DOI: 10.15244/pjoes/208930

ONLINE PUBLICATION DATE: 2025-10-02

Original Research

# Effects of Combined Supplementation of Germanium and Selenium on the Antioxidant Function of Przewalski's Gazelles

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Received: 2 March 2025 Accepted: 3 August 2025

## **Abstract**

The Przewalski's gazelle (P. przewalskii) is a rare and endangered species endemic to China, primarily distributed in the Qinghai-Tibet Plateau region. Due to the unique characteristics of the plateau environment, P. przewalskii often faces issues such as nutritional deficiencies and oxidative stress. This study focused on the differences between P. przewalskii in different areas of Qinghai Lake, revealing that P. przewalskii in the Hudong region exhibited symptoms of emaciation, sluggish movement, and lethargy, with distinct differences in body size and shape compared to those in the Bird Island region. Serum parameter analysis showed that the antioxidant function of P. przewalskii in the Bird Island region was significantly higher than that in the Hudong region (p < 0.05). The levels of germanium (Ge) and selenium (Se) in the soil, forage, and serum of P. przewalskii in the Hudong region were significantly lower than those in the Bird Island region (p < 0.05). To further evaluate the therapeutic effects of combined Ge and Se supplementation, this study designated P. przewalskii in the Bird Island region as the control group and those in the Hudong region as the experimental group. The experimental group was orally administered 5 mg/kg·BW·day of Ge-132 and 1 mg/kg·BW·day of Na, SeO,. The results showed significant improvements in antioxidant indicators (T-AOC +36.3%, MDA -50.6%) and immune indicators (IgG +29.8%, IgM +17.1%), and inflammatory factors (IL-6 -31.2%, TNF- $\alpha$  -26.5%) were significantly reduced (p<0.05). The combined Ge-Se supplementation enhanced antioxidant function (T-AOC +5.5%, MDA -9.3%) and immunity (IgG +11.0%, IgM +9.8%) significantly more than the best individual supplement (p<0.05). After 30 days of supplementation, antioxidant indices showed no significant differences between groups (p>0.05). In conclusion, the combined application of Ge and Se demonstrated the best therapeutic effect on the antioxidant capacity of P. przewalskii. This discovery not only provided new insights for improving the habitat of P. przewalskii but also represented a significant breakthrough in the conservation efforts for rare species.

Keywords: germanium, selenium, P. przewalskii, antioxidant function

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# Introduction

The Przewalski's gazelle (P. przewalskii) belongs to the genus Procapra within the subfamily Antilopinae of the family Bovidae in the order Artiodactyla. It has a narrow distribution range and is primarily found in Qinghai, Ningxia, Inner Mongolia, and Gansu [1]. For P. przewalskii, antioxidant function is crucial, and minerals are closely related to this antioxidant capability [2]. A deficiency in minerals can lead to a decline in the animal's immunity and antioxidant capacity, which further inhibits their reproduction and growth [3]. In the 1990s, it was estimated that only 200-300 individuals survived around Qinghai Lake [4]. After more than 20 years of continuous conservation and research, the population of P. przewalskii has partially recovered but remains below 2,000 individuals [5]. Therefore, the endangered status of P. przewalskii has attracted widespread attention.

Antioxidant substances can help animals resist oxidative stress, a condition caused by excessive free radicals in the body, which may lead to cellular damage and functional impairment [6, 7]. Antioxidants protect the health of cells and tissues by neutralizing free radicals [8]. The antioxidant system consists of enzymatic and nonenzymatic systems. The enzymatic system is composed of antioxidant enzymes, such as superoxide dismutase (SOD), glutathione peroxidase (GSH-PX), and catalase (CAT) [9]. The non-enzymatic system includes elements such as copper (Cu), iron (Fe), zinc (Zn), and glutathione (GSH), which can effectively eliminate excess free radicals in the body. This prevents the chain reaction of lipid peroxidation, thereby reducing the accumulation of malondialdehyde (MDA) [10, 11]. MDA is closely related to antioxidants [12, 13]. Animals with strong antioxidant functions typically exhibit higher immunity, enabling them to resist diseases and infections more effectively [14, 15].

Previous studies have conclusively confirmed that Se plays an important role in improving the antioxidant function of *P. przewalskii* [16]. Selenium (Se) is an essential component of selenoproteins and various antioxidant enzymes, such as GSH-PX, thioredoxin reductase, and iodothyronine deiodinase. These enzymes protect cells from the harmful effects of free radicals generated during oxidation processes [17].

Organic germanium can promote the synthesis and activity of antioxidant enzymes, such as SOD and GSH-PX, thereby enhancing the antioxidant function of cells [18, 19]. Among various organic germanium compounds, Ge-132 is considered to have the lowest toxicity and plays an important role in maintaining health [20, 21]. Ge-132 has a significant free radical scavenging effect; it can enhance SOD activity and improve the ability to resist lipid peroxidation [22]. In addition, it has been shown to possess various biological activities, including immune stimulation, anti-tumor effects, pain relief, and anti-inflammatory activity [23, 24].

Despite the well-established individual roles of selenium (Se) and germanium (Ge) in antioxidant function, the potential synergistic effects of combining Se with other trace elements such as germanium (Ge) remain unexplored. Critically, despite the individual antioxidant benefits of both elements, no prior studies have investigated their combined use in P. przewalskii or similar endangered species, nor have interactions between Se and Ge and their potential synergistic impacts on antioxidant capacity been examined in this context. Given the limited efficacy of single-element interventions in complex ecological systems and animal health maintenance, this study addresses a critical research gap by evaluating the combined effects of Ge and Se supplementation in P. przewalskii [25]. This innovative approach advances understanding of multielement synergism in antioxidant defense and establishes a novel strategy for enhancing health management and conservation of this endangered species.

#### **Materials and Methods**

# Study Location

The study area was located in the region surrounding Qinghai Lake (99°36′-100°16′ E, 36°32′-37°15′ N). The elevation ranges from 3,195 to 3,305 m, characterized by an extended cold season, intense solar radiation, large daily temperature fluctuations, and arid conditions with little rainfall. The annual average temperature ranges from -1°C to -1.5°C, with cool and cloudy summers and cold, dry winters [26]. The main vegetation types include Oxytropis aciphylla Ledeb, Lagotis brachystachya Maxim, Stellera chamaejasme, Iris loczyi, Kobresia littledalei, Kobresia tibetica, Kobresia schoenoides, and Blysmus sinocompressus [27].

