. These variable components can be used efficiently in the pharmaceutical industries, cosmetics, and drugs due to their huge role in healthcare [3]. The flowers are recommended for the curing of dangerous bites such as snakes and scorpions, as well as for boosting memory, improving the intellect, inhibiting platelet aggregation, and controlling some neurological disorders [4]. It has excellent antioxidant, antidiabetic, anti-obesity, anti-inflammation, anticancer, antihyperlipidemic, and anti-asthma properties. Parimaladevi et al. [5] reported the presence of some anti-fungal proteins that were homologous to the defensin plants. Several techniques have been applied for the bioactive extractions of fresh or dried blue pea flowers, especially brewing and boiling them for daily use [6]. Those bioactive components can be influenced by those techniques, especially anthocyanin, which is affected by temperature, according to several studies [7, 8]. Several anthocyanin types have been detected in the blue butterfly flowers, such as ternatins A3, (C1-C5), (B2-B4), and (D2-D3) [9]. Antioxidants can slow down lipoxygenase, chelate metals, lipid peroxidation, singlet oxygen quenching, and free radical formation to defend the body from a lot of serious diseases [10]. Heydari et al. [11] reported the negative effect of the dried flower intake against the bioactive compounds and the antioxidant activities. Marpaung et al. [12] confirmed that the color indicators of the blue flower extracts, such as intensity, violet, and browning indexes, can be varied according to the extraction techniques. Parimaladevi et al. [5] reported that changes in antioxidant values can be influenced by various extract solvents; DPPH activity was the highest with the ethanolic extract of the flower. Jain et al. [9] noted that the methanolic extracts had higher values compared to acetone extracts for the free radical scavenging activities.

The current research was arranged on the various extracts and preparation assays on butterfly blue pea flower with evaluating the antibacterial potential against Staphylococcus aureus, some phytochemical properties

such as (acidity, total soluble sugar, color index), total antioxidant capacity, oxygen radical absorbance capacities (ORAC), ABTS scavenging activity with phenolic and anthocyanin contents.

## Experimental

#### Flower Materials

The pea flowers (*C. ternatea*) were collected from a private farm in Taif, KSA. The petal parts were divided from the sepals. The flowers were separated into three sets. The fresh ones were preserved in a refrigerator at 4°C. The dried ones were achieved by air drying at an ambient temperature of 25°C and then were preserved at -80°C for advanced analysis. In contrast, the ultrasound processing at 40°C for 60 min has been achieved for both fresh and dried flowers, especially for the anthocyanin determination.

## Preparation of Clitoria Ternatea Extracts

Several assays have been applied to prepare five extracts coded as ME, WE, EE, GE, and GWE for methanol, water, ethanol, glycerol, and glycerol/water, especially for phenolic determination. Solvents were prepared according to the previous study by Sami et al. [13]. Fig. 1. shows the experimental work processes of *Clitoria ternatea* extracts.

#### Microbial Determination

Staphylococcus aureus is well-known as a ubiquitous communal bacterium on the skin and anterior nare, while it can cause infection in the case of the bacterial generation increase [14]. The antimicrobial potential determination was detected for *C. ternatea* flower against Gram-positive *Staphylococcus aureus* as a food-borne bacterial on blood agar, then Tryptone Soya, which was purchased from the cell bank, Jeddah, Saudi Arabia. *Staphylococcus aureus* bacteria was suspended and diluted, then incubated at 37°C for 24 h. The result for the antimicrobial potential determination of *Staphylococcus aureus* bacteria was expressed as the zone of inhibition in (mm).

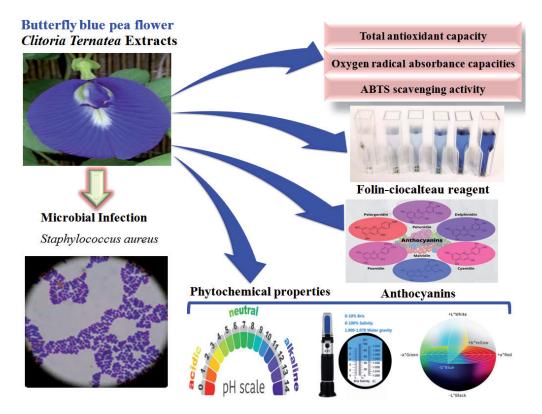


Fig.1. The summary of the experimental work on Clitoria ternatea extracts.

# Determination of some Phytochemical Properties

## **Acidity Determination**

pH is the quantity of acidity in liquids by using a digital pH meter (LPV2500.97.0002, PH1, Spain) at the ambient temperature, which equals the activity Log (-) of the (H) ions. pH determination may help for prolonging the shelf-life of liquids and it also can control the customer's acceptability [15]. High acidic values may cause some harmful health effects, such as gastroesophageal reflux diseases and dental [16].

### **Total Soluble Sugar Determination**

Total soluble sugar is a percentage of the presence of sucrose in a sugary *C. ternatea* flower solution [17]. Determination of the total soluble sugar content is essential in beverages to evaluate customer acceptability. The total soluble sugar content can be detected via the gravimetric assay using a hand refractometer (Kyowa, HR-1, Japan) [18].

#### Color Index Determination

The color index is an essential factor for sensorial evaluation and can influence the selection of the beverage itself [19]. Furthermore, *C. ternatea* flower extracts are affected by acidity due to their anthocyanin contents [20]. The color ranged from purple-blue near

pH=7, then it turned to red at pH $\leq$ 7 pH, while it turned to dark red at pH $\leq$ 3 and turned to green at pH $\geq$ 10.5 [21]. The color index profile ( $L^*$ ,  $a^*$ , and  $b^*$ ) was detected by a Miniscan XE Plus spectrophotometer. The final average was calculated when (n = 3) for various values [22].

## Determination of some Antioxidant Potentials

## **Total Antioxidant Capacity Determination**

The antioxidant capacity of *C. ternatea* flower was detected by the phosphomolybdate assay by using the ascorbic acid as a blank [23]. It was determined with the help of a reagent solution (0.6 M H<sub>2</sub>SO<sub>4</sub>, 28 mM Na<sub>3</sub>PO<sub>4</sub>, and four mM (NH<sub>4</sub>)6Mo<sub>7</sub>O<sub>24</sub>). The results were detected using a spectrophotometer (Helios Zeta, UV-VIS, Thermo Fisher, USA) and were expressed in percentage after absorbance at 695 nm wavelength [24].

# Determination of Oxygen Radical Absorbance Capacity

The ORAC was detected according to [25] assay. Approximately 75 mM Phosphate-buffered saline with (pH = 7.4) was mixed with 40 mg/mL 2, 2'-azobis (2-amidino-propane) dihydrochloride, 4.8  $\mu$ M fluorescein, and 1.5 and 0.75  $\mu$ g/mL Trolox standard solutions. In comparison, *C. ternatea* flower was dissolved in 2% dimethyl sulfoxide. The mixture

was incubated for 10 mins at 37°C. The result was absorbed at 494 nm and 535 wavelengths. Results were expressed as mg TE/L.

# Determination of ABTS Scavenging Activity

The ABTS scavenging activity was detected by using diammonium salt ABTS+ 2, 2-Azinobis-(3 ethylbenzothiazoline-6-sulfonate) [26]. Approximately 2  $\mu$ L of *C. ternatea* flower was mixed with 198  $\mu$ L of ABTS reagent against 200  $\mu$ L of Trolox solution [27]. The mixtures were incubated for 10 mins at 37°C. The mixture was absorbed at 745 nm wavelength. The lightening of the colors indicates the ABTS scavenging activity effects. The results were detected using a spectrophotometer and expressed as IC<sub>50</sub> ( $\mu$ L/mL) and mg TE/L [28].

#### **Determination of Total Phenolic Contents**

The total phenolic contents (TPC) of *C. ternatea* flower (5 mL) extracts as (ME), (WE), (EE), (GE), and (GWE) were detected according to the Folin-Ciocalteu assay by Mazzucotelli et al. [29]. Approximately 0.5 mL of the Folin-Ciocalteu reagent was mixed by vortex with 1.5 mL 7.5% Na<sub>2</sub>CO<sub>3</sub> and 7.9 ml H<sub>2</sub>O against 0.1 mL gallic acid as a standard. The mixtures were diluted to reach a concentration of 10 mg/mL and incubated for 1 hour in a dark area. The mixtures were absorbed using a spectrophotometer at a wavelength of 750 nm, while the results were expressed as gallic acid equivalents (mg GAEg<sup>1</sup>).