# Experimental Design

This study was conducted in the Hudong and Bird Island Nature Reserves within the Qinghai Lake region in June 2024. Sixteen P. przewalskii with obvious differences in body size and posture from those in the Hudong area were selected in the Bird Island area as the control group, allowing them to graze freely on local forage in the natural environment. In the Hudong District, a designated enclosed area of 40 hm<sup>2</sup> has been established as an experimental zone. Within this area, 16 P. przewalskii of similar age and physical condition, exhibiting symptoms such as emaciation, sluggish movement, and lethargy, were selected. These gazelles were allowed to graze freely on local pasture in their natural environment and were randomly divided into four groups. These gazelles are allowed to graze freely on local pasture in their natural environment. The experimental groups were set as follows: Group I: No treatment. Group II: Administered 5 mg/kg BW·day of Ge-132 orally each day [28-30].

Group III: Administered 1 mg/kg BW·day of Na<sub>2</sub>SeO<sub>3</sub> orally each day [31-35]. Group IV: Administered 5 mg/kg BW·day of Ge-132 and 1 mg/kg BW·day of Na<sub>2</sub>SeO<sub>3</sub> orally each day. The Ge-132 and Na<sub>2</sub>SeO<sub>3</sub> used in this experiment were purchased from Shanghai Aladdin Biochemical Technology Co., Ltd., with a purity of > 99% for both compounds.

# Sample Collection

Through systematic observation of the activity areas and foraging behavior of *P. przewalskii*, 10 sampling plots of 1000 m<sup>2</sup> each were randomly established along their foraging routes in both regions, with an interval distance of 500 m between plots.

At each sampling site, the S-shaped sampling method was used to select 4 points as soil sampling locations. During soil sample collection, a shovel was used to remove surface vegetation, fallen leaves, and other impurities. Then, approximately 0-30 cm of topsoil was collected, with about 250 g taken from each sampling point. The 4 soil samples from each site were crushed, and impurities such as grass roots were removed. The samples were then thoroughly mixed, and the quartering method was used to retain 100 g. After soil sampling was completed, the samples were labeled in preparation for mixed forage collection. The soil samples were air-dried indoors at 20-25°C in a cool place, crushed, sieved through a 2 mm mesh, and fine sand was removed using a 0.075 mm sieve. Finally, the samples were bagged, labeled, and stored in a silica gel vacuum desiccator for later use.

Within each sampling plot, mixed forage samples were collected at intervals of more than 500 m (each sample weighed 500 g), resulting in a total of 20 samples (10 from each of the Hudong and Bird Island regions). During sample collection, stainless steel scissors were used to cut the vegetation at a height of 1-2 cm above the ground to avoid soil contamination. The sample processing procedure was as follows: First, the collected mixed forage samples were naturally dried at a constant temperature of 20-25°C until a constant weight was achieved. Subsequently, the dried samples were crushed using a mortar and sequentially passed through 2 mm and 0.075 mm standard sieves to remove impurities and fine sand. Finally, the processed samples were stored in a vacuum drying oven (Shanghai Boxun Industrial Co., Ltd., BOV series) for subsequent chemical analysis. All sample processing steps were conducted in a clean laboratory environment to ensure sample quality.

Before the experiment began, we selected 16 *P. przewalskii* from each of the Hudong and Bird Island regions of Qinghai Lake and collected whole blood samples. Blood was collected from experimental animals on days 10, 20, 30, 40, and 50. After the *P. przewalskii* were manually restrained, whole blood samples were collected from their jugular veins using a blood collection needle (Weigao Group, Shandong, China) and a special vacuum blood collection tube for high-

altitude areas without additives (SL-001, Wuhan Sanli Medical Technology Company, Wuhan, China). All the collected samples were numbered and labeled to ensure the accuracy of the data. The whole blood samples were left standing at room temperature for 20 to 30 minutes. After they naturally coagulated, a centrifuge (ZJU-3000, Jinhua Shenzhou Centrifuge Co., Ltd, Zhejiang, China) was used to centrifuge the samples at a low speed (1000 to 3000 rpm) for 10 to 15 minutes to separate the serum for subsequent analysis.

# Sample Preparation and Analysis

The soil samples were digested using microwave digestion. Before the experiment, the digestion tubes were rinsed with deionized water. Approximately 0.5 g of soil sample was placed into a digestion tube, followed by the addition of 6 mL of nitric acid (HNO<sub>2</sub>), 3 mL of hydrochloric acid (HCl), and 2 mL of hydrofluoric acid (HF). The mixture was shaken well and allowed to stand for about 5 minutes. The lid was then tightly screwed on, and the digestion tube was placed into the digestion rack and transferred to the turntable of the microwave oven. After confirming that the pressure and temperature sensors of the digestion instrument were functioning properly, the samples were automatically digested according to the microwave digestion program. After cooling, the lid was loosened, and the solution was transferred to a polytetrafluoroethylene (PTFE) crucible for acid evaporation. Subsequently, the solution was transferred to a 50 mL volumetric flask, diluted to the mark, labeled, and set aside for further use.

Nitric acid (HNO<sub>3</sub>), hydrofluoric acid (HF), and perchloric acid (HClO<sub>4</sub>) were mixed in a volume ratio of 5:2:5 and added to the mixed forage samples, followed by thorough shaking. Subsequently, a mixed solution of HNO<sub>3</sub> and HClO<sub>4</sub> in a 4:1 volume ratio was added to the forage samples and shaken well.

The concentrations of mineral elements in the mixed soil-forage from Hudong and Bird Island, as well as in animal serum, were determined using inductively coupled plasma atomic emission spectroscopy (ICP-AES, HK9600, Huakexing Co., Beijing, China). The analyzed elements included Ge, Se, Cu, Zn, and Fe.

# Blood Biochemical Analysis

Antioxidant and immune function indicators in blood samples were tested. The diagnostic kits used were provided by Nanjing Biotechnology Co., Ltd. (Jiangsu, China). The antioxidant capacity indicators included T-AOC, CAT, GSH-PX, SOD, and MDA. The immunological indicators included IgG (Immunoglobulin G), TNF-α (Tumor Necrosis Factoralpha), IL-2 (Interleukin-2), IgA (Immunoglobulin A), IL-1β (Interleukin-1 beta), IgM (Immunoglobulin M), and IL-6 (Interleukin-6).

# Statistical Analysis

Data were processed using SPSS statistical software (SPSS, version 23.0, Inc., Chicago, Illinois, USA). After confirming homogeneity of variances by Levene's test (p>0.05) for all elements, differences among groups were analyzed by one-way ANOVA followed by Tukey's post-hoc test for multiple comparisons. Results are expressed as mean $\pm$ standard error (SE). Significant differences (p<0.05) are denoted in tables by superscript letters (a, b, c) and in figures by asterisks (\*).