## Determination of Total Anthocyanin Contents

The total anthocyanin contents (TAC) were evaluated according to the previous pH-differential assay [30]. Approximately 1 mL of *C. ternatea* extracts was mixed in a beaker glass with KCl buffer (pH= 1.0) and CH<sub>3</sub>COONa buffer (pH= 4.5) to reach a volume of 10 mL. The absorbance using a spectrophotometer was detected at 520 nm and 700 nm wavelengths against using distilled water a blank. The results were expressed as mg Cyanidin-3-glucoside/g and calculated according to the protocol described by Ahmad et al. [31].

#### Statistical Analysis

The data from all replications of the current study with standard deviations have been reported as final results. The significant differences between all means were determined by Duncan's Multiple Range Test  $(p \le 0.05)$ .

#### **Results and Discussions**

#### **Antimicrobial Potential**

The result of the antimicrobial potential for *C. ternatea* extract is exposed in Table 1. The antimicrobial potential in the agar media for *Staphylococcus aureus* was detected in a zone of inhibition and reported a 1.2 mm zone of inhibition. The antimicrobial potential was linked with the previous study for several micros, such as *Staphylococcus aureus*, *Enterococcus faecalis*, and *Salmonella typhimurium* [32]. The result was also linked with Dacullo and Bitacura [33], who reported that the function inhibited the antimicrobial potential activity.

## Acidity and Total Soluble Solids

Clitoria ternatea is a simply accessible flower the acidity may be influenced by several indicators. As shown in Table 1, the acidity value reported was 6.4, which is linked with Rangel et al. [34], who contributed that the changes in color for the butterfly pea beverages can be influenced by the variation in pH values and high temperature. Based on the research work, C. ternatea flower extract reported 10.29°Brix. The results were in agreement with Featherstone [35], who detected the <sup>o</sup>Brix and acid ratios. Heating can cause changes in pH values in several herbal teas, such as acacia and squash blossom, and a reduction in the titration value in pumpkin flower tea. On the other hand, heating can reduce the acidity values in other herbal teas, such as honeysuckle, lilac, and clove [36]. The heating period for the butterfly flower can reduce the pH value and increase the titration value [37, 38]. Also, Adams [39] reported a deep purple color can be detected in sports drinks with lower acidity.

#### Color Index Profile

The color index is one of the essentially important signs of the customer's acceptance [40]. The C. ternatea flower extract had a transparent, dark, and more opaque blue color due to a higher  $L^*$  value (29.52) than lower  $a^*$  and  $b^*$  values (2.5 and 40.32-), respectively (Fig. 2.). Amaolo [41] used C. ternatea flower as a low-cost pH indicator. The flavylium cations and quinonoid anion can absorb various wavelengths of light, and hence color gradients are formed, especially with lower pH. The best method for using C. ternatea flower as a food additive for a long-term storage period is in a powdered form [42].

## Total Antioxidant Capacity and Oxygen Radical Absorbance Capacity

The total antioxidant capacity and ORAC of *C. ternatea* extract were detected by phosphomolybdate assay, and the results were expressed in percentage and

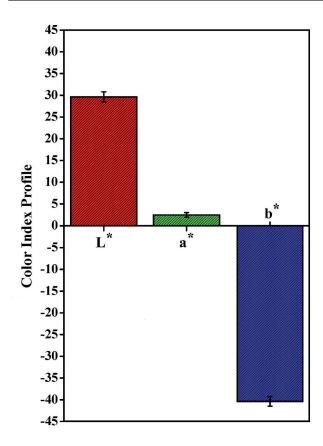


Fig. 2. Color index profile of blue pea flowers

mg TE/L of Trolox, respectively. *C. ternatea* flower extract exhibited a high total antioxidant capacity (59.11%), while ORAC reported (109.22 mg TE/L), Table 1. Rabeta and Nabil [43] reported that the high values of oxidation were due to the contributions of phenolics and oxygen radical absorbance capacities in the *C. ternatea* flower.