## Results

Antioxidant and Immune-Inflammatory Profiles of *P. przewalskii* in Qinghai Lake and Bird Island Pre-Experiment

As shown in Fig. 1, a significant difference in antioxidant indicators was observed between the Bird Island group and the Hudong group of *P. przewalskii* 

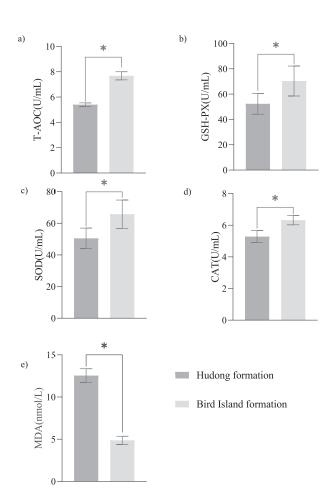


Fig. 1. The blood antioxidant indexes before the experiment: a) T-AOC, total antioxidant capacity; b) GSH-PX, glutathione peroxidase; c) SOD, superoxide dismutase; d) CAT, catalase; e) MDA, malondialdehyde.

Notes: Error bars represent standard error, SE.

(p<0.05). The serum levels of T-AOC, GSH-PX, SOD, and CAT in the Bird Island group were significantly higher than those in the Hudong group (p<0.05). Additionally, the MDA levels in the Bird Island group were significantly lower compared to the Hudong group (p<0.05).

Fig. 2 illustrates significant differences in multiple immune and inflammatory indicators between the Eastern Qinghai Lake and Bird Island groups of *P. przewalskii*. The concentrations of IL-1 $\beta$ , IL-6, IL-2, and TNF- $\alpha$  were significantly higher in the Hudong group compared to the Bird Island group (p<0.05). Conversely, the concentrations of IgA, IgG, and IgM were significantly higher in the Bird Island group (p<0.05).

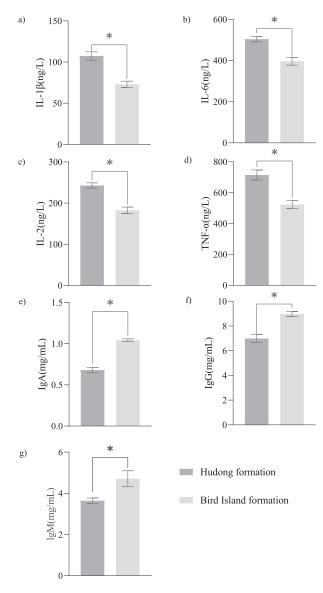


Fig. 2. The blood immune and anti-inflammatory indexes before the experiment: a) IgA, immunoglobulin A; b) IgG, immunoglobulin G; c) IgM, immunoglobulin; d) IL-1 $\beta$ , interleukin-1 $\beta$ ; e) IL-2, interleukin-2; f) IL-6, interleukin-6; g) TNF- $\alpha$ , tumor necrosis factor-alpha.

Notes: Error bars represent standard error, SE.

<sup>\*</sup> indicates great difference at p<0.05.

<sup>\*</sup> indicates great difference at p < 0.05.

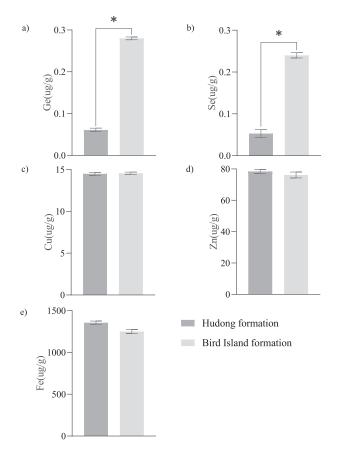


Fig. 3. The mineral content in the soil: a) Ge, germanium; b) Se, selenium; c) Cu, copper; d) Zn, zinc; e) Fe, iron. Notes: Error bars represent standard error, SE.

\* indicates great difference at *p*<0.05

As shown in Fig. 3, the contents of Ge and Se in the soil from the Bird Island area of Qinghai Lake were significantly higher than those in the Hudong Qinghai Lake area (p<0.05). Meanwhile, the contents of other mineral elements, such as Cu, Zn, and Fe, showed no significant differences between the two areas (p>0.05).

As shown in Fig. 4, the contents of Ge and Se in the mixed forage from the Bird Island area of Qinghai Lake were significantly higher than those in the Hudong Qinghai Lake area (p<0.05). Meanwhile, the contents of other mineral elements, such as Cu, Zn, and Fe, showed no significant differences between the two areas (p>0.05).

As shown in Fig. 5, the serum levels of Ge and Se in *P. przewalskii* from the Bird Island area of Qinghai Lake were significantly higher than those in the Hudong Qinghai Lake area (p<0.05). However, there were no significant differences in the levels of other mineral elements, such as Cu, Zn, and Fe, between the two regions (p>0.05).

# Serum Mineral Content of Experimental Groups of *P. przewalskii*

As shown in Table 1, there were no significant differences in mineral element levels in Group I

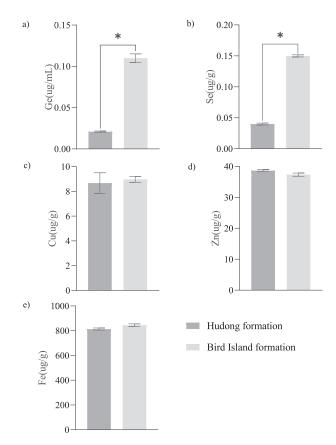


Fig. 4. The mineral content in the mixed forage: a) Ge, germanium; b) Se, selenium; c) Cu, copper; d) Zn, zinc; e) Fe, iron).

Notes: Error bars represent standard error, SE.

\* indicates great difference at p<0.05

(p>0.05). The Ge levels in Groups II and IV increased significantly (p<0.05), while the Se levels in Groups III and IV also showed significant increases (p<0.05). Over the 50 days, there were no significant changes in the levels of Cu, Zn, and Fe (p>0.05).

# Serum Antioxidant Parameters of Experimental Groups of *P. przewalskii*

There was no significant difference in T-AOC, GSH-PX, SOD, CAT, and MDA activity in Group I (p>0.05). In Groups II, III, and IV, serum T-AOC, GSH-PX, SOD, and CAT activity increased significantly during the first 0-30 days (p<0.05) but showed no significant difference after 30 days (p>0.05) (Fig. 6(a-d)).

In Groups II, III, and IV, serum MDA activity decreased significantly during the first 0-30 days (p<0.05) but showed no significant difference after 30 days (p>0.05) (Fig. 6e)).

# Serum Immune and Inflammatory Indicators of *P. przewalskii*

Group I received no treatment. Group II was orally administered 5 mg/kg·BW of Ge-132 daily. Group III

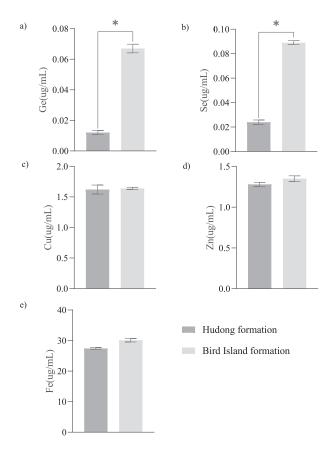


Fig. 5. The mineral content in the blood: a) Ge, germanium; b) Se, selenium; c) Cu, copper; d) Zn, zinc; e) Fe, iron.

Notes: Error bars represent standard error, SE.

\* indicates great difference at p<0.05

was orally administered 1 mg/kg·BW of Na<sub>2</sub>SeO<sub>3</sub> daily. Group IV was orally administered 5 mg/kg·BW of Ge-132 combined with 1 mg/kg·BW of Na<sub>2</sub>SeO<sub>3</sub> daily.

As shown in Table 2, the levels of IL-1 $\beta$ , IL-2, TNF- $\alpha$ , IgA, IgG, IL-6, and IgM in Groups II, III, and IV were significantly higher than those in Group I. In Group IV, these levels returned to levels comparable to those of the control group.

# **Discussion**

The desertification of habitats in the Qinghai Lake region has impacted the quality of local soil [36]. Subsequently, this has affected the mineral element content in pasture. Pasture serves as the primary pathway for the transfer of mineral elements from soil to animals, and the levels of Ge and Se in animals are crucial for their antioxidant function [37-39]. This study aimed to enhance the antioxidant function of *P. przewalskii* through the combined supplementation of Ge and Se, thereby boosting its immune function. This approach holds significant ecological and conservation value for protecting this endangered species.

In this study, we analyzed the mineral element contents of mixed pastures from the Hudong and Bird Island regions, as well as the hematological parameters of *P. przewalskii*. The results showed that the Ge and Se contents in mixed pasture from the Hudong region were substantially lower than those in the Bird Island region. Similarly, the antioxidant parameters in the serum of *P. przewalskii* from the Hudong region were significantly lower than those of gazelles from the Bird Island region. This suggests that Ge and Se may play a crucial role in enhancing the antioxidant function of *P. przewalskii*. To validate this hypothesis, we conducted supplementation experiments with Ge and Se on *P. przewalskii* in the Hudong region of Qinghai Lake. The antioxidant function of *P. przewalskii* can be

Table 1. Concentrations of Ge, Se, Cu, Zn, and Fe in the blood of *P. przewalskii* in the Hudong region.

Groups									
Days	Elements	I	II	III	IV				
0d	Ge(ug/ml)	0.016±0.0016	0.017±0.0004	0.017±0.0026	0.016±0.0003				
	Se(ug/ml)	0.021±0.0006	0.021±0.0012	0.022±0.0007	0.021±0.0001				
	Cu(ug/ml)	1.59±0.16	1.63±0.14	1.65±0.16	1.60±0.19				
	Zn(ug/ml)	1.21±0.044	1.25±0.031	1.22±0.040	1.23±0.077				
	Fe(ug/ml)	25.47±1.98	25.34±29.45	25.69±1.67	25.94±0.93				
50d	Ge(ug/ml)	0.016±0.0007 <sup>b</sup>	0.067±0.0009a	0.017±0.0016 <sup>b</sup>	0.072±0.0015a				
	Se(ug/ml)	0.022±0.0002°	0.021±0.0006°	0.074±0.0003 <sup>b</sup>	0.083±0.0004ª				
	Cu(ug/ml)	1.59±0.15	1.60±0.13	1.56±0.07	1.62±0.05				
	Zn(ug/ml)	1.20±0.14	1.21±0.09	1.20±0.12	1.22±0.03				
	Fe(ug/ml)	24.85±1.69	25.12±1.08	24.88±1.21	25.20±0.99				

Notes: Ge, germanium; Se, selenium; Cu, copper; Zn, zinc; Fe, iron.

Within each row, values with different superscript letters (a, b, c) differ significantly (p < 0.05).

<sup>\*</sup> At the level of p < 0.05.

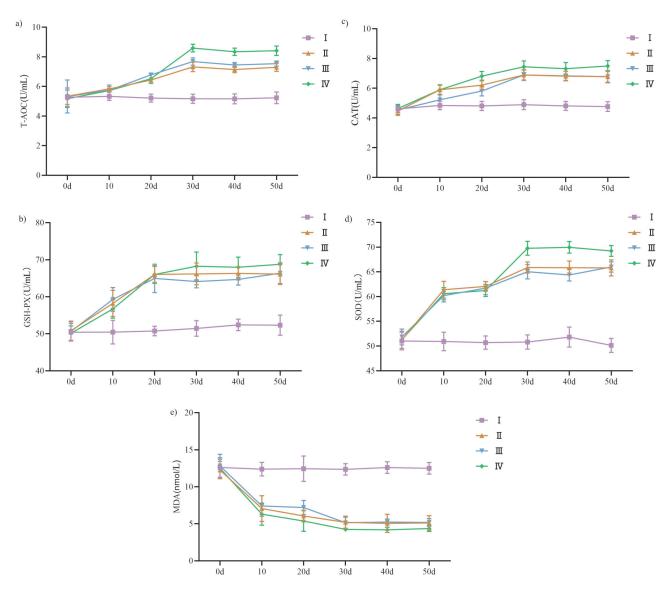


Fig. 6. Antioxidant indicators: a) T-AOC, total antioxidant capacity; b) GSH-PX, glutathione peroxidase; c) SOD, superoxide dismutase; d) CAT, catalase; e) MDA, malondialdehyde; I: Group II; III: Group III; IV: Group IV. Notes: Error bars represent standard error, SE.

evaluated through trace elements and antioxidant parameters in their serum. In our experiment, Se supplementation significantly improved the antioxidant function of *P. przewalskii*. This finding further validates the critical role of Se in enhancing the antioxidant capacity of animals. It suggests that appropriate Se supplementation in the management and feeding of *P. przewalskii* can effectively boost their antioxidant capacity and improve their overall health.