#### **ABTS Scavenging Activity**

ABTS scavenging activity may determine the free radical diammonium salt of ABTS+ 2, 2-Azinobisn [44]. The results were expressed as  $IC_{50}$  ( $\mu L/mL$ ) and mg

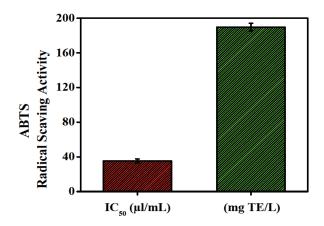


Fig. 3. ABTS radical scavenging activity of blue pea flowers.

TE/L in blue-green solutions with the phosphomolybdate assay. Fig. 3. shows ABTS results as  $IC_{50} = 35.27 \ \mu L/mL$  and 189.68 mg TE/L. Previous studies reported a relationship between heating temperature and the infusion times of the flower extract with the increase in ABTS scavenging activity [45, 46]. The previous factors, temperature and time, allow the bioactive components to become free due to tissue degradation.

## **Total Phenolic Contents**

The calibration curve of gallic acid represents the oxidation by phosphotungstic and phosphomolybdic acids [47]. TPC different solvents were evaluated, while the data were expressed in mg GAEg<sup>-1</sup>. The experiment presented various results on the blue color. GE had the deepest blue color and reported (18.76 mg GAEg<sup>-1</sup>), followed by EE (18.55 mg GAEg<sup>-1</sup>), GWE (18.53 mg GAEg<sup>-1</sup>), and ME (15.17 mg GAEg<sup>-1</sup>), Fig. 4. WE had the lightest blue color and reported the lowest value (9.79 mg GAEg<sup>-1</sup>). The glycerol extracts had a better ability to extract TPC than the ethanol, methanol, and water due to the polar structure of the phenols [48]. The TPC results were in range with the previous research by Widowati et al. [26], who reported

Table 1. Antioxidant activities, phytochemical analysis, and microbial infection of butterfly pea flowers.

Antioxidant Activities	
Total Antioxidant Capacity	59.11±2.48%
Oxygen Radical Absorbance Capacity	109.22±5.78 mg TE/L
Phytochemical Analysis	
Total Soluble Sugar	10.29±1.24 °Brix
pH	6.40±1.14
Antibacterial Potential	
Staphylococcus aureus	1.20±0.10 Zone of Inhibition (mm)

<sup>\*</sup>Each value presents as mean $\pm$ standard deviation. (n = 3)

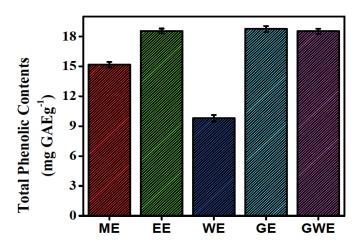


Fig. 4. Total phenolic contents of blue pea flowers.

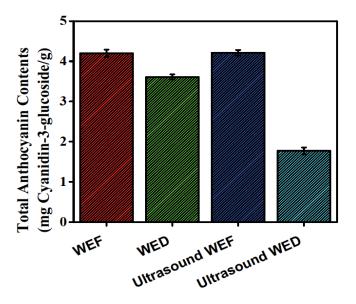


Fig. 5. Total anthocyanin contents of blue pea flowers.

16.20 µg GAE%. Cacace and Mazza [49] reported the migration of phenolic components into water during higher temperatures and infusion time in milled berries tea. The same increase in TPC was reported by (corn tassels, walnut shells, cherry stalks, banana, pomegranate, mandarin, eggplant, and red onion peel) teas [50, 51].