Ge plays a vital role in enhancing the antioxidant function of animals [40, 41]. Ge is a trace element that is typically present in low concentrations in plants, primarily influenced by the Ge content in the soil and the surrounding environment [42, 43]. Although Ge is widely found in various plants, its concentration is relatively low [44]. Ge can be classified into inorganic and organic forms, with inorganic Ge exhibiting lower biological and pharmacological activity compared to

organic Ge [45]. Ge can improve insulin sensitivity in animals [46], inhibit damage caused by free radicals, and exert antioxidant effects [18]. Our study results showed that Ge supplementation significantly improved the antioxidant function of *P. przewalskii*. This finding further validates the critical role of Ge in enhancing the antioxidant capacity of animals. It indicates that appropriate Ge supplementation in the management and feeding of *P. przewalskii* can effectively boost their antioxidant capacity and improve their overall health.

The Ge and Se content in the serum of *P. przewalskii* significantly increased after Ge and Se supplementation. Meanwhile, the results of antioxidant parameter analysis indicated that supplementation with Ge-132 and sodium selenite significantly enhanced the T-AOC, GSH-PX, CAT, and SOD activities in the serum. Changes in their activity and levels reflect the antioxidant function and health status of *P. przewalskii*. Under conditions of

Table 2. The differences	in the immune	inflammation	index of <i>P</i> .	przewalskii among variou	s groups.

Immune -inflammatory	Groups					
markers	I	II	III	IV		
IgA(mg/mL)	0.680±0.066°	0.897±0.052b	$0.864 \pm 0.076^{b}$	1.060±0.09ª		
IgG(mg/mL)	6.99±0.62°	8.17±0.024 <sup>b</sup>	8.14±0.016 <sup>b</sup>	9.07±0.64ª		
IgM(mg/mL)	3.51±0.13°	3.74±0.26 <sup>b</sup>	3.77±0.12 <sup>b</sup>	4.11±0.36a		
IL-1β (ng/L)	104.91±10.28 <sup>a</sup>	79.87±8.12 <sup>b</sup>	78.76±5.98 <sup>b</sup>	72.98±7.64°		
IL-6 (ng/L)	502.96±55.47ª	379.45±32.87 <sup>b</sup>	375.98±24.54 <sup>b</sup>	346.15±38.17 <sup>C</sup>		
IL-2(ng/L)	240.24±18.01ª	200.67±19.26 <sup>b</sup>	203.17±16.98 <sup>b</sup>	182.54±15,78°		
TNF-α(ng/L)	711.47±79.51ª	563.88±45.42 <sup>b</sup>	580.01±72.76 <sup>b</sup>	523.33±50.41°		

Notes: IgA, immunoglobulin A; IgG, immunoglobulin G; IgM, immunoglobulin; IL-1 $\beta$ , interleukin-1 $\beta$ ; IL-2, interleukin-2; IL-6, interleukin 6; TNF- $\alpha$ , tumor necrosis factor-alpha.

Within each row, values with different superscript letters (a, b, c) differ significantly (p < 0.05).

disease or environmental stress, the activity of these enzymes increases to counteract the elevated levels of free radicals in the body [47, 48]. Meanwhile, it significantly reduced the content of MDA. MDA levels serve as a marker for the degree of lipid peroxidation, with higher MDA levels indicating an increased extent of lipid peroxidation, which indirectly reflects the severity of cellular damage [49, 50]. The supplementation of Ge and Se, either individually or in combination, had significant impact on the growth, element accumulation, and ergosterol content of Ganoderma and Pleurotus fruiting bodies. The synergistic effect of Ge and Se plays an important role in the accumulation of these elements in the studied mushroom species [51]. With increasing concentrations of Se or Ge, the free radical scavenging rate was significantly enhanced. Moreover, the combined use of Ge and Se exhibited a superior free radical scavenging effect compared to their individual use [52]. The combined supplementation of Se and Ge enhances glutathione peroxidase (GPX) activity and GSH levels while reducing MDA levels, thereby exerting antioxidant effects and protecting the body from oxidative damage [53].

We also measured the immune indicators in the serum of P. przewalskii. The combined use of Ge and Se significantly enhanced the immune capacity of P. przewalskii [54]. When the body's antioxidant function declines, reactive oxygen species (ROS), such as oxygen free radicals, increase, leading to lipid peroxidation. This, in turn, causes tissue damage, cellular metabolic disorders, and a reduction in immune function [55-59]. Ge-132 has been shown to induce the production of endogenous interferons, enhance natural killer (NK) cell activity, activate macrophages, promote antibody production, and exhibit biological effects such as antitumor and anti-aging properties [60]. In addition, Se influences immune and inflammatory responses by regulating macrophage activity, phagocytic capacity, and cytokine secretion [61]. Studies have shown

that Se polysaccharides can enhance macrophage immune activity and improve the immune function of immunosuppressed and Se-deficient mice by activating the MAPK signaling pathway and regulating the expression of related proteins and transcription factors [62]. Combined supplementation of Ge and Se significantly improved the antioxidant function of P. przewalskii, with effects superior to those of supplementing either Ge or Se alone. This finding further confirms the synergistic effect of Ge and Se in enhancing antioxidant function, providing strong scientific support for the health management of P. przewalskii. Therefore, Ge and Se can each enhance the antioxidant function of P. przewalskii when used individually, but their combined use demonstrates a more pronounced effect, effectively strengthening the body's defense against oxidative damage. In addition, during the 50-day period of daily oral administration of 5 mg/kg BW/day Ge-132 and 1 mg/kg BW/day Na<sub>2</sub>SeO<sub>3</sub>, the antioxidant parameters of P. przewalskii returned to levels consistent with those of the P. przewalskii from the Bird Island region by the 30th day. From day 30 onwards, all parameters remained stable. In the past, melatonin treatment of Sarda sheep for 35 days significantly improved their reproductive performance [63]. Plasma rich in growth factors shows significant therapeutic effects on grade II muscle injuries in sheep after approximately 30 days of treatment [64]. In addition, to ensure adequate Se intake, Se supplementation was administered approximately 30 days before lambing, significantly increasing Se levels in the ewes [65]. The aforementioned cases all indicate that a 30-day treatment yields significant effects, which aligns with the conclusions of this study. Therefore, for the P. przewalskii gazelles in the Hudong area of Qinghai Lake that exhibit adverse symptoms such as emaciation, sluggish movement, and listlessness due to the deficiency of Ge and Se, a treatment regimen of jointly supplementing germanium and selenium for

<sup>\*</sup> At the level of p<0.05.

30 days can be adopted. In summary, the combined use of Ge and Se in *P. przewalskii* can significantly improve their antioxidant function. This supplementation not only enhances the stability of antioxidant indicators but also boosts immune function, providing effective support for the health and well-being of *P. przewalskii*.

#### **Conclusions**

This study found that the *P. przewalskii* living in the Hudong region of Qinghai Lake exhibited significant weight loss and lethargy due to deficiencies in Ge and Se. To address this issue, the combined supplementation of Ge and Se significantly improved the antioxidant and immune-inflammatory indicators of P. przewalskii, positively impacting their antioxidant function. Based on these findings, we recommend a treatment regimen for P. przewalskii showing significant weight loss and lethargy due to Ge and Se deficiencies. The regimen involves a 30-day cycle of daily oral administration of 5 mg/kg·BW·day of Ge-132 and 1 mg/kg·BW·day of Na<sub>2</sub>SeO<sub>3</sub>. This dosage ensures optimal antioxidant effects and aids in restoring the overall health of the P. przewalskii. This study not only provides a scientific basis for the health management of P. przewalskii but also offers strong support for their population recovery and ecological conservation. In the future, we recommend further research on the effects of Ge and Se supplementation under different environmental conditions to optimize dosage and administration methods, ensuring effective application in broader wildlife conservation practices.

# Acknowledgements

This work was supported by the National Natural Science Foundation of China (42171060), the Innovation Development Supporting Plan Project of Key Industries in Southern Xinjiang (2021DB014), and the Modern Agricultural Industry Technology System in China (CARS-38).

# **Conflict of Interest**

The authors declare no conflict of interest.

## References

- LIANG D., LI C. Habitat Suitability, Distribution Modelling and GAP Analysis of Przewalski's Gazelle Conservation. Animals. 14 (1), 149, 2024.
- ZHANG Q., ZHAO K., SHEN X. Metabolomic Analysis Reveals the Adaptation in the P. przewalskii to Se-Deprived Environment. Biological Trace Element Research. 200 (8), 3608, 2021.

- AMIR ASLANI B., GHOBADI S. Studies on oxidants and antioxidants with a brief glance at their relevance to the immune system. Life Sciences. 146, 163, 2016.
- ZHANG L., LIU J., WANG D., SCHALLER G.B., WU Y., HARRIS R.B., ZHANG K., LÜ Z. Distribution and population status of Przewalski's gazelle, Procapra przewalskii (Cetartiodactyla, Bovidae). Mammalia. 77 (1), 31, 2013.
- PING X., LI C., LI C., TANG S., FANG H., CUI S., CHEN J., WANG E., HE Y., CAI P., ZHANG Y., WU Y., JIANG Z. The distribution, population and conservation status of Przewalski's gazelle, Procapra przewalskii. Biodiversity Science. 26 (2), 177, 2018.
- GULCIN İ. Antioxidants and antioxidant methods: an updated overview. Archives of Toxicology. 94 (3), 651, 2020.
- ISMAEEL A., HOLMES M., PAPOUTSI E., PANTON L., KOUTAKIS P. Resistance Training, Antioxidant Status, and Antioxidant Supplementation. International Journal of Sport Nutrition and Exercise Metabolism. 29 (5), 539, 2019.
- 8. NISANSALA C., SEON GYEONG B., EUN HYUN P., HYUNG-JIN L., YEONG-SEON W., EUN-KYUNG K., SANG-IK P., SEUNG JAE L. Free radicals and their impact on health and antioxidant defenses: a review. Cell Death Discovery. 12 (23), 4008, 2025.
- CHIRINOS R., DELGADO-PARIONA J., AGUILAR-GALVEZ A., FIGUEROA-MERMA A., PACHECO-ÁVALOS A., CAMPOS D., PEDRESCHI R. Postharvest Storage Differentially Modulates the Enzymatic and Non-Enzymatic Antioxidant System of the Exocarp and Mesocarp of Hass Avocado: Implications for Disorders. Plants. 12 (23), 4008, 2023.
- YANG X.-J., DENG D.-M., LIU K.-H., YU F.-M. Response of enzymatic and non-enzymatic antioxidant defense systems of Polygonum hydropiper to Mn stress. Journal of Central South University. 23, 793, 2016.
- 11. ZHOU X., SHEN X. Environmental sulfur and molybdenum stress disrupts mineral homeostasis and induces physiological and molecular alterations in goats. Environmental Research. 282, 122033, 2025.
- 12. CHOWDHURY M.I., HASAN M., ISLAM M.S., SARWAR M.S., AMIN M.N., UDDIN S.M.N., RAHAMAN M.Z., BANIK S., HUSSAIN M.S., YOKOTA K., HASNAT A. Elevated serum MDA and depleted non-enzymatic antioxidants, macro-minerals and trace elements are associated with bipolar disorder. Journal of Trace Elements in Medicine and Biology. 3, 162, 2016.
- 13. WAHYUNI S. Role of natural antioxidant after exercise in reducing malondialdehyde (MDA) levels during pregnancy. JPMA. The Journal of the Pakistan Medical Association. 71 (2), 14, 2021.
- CAO S., LI X., YIN H., WANG J., LIU J. Dietary Puerarin Supplementation Improves Immune Response and Antioxidant Capacity of Sows. Antioxidants. 13 (3), 290, 2024.
- 15. JIN Y., HE Y., LIU L., TAO W., WANG G., SUN W., PEI X., XIAO Z., WANG H., WANG M. Effects of Supranutritional Selenium Nanoparticles on Immune and Antioxidant Capacity in Sprague-Dawley Rats. Biological Trace Element Research. 199 (12), 4666, 2021.
- SHEN X., HUO B., LI Y., SONG C., WU T., HE J. Response of the critically endangered Przewalski's gazelle (*Procapra przewalskii*) to se-lenium deprived environment. Journal of Proteomics. 241, 104218, 2021.

17. KIM E., JEON Y., KIM D.Y., LEE E., HYUN S.-H. Antioxidative effect of carboxyethylgermanium sesquioxide (Ge-132) on IVM of porcine oocytes and subsequent embryonic development after parthenogenetic activation and IVF. Theriogenology. 84 (2), 226, 2015.