## **Total Anthocyanin Contents**

The anthocyanin content in butterfly peas is extremely high in a delphinidin group with a red-blue color and a high number of hydroxyls, which tend to have high antioxidant activities. Anthocyanins may prevent brain inflammation, blood clogging, and tumor cells, inhibit bacterial growth, control blood sugar, improve vision with analgesic, and exhibit antiasthma, hepatoprotective, anthelmintic, antiparasitic, antiulcer, and anticholesterol activities [52]. Four extractions

were prepared for TAC, which showed the efficacy of the ultrasound process to get the highest anthocyanin content as 4.21 mg Cyanidin-3-glucoside/g on fresh petal compared to 1.77 mg Cyanidin-3-glucoside/g for the dry one, Fig. 5. On the other hand, the commercial water extract reported similar value 4.20 mg Cyanidin-3-glucoside/g on fresh petal, while the dry one reported mg Cyanidin-3-glucoside/g. Therefore, both procedures were efficient in extracting the TAC in fresh condition, as anthocyanins are well-known as polar substances which are easily soluble in water droplets [53]. The current research was in agreement with Sofyan et al. [54], who applied some treatments for serving the butterfly pea flower drinks. Shen et al. [55] reported that anthocyanin is quite stable during the extraction at high temperatures under both neutral and alkaline conditions. Zor et al. [56] reported a reduction in the TAC at 80°C due to the degradation of pigment.

#### **Conclusions**

C. ternatea is generally used in food and beverage preparations. It gives an acceptable blue tint without artificial food coloring additives. It has antimicrobial activity against Staphylococcus aureus bacteria. Glycerol extract had the highest extraction effectiveness for the total phenolic contents. The ultrasound extraction method was the most effective, followed by water extract to get the highest anthocyanin content on fresh petals. It is essential to carry out more research about butterfly pea flowers to use as antioxidants and anti-inflammatory boosters in food and cosmeceutical products.

#### Acknowledgment

The authors extend their appreciation to Taif University, Saudi Arabia, for supporting this work through project number (TU-DSPP-2024-10).

#### **Funding**

This research was funded by Taif University, Saudi Arabia, Project No. (TU-DSPP-2024-10).

#### **Conflict of Interest**

The authors declare no conflict of interest.

#### References

- 1. JAIN N.N., OHAL C.C., SHROFF S.K., BHUTADA R.H., SOMANI R.S., KASTURE V.S., KASTURE S.B. *Clitoria ternatea* and the CNS. Pharmacology Biochemistry and Behavior, **75** (3), 529, **2003**.
- DEY ANKITA D.A., BHATTACHARYYA SAURYYA B.S., KUMAR P.T. Antioxidant activities of Moringa concanensis flowers (fresh and dried) grown in West Bengal. 2014.
- PANDA S., DEEP CHOUDHURY L., SOLANKI M. Formulation And Evaluation Of Antipigment Herbal Blue Pea Based Face Wash Cream Scrub. JETIR, 2020.
- KAZUMA K., NODA N., SUZUKI M. Malonylated flavonol glycosides from the petals of *Clitoria ternatea*. Phytochemistry, 62 (2), 229, 2003.
- DEVI B.P., BOOMINATHAN R., MANDAL S.C. Antiinflammatory, analgesic and antipyretic properties of *Clitoria ternatea* root. Fitoterapia, 74 (4), 345, 2003.
- ZOU Z., XI W., HU Y., NIE C., ZHOU Z. Antioxidant activity of Citrus fruits. Food Chemistry, 196, 885, 2016.
- MARPAUNG A.M., ANDARWULAN N., HARIYADI P., FARIDAH D.N. Thermal degradation of anthocyanins in butterfly pea (Clitoria ternatea L.) flower extract at pH 7. American Journal of Food Science and Technology, 5 (5), 199, 2017.
- 8. MARPAUNG A.M. Tinjauan manfaat bunga telang *(Clitoria ternatea L.)* bagi kesehatan manusia. Journal of Functional Food and Nutraceutical, 63, **2020**.