- VISHTORSKAYA A.A., SAVERINA E.A., PECHENNIKOV V.M., KRYLOVA I.V., LALOV A.V., SYROESHKIN M.A., EGOROV M.P., JOUIKOV V.V. Assessing Ge-132 as an antioxidant in organic and watercontaining media. Journal of Organometallic Chemistry. 858, 8, 2018.
- AZUMI J., SHIMADA Y., TAKEDA T., ASO H., NAKAMURA T. The Organogermanium Compound 3-(Trihydroxygermyl) Propanoic Acid (THGP) Suppresses Inflammasome Activation Via Complexation with ATP. International Journal of Molecular Sciences. 23 (21), 13364, 2022.
- ZHOU X., SHEN X. Multi-omics insights into the mechanisms of muscle damage induced by molybdenum exposure in goats. Ecotoxicology and Environmental Safety. 299, 118373, 2025.
- 21. TEZUKA T., HIGASHINO A., AKIBA M., NAKAMURA T. Organogermanium (Ge-132) Suppresses Activities of Stress Enzymes Responsible for Active Oxygen Species in Monkey Liver Preparation. Advances in Enzyme Research. 5 (2), 13, 2017.
- KIELISZEK M., BŁAŻEJAK S., GIENTKA I., BZDUCHA-WRÓBEL A. Accumulation and metabolism of selenium by yeast cells. Applied Microbiology and Biotechnology. 99, 5373, 2015.
- LIM D.-H., LI M.-H., KIM E.-H., HAM S.-W. Synthesis of Novel Organogermanium Derivative Conjugated with Vitamin C and Study of its Antioxidant Effects. Bulletin of the Korean Chemical Society. 31 (7), 1839, 2010.
- 24. TZANETOU E., KASIOTIS M.K., THOMAIDIS S.N., ROSENBERG E. Determination of Trace Inorganic Germanium in bis-beta-Carboxyethylgermanium Sesquioxide by Headspace Solid-Phase Microextraction and Gas Chromatography Mass Spectrometry. Current Analytical Chemistry. 9 (2), 244, 2013.
- 25. MARET W. The Metals in the Biological Periodic System of the Elements: Concepts and Conjectures. International Journal of Molecular Sciences. 17 (1), 66, 2016.
- CUI B.-L., XIAO B., LI X.-Y., WANG Q., ZHANG Z.-H., ZHAN C., LI X.-D. Exploring the geomorphological processes of Qinghai Lake and surrounding lakes in the northeastern Tibetan Plateau, using Multitemporal Landsat Imagery (1973-2015). Global and Planetary Change. 152, 167, 2017.
- 27. WU W., ZHANG D., TIAN L., ZHANG M., ZHOU X. Features of artificial plant communities from the east sand region of the Qinghai Lake over the last 10 years. Acta Ecologica Sinica. 39 (6), 2109, 2019.
- 28. REDDEMAN R., GLÁVITS R., ENDRES J.R., MURBACH T.S., HIRKA G., VÉRTESI A., BÉRES E., SZAKONYINÉ I.P. A Toxicological Evaluation of Germanium Sesquioxide (Organic Germanium). Journal of Toxicology. 2020, 6275625, 2020.
- KUNIHIKO A., TAKASHI N., YASUHIRO S., TOMOYA T., JUNYA A., ANGELA C.S. Organogermanium: Potential beneficial effects on the cardiovascular system. Physiological Reports. 13 (3), e70234, 2025.
- KIM E., HYUN S.H. Carboxyethylgermanium sesquioxide confers antioxidative protection for porcine in vitro-fertilized embryos. Reproduction, Fertility and Development. 28 (2), 172, 2015.

- 31. DAVIS T.Z., STEGELMEIER B.L., GREEN B.T., WELCH K.D., HALL J.O. Evaluation of the respiratory elimination kinetics of selenate and Se-methylselenocysteine after oral administration in lambs. Research in Veterinary Science. **95** (3), 1163, **2013**.
- 32. ECHEVARRIA M.G., HENRY P.R., AMMERMAN C.B., RAO P.V. Effects of time and dietary selenium concentration as sodium selenite on tissue selenium uptake by sheep. Journal of Animal Science. 66 (9), 2299, 1988.
- 33. KOJOURI G.A., SADEGHIAN S., MOHEBBI A., MOKHBER DEZFOULI M.R. The effects of oral consumption of selenium nanoparticles on chemotactic and respiratory burst activities of neutrophils in comparison with sodium selenite in sheep. Biological Trace Element Research. 146, 160, 2011.
- 34. LACETERA N., BERNABUCCI U., RONCHI B., NARDONE A. The effects of injectable sodium selenite on immune function and milk production in Sardinian sheep receiving adequate dietary selenium. Veterinary Research. 30 (4), 363, 1999.
- 35. SADEGHIAN S., KOJOURI G.A., MOHEBBI A. Nanoparticles of selenium as species with stronger physiological effects in sheep in comparison with sodium selenite. Biological Trace Element Research. 146 (3), 302, 2011
- WANG S., MA L., YANG L., LONG X., GUAN C., ZHAO C., CHEN N. Quantifying desertification in the Qinghai Lake Basin. Frontiers in Environmental Science. 12, 2024.
- 37. ALIL M.M., NOAMAN E., KAMAL S., SOLIMAN S., ISMAIL D.A. Role of germanium L-cysteine alphatocopherol complex as stimulator of some antioxidant defense systems in gamma-irradiated rats. Acta Pharmaceutica. 57 (1), 1, 2007.
- 38. BURMISTROVA D.A., POMORTSEVA N.P., VORONINA Y.K., KISKIN M.A., DOLGUSHIN F.M., BERBEROVA N.T., EREMENKO I.L., PODDEL'SKY A.I., SMOLYANINOV I.V. Synthesis, Structure, Electrochemical Properties, and Antioxidant Activity of Organogermanium (IV) Catecholate Complexes. International Journal of Molecular Sciences. 25 (16), 9011, 2024.
- 39. CRUZ T.E., SITÓ-SILVA L., FILHO R.A.A., MARTINS A., MARQUI F.N., SOUZA D.G., BERTON T.I.U., FREITA-DELL'AQUA C.P., OBA E. Effects of antioxidant Bis-carboxyethyl germanium sesquioxide added to bovine semen cryopreservation medium on in vitro assessed morphofunctional sperm parameters. Reproduction in Domestic Animals. 59 (8), e14703, 2024.
- 40. CORINO C., ROSSI R. Antioxidants in Animal Nutrition. Antioxidants. 10 (12), 1877, 2021.
- 41. ROSSI L., DELL'ANNO M. Novel Antioxidants for Animal Nutrition. Antioxidants. 13 (4), 438, 2024.
- 42. KAISER S., WAGNER S., MOSCHNER C., FUNKE C., WICHE O. Accumulation of germanium (Ge) in plant tissues of grasses is not solely driven by its incorporation in phytoliths. Biogeochemistry. **148**, 49, **2020**.
- 43. OKOROAFOR P.U., OGUNKUNLE C.O., HEILMEIER H., WICHE O. Phytoaccumulation potential of nine plant species for selected nutrients, rare earth elements (REEs), germanium (Ge), and potentially toxic elements (PTEs) in soil. International Journal of Phytoremediation. 24 (12), 1310, 2022
- 44. WICHE O., SZÉKELY B., MOSCHNER C., HEILMEIER H. Germanium in the soil-plant system – a review. Environmental Science and Pollution Research. 25 (32), 31938, 2018.