- JAIN R.A., SHUKLA S.H., SALUJA A.K. In-vitro evaluation of *Clitoria ternatea* stem extract for antioxidant property. IJPSR, 1, 88, 2010.
- REPETTO M.G., LLESUY S.F. Antioxidant properties of natural compounds used in popular medicine for gastric ulcers. Brazilian Journal of Medical and Biological Research, 35, 523, 2002.
- HEYDARI S., REZAEI R., HAGHAYEGH G.H. Effect of drying processes on stability of anthocyanin extracts from saffron petal. Evolving Trends in Engineering and Technology, 2, 13, 2014.
- MARPAUNG A.M., ANDARWULAN N., HARIYADI P., FARIDAH D.N. The difference in colour shifting of Clitoria ternatea L. flower extract at pH 1, 4, and 7 during storage. Current Nutrition & Food Science, 15 (7), 694, 2019.
- 13. SAMI R., KHOJAH E., ALHARBI M., AM AL-MUSHHIN A., SAEED ALKALTHAM M., MOHAMMAD SALAMATULLAH A., AHMED ISMAIL K., HELAL M. Functional effects of pomegranate peel extracts on milk: Antibacterial measurements, antioxidant activities, and photochemical characterizations. Journal of Biobased Materials and Bioenergy, 15 (5), 571, 2021.
- SHITTU A., LIN J., MORRISON D. Molecular identification and characterization of mannitol-negative methicillin-resistant Staphylococcus aureus. Diagnostic Microbiology and Infectious Disease, 57 (1), 93, 2007.
- 15. CHOE J.W., JOO M.K., KIM H.J., LEE B.J., KIM J.H., YEON J.E., PARK J.-J., KIM J.S., BYUN K.S., BAK Y.-T. Foods inducing typical gastroesophageal reflux disease symptoms in Korea. Journal of Neurogastroenterology and Motility, 23 (3), 363, 2017.
- REDDY A., NORRIS D.F., MOMENI S.S., WALDO B., RUBY J.D. The pH of beverages in the United States. The Journal of the American Dental Association, 147 (4), 255, 2016.
- JAYASENA V., CAMERON I. Brix/acid ratio as a predictor of consumer acceptability of Crimson Seedless table grapes. Journal of food Quality, 31 (6), 736, 2008.
- 18. AL-EISA R.A., BUSHNAQ T., RADHI K., HEMDAN D.I., SAMI R., ADNAN ASHOUR A., HELAL M. Physiological effects of honey on cholesterol and triglycerides in albino mice with studying some physical-chemical characterizes and biological activities. Materials Express, 13 (3), 460, 2023.
- DIAS N.A.A., LARA S.B., MIRANDA L.S., PIRES I.S.C., PIRES C.V., HALBOTH N.V. Influence of color on acceptance and identification of flavor of foods by adults. Ciência e Tecnologia de Alimentos, 32 (2), 296, 2012.
- 20. WIYANTOKO B. Butterfly pea *(Clitoria ternatea L.)* extract as indicator of acid-base titration. Indonesian Journal of Chemical Analysis (IJCA), **3** (1), 22, **2020**.
- KHOO H.E., AZLAN A., TANG S.T., LIM S.M. Anthocyanidins and anthocyanins: Colored pigments as food, pharmaceutical ingredients, and the potential health benefits. Food & Nutrition Research. 2017.
- 22. HELAL M., SAMI R., ALGARNI E., ALSHEHRY G., ALJUMAYI H., AL-MUSHHIN A.A.M., BENAJIBA N., CHAVALI M., KUMAR N., IQBAL A. Active bionanocomposite coating quality assessments of some cucumber properties with some diverse applications during storage condition by chitosan, nano titanium oxide crystals, and sodium tripolyphosphate. Crystals, 12 (2), 131, 2022.

PRASAD N., YANG B., KONG K.W., KHOO H.E., SUN J., AZLAN A., ISMAIL A., ROMLI Z. B. Phytochemicals and antioxidant capacity from Nypa fruticans Wurmb. fruit. Evidence-Based Complementary and Alternative Medicine, 2013, 2013.

8

- 24. ROKAYYA S., LI C.-J., ZHAO Y., LI Y., SUN C.-H. Cabbage (*Brassica oleracea* L. var. capitata) phytochemicals with antioxidant and anti-inflammatory potential. Asian Pacific Journal of Cancer Prevention, 14 (11), 6657, 2013.
- 25. OU B., HAMPSCH-WOODILL M., PRIOR R.L. Development and validation of an improved oxygen radical absorbance capacity assay using fluorescein as the fluorescent probe. Journal of Agricultural and Food Chemistry, 49 (10), 4619, 2001.
- 26. WIDOWATI W., WARGASETIA T.L., ZAKARIA T.M., MARTHANIA M., AKBAR R.A.T.P.P., GUNADI M.S., HALIM N., SANTIADI S. Antioxidant activity of TEMON (*Clitoria ternatea* and *Citrus sp.*) as an infused herbal tea. Majalah Obat Tradisional, 27 (1), 32, 2022.
- 27. LISTER I.N.E., GINTING C.N., GIRSANG E., ARMANSYAH A., MARPAUNG H.H., SINAGA A.P.F., HANDAYANI R.A.S., RIZAL R. Antioxidant properties of red betel (*Piper crocatum*) leaf extract and its compounds. Journal of Natural Remedies, 198, 2019.
- 28. ZOR M., SENGUL M., KARAKÜTÜK İ.A., ODUNKIRAN A. Changes caused by different cooking methods in some physicochemical properties, antioxidant activity, and mineral composition of various vegetables. Journal of Food Processing and Preservation, 46 (11), e16960, 2022.
- 29. MAZZUCOTELLI C.A., GONZÁLEZ-AGUILAR G.A., VILLEGAS-OCHOA M.A., DOMÍNGUEZ-AVILA A.J., ANSORENA M.R., DI SCALA K.C. Chemical characterization and functional properties of selected leafy vegetables for innovative mixed salads. Journal of Food Biochemistry, 42 (1), e12461, 2018.
- JEYARAJ E.J., LIM Y.Y., CHOO W.S. Effect of organic solvents and water extraction on the phytochemical profile and antioxidant activity of *Clitoria ternatea* flowers. ACS Food Science & Technology, 1 (9), 1567, 2021.
- 31. AHMAD A.N., ABDULLAH LIM S., NAVARANJAN N. Development of sago (Metroxylon sagu)-based colorimetric indicator incorporated with butterfly pea *(Clitoria ternatea)* anthocyanin for intelligent food packaging. Journal of Food Safety, **40** (4), e12807, **2020**.
- 32. DZOYEM J.P., MCGAW L.J., ELOFF J.N. In vitro antibacterial, antioxidant and cytotoxic activity of acetone leaf extracts of nine under-investigated Fabaceae tree species leads to potentially useful extracts in animal health and productivity. BMC Complementary and Alternative Medicine, 14, 1, 2014.
- 33. DACULLO R.M.A., BITACURA J.G. In Vitro Antibacterial, Anticoagulant, and Antioxidant Screening of Aqueous Extracts of Blue Ternate (Clitoria ternatea L.) Flower. Herbal Medicines Journal, 7, 2022.
- 34. RANGEL C.N., CARVALHO L.M.J.D., FONSECA R.B.F., SOARES A.G., JESUS E.O. D. Nutritional value of organic acid lime juice (Citrus latifolia T.), cv. Tahiti. Food Science and Technology, 31, 918, 2011.
- FEATHERSTONE S. A complete course in canning and related processes: Volume 3 Processing Procedures for Canned Food Products. Woodhead Publishing, 2015.