- 45. MENCHIKOV L.G., POPOV A.V. Physiological Activity of Trace Element Germanium Including Anticancer Properties. Biomedicines. 11 (6), 1535, 2023.
- 46. MULDER P., VAN DEN HOEK A.M., KLEEMANN R. The CCR2 Inhibitor Propagermanium Attenuates Diet-Induced Insulin Resistance, Adipose Tissue Inflammation and Non-Alcoholic Steatohepatitis. PLoS One. 12 (1), e0169740, 2017.
- 47. ANTUS B., PASKA C., SIMON B., BARTA I. Monitoring Antioxidant Enzyme Activity during Exacerbations of Chronic Obstructive Pulmonary Disease. COPD-Journal of Chronic Obstructive Pulmonary Disease. 15 (5), 496, 2018.
- 48. PANKO N., KHADZHYNOVA I., SHUKREY S. AB0877 Dynamics of Parameters Changes of Antioxidant Defense and Lipid Peroxidation in Patients with Juvenile Idiopathic Arthritis. Annals of the Rheumatic Diseases. 75 (2), 1202, 2016.
- 49. BUSCH C.J., BINDER C.J. Malondialdehyde epitopes as mediators of sterile inflammation. Biochimica et Biophysica Acta (BBA) – Molecular and Cell Biology of Lipids. 1862 (4), 398, 2016.
- REN H., SHEN X. Multi-omics reveals the hepatic metabolic mechanism of neurological symptoms caused by selenium exposure in Przewalski's gazelle (Procapra przewalskii). Environmental Pollution. 375, 126341, 2025.
- 51. SIWULSKI M., BUDZYŃSKA S., RZYMSKI P., GĄSECKA M., NIEDZIELSKI P., KALAČ P., MLECZEK M. The effects of germanium and selenium on growth, metalloid accumulation and ergosterol content in mushrooms: experimental study in Pleurotus ostreatus and Ganoderma lucidum. European Food Research and Technology. 245 (9), 1799, 2019.
- 52. WU Z., CHEN X., YANG K., XIA T. Studies on the hydroxyl free radical-scavenging effect of combined selenium and germanium. Wei sheng yan jiu = Journal of Hygiene Research. 30 (4), 208, 2001.
- ŞLENCU B. G. Chapter 41 Selenium protection against cadmium and lead-induced oxidative stress. Academic Press, 419, 2021.
- 54. MORRIS G., GEVEZOVA M., SARAFIAN V., MAES M. Redox regulation of the immune response. Cellular & Molecular Immunology. 19 (10), 1079, 2022.
- EXNER R., WESSNER B., MANHART N., ROTH E. Therapeutic potential of glutathione. Wien Klin Wochenschr. 112 (14), 610, 2000.
- 56. SAFDARI-ROSTAMABAD M., HOSSEINI-VASHAN S.J., PERAI A.H., SARIR H. Nanoselenium

- Supplementation of Heat-Stressed Broilers: Effects on Performance, Carcass Characteristics, Blood Metabolites, Immune Response, Antioxidant Status, and Jejunal Morphology. Biological Trace Element Research. 178 (1), 105, 2017.
- 57. ASHRAFI H., SADEGHI A.A., CHAMANI M. Effect of Organic Selenium Supplementation on the Antioxidant Status, Immune Response, and the Relative Expression of IL-2 and IFN-γ Genes in Ewes During the Hot Season. Biological Trace Element Research. 202 (5), 2052, 2024.
- 58. ZHANG Y., ZHOU P., SHEN X. Effects of Se-Enriched Malt on the Immune and Antioxidant Function in the Se-Deprived Reclamation Merino Sheep in Southern Xinjiang. Biological Trace Element Research. 200 (8), 3621, 2022.
- 59. QIU H., GAO S., HOU L., LI A., ZHU L.-Q., DONG J., CHEN F. Selenium-enriched Bacillus subtilis Improves Growth Performance, Antioxidant Capacity, Immune Status, and Gut Health of Broiler Chickens. Biological Trace Element Research. 201 (12), 5756, 2023.
- 60. CHO J.M., CHAE J., JEONG S.R., MOON M.J., SHIN D.Y., LEE J.H. Immune activation of Bio-Germanium in a randomized, double-blind, placebo-controlled clinical trial with 130 human subjects: Therapeutic opportunities from new insights. PLoS One. 15 (10), e0240358, 2020.
- NIU R., YANG Q., DONG Y., HOU Y., LIU G. Selenium metabolism and regulation of immune cells in immuneassociated diseases. Journal of Cellular Physiology. 237 (9), 3449, 2022.
- 62. HUANG S., YANG W., HUANG G. Preparation and activities of selenium polysaccharide from plant such as Grifola frondosa. Carbohydrate Polymers. 242, 116409, 2020.
- 63. MURA M.C., LURIDIANA S., FARCI F., DI STEFANO M.V., DAGA C., PULINAS L., STARIČ J., CARCANGIU V. Melatonin treatment in winter and spring and reproductive recovery in Sarda breed sheep. Animal Reproduction Science. 185, 104, 2017.
- 64. AGUILAR-GARCÍA D., FERNÁNDEZ-SARMIENTO J.A., DEL MAR GRANADOS MACHUCA M., RODRÍGUEZ J.M., RASCÓN P.M., CALVO R.N., RUIZ Y.M., POVEDA J.M.C., CASTAÑEDA J.M., BERTOMEU R.C., DOMÍNGUEZ PÉREZ J.M. Histological and biochemical evaluation of plasma rich in growth factors treatment for grade II muscle injuries in sheep. BMC Veterinary Research. 18, (1), 400, 2022.
- JUDSON G.J., ELLIS N.J., KEMPE B.R., SHALLOW M. Long-acting selenium treatments for sheep. Australian Veterinary Journal. 68 (8), 263, 1991.