- 36. DINÇER C., TONGUR T., ERKAYMAZ T. Investigation of the effect of different extraction methods on the quality properties of hibiscus extracts. 2020.
- 37. LIU Y., LUO L., LIAO C., CHEN L., WANG J., ZENG L. Effects of brewing conditions on the phytochemical composition, sensory qualities and antioxidant activity of green tea infusion: A study using response surface methodology. Food Chemistry, 269, 24, 2018.
- 38. BOONSIRIWIT A., LEE M., KIM M., INTHAMAT P., SIRIPATRAWAN U., LEE Y.S. Hydroxypropyl methylcellulose/microcrystalline cellulose biocomposite film incorporated with butterfly pea anthocyanin as a sustainable pH-responsive indicator for intelligent foodpackaging applications. Food Bioscience, 44, 101392, 2021.
- 39. ADAMS R. Sensient's new blue food colorant, derived from butterfly pea flower. Focus on Pigments. Elsevier BV, Radarweg, 29, 1043, 2022.
- 40. MENG X., ZHANG M., ZHAN Z., ADHIKARI B. Changes in quality characteristics of fresh-cut cucumbers as affected by pressurized argon treatment. Food and Bioprocess Technology, 7, 693, 2014.
- 41. AMAOLO A. The butterfly pea flower as a pH indicator. International Scholastic Journal of Science, 11 (1), 2, 2017.
- 42. POH N.A.B.A. Research of the ecology and morphology of *Clitoria ternatea*. Singapore School of Science and Technology. **2019**.
- RABETA M.S., AN NABIL Z. Total phenolic compounds and scavenging activity in *Clitoria ternatea* and *Vitex* negundo linn. International Food Research Journal, 20 (1), 2013
- 44. BENAJIBA N., KHOJAH E. DPPH, FRAP and TAEC Assays with Postharvest Cabbage (Brassica oleracea) Parameters During the Packaging Process. Pakistan Journal of Biological Sciences: Pakistan Journal of Biological Sciences, 24 (2), 182, 2021.
- 45. WIDOWATI W., WARGASETIA T.L., ZAKARIA T.M., MARTHANIA M., GUNADI M.S., HALIM N., DEWI N.S.M., SANTIADI S. Antioxidant activities of ginger (Zingiber officinale) and telang flower (Clitoria ternatea L.) combination tea. Majalah Kedokteran Bandung, 54 (3), 154, 2022.
- 46. CHANG M.-Y., LIN Y.-Y., CHANG Y.-C., HUANG W.-Y., LIN W.-S., CHEN C.-Y., HUANG S.-L., LIN Y.-S. Effects of infusion and storage on antioxidant activity and total phenolic content of black tea. Applied Sciences, 10 (8), 2685, 2020.
- 47. WONG S.P., LEONG L.P., KOH J.H.W. Antioxidant activities of aqueous extracts of selected plants. Food Chemistry, **99** (4), 775, **2006**.
- 48. HUAMÁN-CASTILLA N.L., MARIOTTI-CELIS M.S., MARTÍNEZ-CIFUENTES M., PÉREZ-CORREA J.R. Glycerol as alternative co-solvent for water extraction of polyphenols from Carménère pomace: Hot pressurized liquid extraction and computational chemistry calculations. Biomolecules, 10 (3), 474, 2020.
- 49. CACACE J.E., MAZZA G. Mass transfer process during extraction of phenolic compounds from milled berries. Journal of Food Engineering, **59** (4), 379, **2003**.
- ACAR A., AYDIN M., ARSLAN D. Development of infusion tea formulations with food wastes: Evaluation of temperature and time effects on quality parameters. Applied Food Research, 2 (1), 100087, 2022.
- 51. KILIÇ C., CAN Z., YILMAZ A., YILDIZ S., TURNA H. Antioxidant properties of some herbal teas (green tea, senna, corn silk, rosemary) brewed at different

- temperatures. International Journal of Secondary Metabolite, **4** (3, Special Issue 1), 142, **2017**.
- MIGUEL M.G. Anthocyanins: Antioxidant and/or antiinflammatory activities. Journal of Applied Pharmaceutical Science, 07, 2011.
- 53. SAPTARINI N.M., SURYASAPUTRA D. Total anthocyanins content in various extract of butterfly pea (Clitoria ternatea linn) flower. Research Journal of Pharmaceutical Biological and Chemical Sciences, 9 (2), 185. 2018.
- 54. SOFYAN J., PERMANA T., MARPAUNG A.M. The Study of Several Applicable Treatments for Serving Butterfly Pea Flower Drinks. Atlantis Press,
- 55. LÓPEZ PRADO A. S., SHEN Y., ARDOIN R., OSORIO L.F., CARDONA J., XU Z., PRINYAWIWATKUL W. Effects of different solvents on total phenolic and total anthocyanin contents of *Clitoria ternatea L.* petal and their anti-cholesterol oxidation capabilities. International Journal of Food Science & Technology, 54 (2), 424, 2019.
- 56. MELEK Z.O.R., ŞENGÜL M., KARAKÜTÜK İ.A., AKSOY S. Investigation about Various Infusion Conditions on Physical, Chemical and Antioxidant Properties of *Clitoria ternatea L.* Tea. Journal of the Institute of Science and Technology, 13 (3), 1738, 2023.